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Effect of *Thymus vulgaris* extract on systemic antibody responses against Influenza and Newcastle disease vaccine in broiler chickens

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ABSTRACT

Background & Aim: The recent study was conducted to investigate the effects of Thyme extract in drinking water on immune response of broiler chickens.

Experimental: A total of 245 day-old broiler chicks were purchased and 20 chicks were bled for determination maternal antibody and remaining chicks divided into 5 equal groups. Chickens of group A, B and C received 0.1%, 0.15% and 0.2% of Thyme extract in drinking water for all of the period of experiment, respectively. Chickens of group D did not receive Thyme extract but vaccinated against Newcastle and Influenza diseases. Chickens of group E were kept as control group and did not receive Thyme extract and Newcastle and Influenza diseases vaccines. Chickens of group A, B, C and D were vaccinated with live Newcastle vaccine (B1 strain) intraocularly and AI-ND killed vaccine (subtype H9N2) subcutaneously of neck back. Blood samples were collected before vaccination as well as on days 14, 28 and 35 after vaccination. Ten chickens of each group were bled randomly and antibody titer against Newcastle and Influenza vaccine virus was determined by HI test.

Results & Discussion: Results indicted that the extract had no effect on antibody response against Newcastle vaccine virus, but 14 days after vaccination, receiving of 0.2% of extract, significantly increased the specific antibody response against Influenza vaccine compared to all groups.

Industrial and practical recommendations: Herbs that are rich in flavonoids such as thyme (*Thymus vulgaris*) extend the activity of vitamin C, act as antioxidants and may therefore enhance the immune function.

1. Introduction

Nowadays, using antibiotics at sub-therapeutic levels has caused concerns about antibiotic residues in the animal productions which lead to the development of drug-resistant bacteria in animals and human. Thus, medical and public concerns focused on the complete removal of the antibiotics from animal feed in the European Union at the beginning of 2006 (Nollet, 2005; Wakeman, 2005; Cervantes, 2006). Therefore, poultry industry has been looking for the substances that could replace antibiotic growth promoters (AGP) in the feed (Bach Knudsen, 2001). Application of feed additives has two objectives: controlling pathogenic microorganisms and enhancing beneficial microorganisms in the digestive content of the gut (Vahdatpour et al., 2011). Recently some alternative components such as probiotics, prebiotics, organic acids and phytogenics fed additive have been introduced instead of antibiotics (Patterson and Burkhoder, 2003; Ricke, 2003). Recent bans and restrictions on the use of animal antibiotic growth promoters stimulated interest in bioactive secondary metabolites of plant source as alternative performance enhancers (Greathead, 2003). Medicinal plants and their essential oils have been used extensively in food products, perfumery, dental and oral products due to their different medicinal properties (Suppakul et al., 2003). Also in poultry production, it's very important to improve immunity so as to prevent infectious diseases. A variety of such factors as vaccination failure, infection by immune suppressive diseases, and abuse of antibiotics can induce immunodeficiency. Utilization of immunostimulants is a solution to improve the immunity of animals and to decrease their susceptibility to infectious disease (Liu, 1999). Herbs that are rich in flavonoids such as thyme (Thymus vulgaris) extend the activity of vitamin C, act as antioxidants and may therefore enhance the immune function (Manach et al., 1996; Cook and Samman, 1996). Thymus vulgaris is a medicinal herb in the Lamiaceae family, cultivated worldwide for culinary, cosmetic perennial and medical purposes. This species has special functions such as antispasmodic, expectorant, antiseptic, antimicrobial and antioxidant (Hertrampf, 2001; Abu-Darwish et al., 2009). Thymol (5-methyl-1-2-isopropyl phenol) and carvacrol (5isopropyl-2-methyl phenol) are the main phenolic components in Thymus vulgaris (Masada, 1976). Thyme (extract, oil, and the major components) have shown antibacterial activity against the Clostridium botulinum, Clostridium perfringens, Bacillus subtilis, Salmonella sonnei, Escherichia coli, Helicobacter pylori, Salmonella typhimurium, Bacillus cereus, Listeria monocytogenes, Campylobacter jejuni and Salmonella enteric reported in previous literatures (Nevas et al., 2004; Fan and Chen, 2001; Tabak et al., 1996; Juven et al., 1994; Ultee et al., 2000; Friedman et al., 2002; Thakare, 2004). Performance promoting effects of essential oil, extract, powder or principal components of thyme have been demonstrated in poultry (Al-Kassie, 2009; cross et al., 2007, Al-Mashhadani et al., 2011; Lee et al., 2003; Bolukbasi and Erhan., 2007; El-Ghousein and El-Beitawi., 2009). For example, supplementation of 200 ml Thyme extract per 1000 liter drinking waters in broiler chickens led to improvement of body weight gain and feed conversion ratio (Feisi and Bijanzad., 2010). Moreover, Rahimi et al. (2011) reported that dietary Thyme extract solution (0.1%) in water increased the performance and *lactic* acid counts whereas reduced the Escherichia coli numbers (p < 0.05). A variety of factors such as vaccination failure, infection by immune suppressive diseases, and abuse of antibiotics can induce immunodeficiency. Utilization of immunostimulants is one solution to improve the immunity of animals and to decrease their susceptibility to infectious disease (Liu, 1999). Herbs that are rich in flavonoids such as thyme (Thymus vulgaris) extend the activity of vitamin C, act as antioxidants and may therefore enhance the immune function (Manach et al., 1996; Cook and Samman, 1996). The current study was conducted to evaluate the potential of applying different levels of Thyme extract (Thymus vulgaris) on systemic antibody responses against Influenza and Newcastle disease vaccine in broiler chickens.

2. Materials and Methods

2.1. Chickens

A total of 245 day-old broiler chicks (Ross 308) were procured. All chickens were divided into 5 groups and raised under standard conditions. Chickens of group A, B, C and D were submitted to vaccinations against AIV and NDV.

2.2. Vaccinia

Hitchner B1 vaccine Cevac®, AI- ND killed vaccine (subtype H9N2).

2.3. Thyme extract

Thymus vulgaris aqueous extract was purchased commercially as solution (Dineh Iran - The Group of Pharmaceutical, Hygienic & Food industries (P.J.S)). The product contained 5 mg thymol per 5 ml of the solution.

2.4. Experimental design

Chickens of group A, B and C received 0.1%, 0.15% and 0.2% of Thyme extract respectively in drinking water for all of the period of experiment. Chickens of group D did not receive Thyme extract but vaccinated against Newcastle and Influenza diseases. Chickens of group E were kept as control group and did not receive Thyme extract and Newcastle and Influenza diseases vaccines. Chickens of group A, B, C and D were vaccinated with live Newcastle vaccine (B1 strain) intraocularly and AI-ND killed vaccine (subtype H9N2) subcutaneously of neck back.

2.5. Blood collection and serological tests

Blood samples were collected before vaccination as well as on days 14, 28 and 35 post vaccination. Ten chickens of each group were bled randomly and antibody titer against Newcastle and Influenza vaccine virus was determined by HI test. Blood samples were drained from the brachial vein and sera were separated, identified and frozen at -20° C until the serological tests were performed. Serum samples were analyzed by Hemagglutination inhibition test (HI) to detect antibodies against AIV and NDV (Alexander *et al.*, 1983).

2.6. Microplate heamagglutination inhibition (HI) assay

Beta procedure of micro-plate HI test was performed in U-bottomed 96-well microtiter plates with 1% chicken erythrocytes to determine the antibody level of the sera samples collected from the chicks of different groups. The test was conducted using constant 4HA unit AIV and ND virus and diluted.

2.7. Statistical analysis

The titers obtained by HI were submitted to analysis of variance using the Statistical Package for social Sciences (SPSS) version 18.0 program. One Way ANOVA LSD Test were performed to determine the significant differences in HI titers of chickens of each group after vaccination. Means were compared at a significance level of 5%.

3. Results and discussion

Results of Table 1, indicated that before vaccination, there was no significant difference between all groups. On the other hand, 14, 28 and 35 days after vaccination, there was no significant

difference between groups A, B, C as compared to the group D ,but groups A, B, C and D had significant difference as compared to the group E (P < 0.05). The results of present study showed that Thyme extract had no effect on antibody response against Newcastle vaccine virus.

The results of Table 2, indicated that before vaccination, there was not any significant difference between all groups, whereas 14 days after vaccination, there was significant difference between group C and all groups. Antibody titers in groups A, B, and C were higher than group D and E on the 28 and 35 days after vaccination, but there was not any significant difference between groups A, B, C as compared to group D. 14, 28 and 35 days after vaccination, group C has the highest antibody levels. The results of present study showed that14 days after vaccination, receiving of 0.2% of Thyme extract, significantly increased the specific antibody response to avian Influenza vaccine compared to all groups. In poultry industry, it's very important to improve immunity so as to prevent infectious diseases. A variety of such factors as vaccination failure, infection by immune suppressive diseases, and abuse of antibiotics can induce immunodeficiency. Utilization of immunostimulants is one solution to improve the immunity of animals and to decrease their susceptibility to infectious disease (Liu, 1999). Herbs that are rich in flavonoids such as thyme (Thymus vulgaris) extend the activity of vitamin C, act as antioxidants and may therefore enhance the immune function (Manach et al., 1996; Cook and Samman, 1996). In agreement with our results, Teymouri Zadeh et al. (2009) reported no significant difference in antibody responses to red blood cell and Newcastle disease viruses no between 0.1% thymus vulgaris extract received birds and control group (Teymouri Zadeh et al., 2009). None of the immune related parameters such as antibody titer against Newcastle, Influenza viruses and sheep red blood cell, heterophil to lymphocyte ratio and albumin to globulin ratio were differed significantly in broilers treated with 5 and 10 g/kg thyme powder while compared with control birds (Toghyani et al., 2010). Furthermore, Rahimi et al. (2011) reported that dietary thyme extract (0.1%)soluble in water increased performance and lactic acid counts and reduced E.coli numbers but did not affect immune system compared with control group (P<0.05) (Rahimi et al.,2011).

groups	Days after vaccination				
	0	14	28	35	
A (0.1%)	6.7±0.29	2.37±0.9 ^{C*}	4±0.8 ^E	3.04±0.2 ^E	
B (0.15%)	6.7±0.29	2.19±0.35 ^C	4.1±0.9 ^E	3.25 ± 0.53^{E}	
C (0.2%)	6.7±0.29	3.25±0.9 ABDE	4.3±0.9 ^E	$3.47{\pm}0.4^{\rm E}$	
D (vaccinated)	6.7±0.29	$2.05\pm0.7^{\circ}$ C	3. 5 ± 0.3^{E}	2.9 ± 0.37^{E}	
E (unvaccinated)	6.7±0.29	1.85±0.35 [°]	-	-	

Table 1. The effect of Thyme extract on HI antibody titer against Newcastle disease virus in broiler chicks.

Different superscripts in the same column represent significant difference (p <0.05). *Mean \pm standard deviation

groups	Days after vaccination				
	0	14	28	35	
A (0.1%)	6.05±0.8	4.1±0.87 ^{e*}	5.54±0.82 °	4.72±0.9 °	
B (0.15%)	6± 0.15	4.6±0.96 ^e	5.09±0.75 ^e	4.5±0.89e	
C (0.2%)	6.13±0.57	4.6±0.84 °	5.02±0.9 ^e	4.5±0.42 ^e	
D (vaccinated) E (unvaccinated)	5.9±0.33 6.1±0.19	$4\pm 0.85^{\text{ e}}$ $1.7\pm 0.82^{\text{abcd}}$	5.1±0.56 e _ abcd	4.63±0.48 ° _ abcd	

Table 2. Effect of probiotic on HI antibody titer against avian Influenza disease virus.

Different superscripts in the same column represent significant difference (p < 0.05). *Mean \pm standard deviation

The beneficial effects of thyme plant on bacterial and fungal activities and also potent antioxidant properties of major components of thyme essential oil such as thymol and carvacrol has been reported (Vincent 2002; Basilico and Basilico 1999). Considering the thyme characteristics, we anticipated that an increase in immune response of chicks would be observed. The lower results of thyme extract on immune system is probably related to the dose of additives, type of thyme, possess and preparation period and also vaccination program times and stimulator material that used in our study. Regarding this fact that a few reports are available on the impact of thyme or thyme component on bird immune response, more studies will be needed to investigate thyme extract immonomodulatory properties and principal components (Thymol and carvacol) on broiler health.

4. Conclusions

In conclusion, the results of the present study showed that supplementation of 0.1, 0.15 and 0.2% thyme extract in drinking water did not improve the systemic antibody responses against Newcastle disease vaccine in broiler chickens in the whole experimental period but 14 days after vaccination, receiving of 0.2% of Thyme extract significantly increased the specific antibody response against Influenza vaccine virus compared to all groups.

5. References

Abu-Darwish, M.S., Abu-Dieyeh, Z.H., Batarseh, M., Al-Tawaha, A.R.M and Al-Dalian, S.Y.A. 2009. Trace element contents and essential oil yields from wild thyme plant (*Thymus serpyllum*. L) growth at different natural variable environments, Jordan. *Journal of Food Agriculture and Environment*, 7: 920-924.

- Al-Kassie, G.A.M. 2009. Influence of two plant extracts derived from thyme and cinnamon on broiler performance. *Pakistan Veterinary Journal*, 29:169-173.
- Al-Mashhadani, E., Farah, H., Al-Jaff, K., Farhan, Y. M. and AL-Mashhadani, H.E. Effect of anise, thyme essential oils and their mixture (EOM) on broiler performance and some physiological traits. 2011. Egyptian Poultry Science, 31(2): 481-489.
- Bach Knudsen, K.E. 2001. Development of antibiotic resistance and options to replace antimicrobials in animal diets. *Proceedings of the Nutrition Society*, 60:291-299.
- Basilico, M.Z. and Basilico, J.C. 1999. Inhibitory effects of some spice essential oils on Aspergillus ochraceus NRRL 3174 growth and ochratoxin A production. *Letter Applied Microbiology*, 29: 238-24
- Bolukbasi, S., Erhan, M. 2007. Effect of dietary Thyme (*Thymus vulgaris*) on laying hens performance and *Escherichia coli* (*E. coli*) concentration in feces. *International journal of natural and engineering science*, 1(2): 55-58.
- Cervantes H. 2006. Banning antibiotic growth promoters: Learning from the European experience. *Poultry International*, 45: 14-15.
- Cook, N. C. and Samman, S. 1996. Flavonoidschemistry, metabolism, cardio protective effects, and dietary sources. *Journal of Nutrition and Biochemistry*, 7: 66-76.
- Cross, D.E., McDevitt, R.M., Hillman, K. 2007. Acamovic. The effect of herbs and their associated essential oils on performance, dietary digestibility and gut microflora in chickens from 7 to 28 days of age. *British Poultry Science*, 48: 496–506.
- El-Ghousein, S.S and N.A. Al-Beitawi. 2009. The effect of feeding of crushed Thyme (*Thymus vulgaris*) on growth, blood constituents, and gastrointestinal tract and carcass characteristics of broiler chickens. *Journal of Poultry Science*, 46: 100-104
- Fan, M and J. Chen. 2001. Studies on antimicrobial activity of extracts from thyme. *Wei Sheng Wu Xue Bao*, 41: 499-504.

- Feizi, A., Bijanzad, P. 2010. Evaluating the Effects of *Thymus vulgaris* extract on Growth Performance Parameters in Broiler Chicken. *Journal of Veterinary Medicine*, 12: 39-45.
- Friedman M., Henika, P. R. and Mandrell, R. E. 2002. Bactericidal activities of plant essential oils and some of their isolated constituents against *Campylobacter jejuni*, *Escherichia coli*, *Listeria monocytogenes*, and *Salmonella enterica*. *Journal* of Food Protection, 65:1545-1560.
- Greathead, H. 2003. Plants and plant extracts for improving animal productivity. *Proceedings of the Nutrition Society*, 62:279-290.
- Hertrampf, J.W. 2001. Alternative antibacterial, performance promoters. *Poultry International*, 40:50-52.
- Juven, B. J., Kanner, J., Schved, F. and Weisslowicz, H. 1994. Factors that interact with the antibacterial action of thyme essential oil and its active constituents. *Journal of Applied Bacteriology*. 76:626-631.
- Lee, K. W., Everts, H., Kappert, H. J., Frehner, M., Losa, R. and Beynen, A. C. 2003. Effects of dietary essential oil components on growth performance, digestive enzymes and lipid metabolism in female broiler chickens. *British Poultry Science*, 44: 450-457.
- Liu, X. Y. 1999. Stress and Immunity. In: "Poultry Immunology", (Ed.): Yin, T. B.China Agriculture Press, Beijing, China, 230–252.
- Manach, F., Regerat, F. and Texier, O. 1996. Bioavailability, Metabolism and Physiological Impact of 4-oxo-flavonoids. *Nutrition Research*, 16: 517-44.
- Masada, Y. 1976. Analysis of oils by gas chromatography and mass spectrometry. Johan Wiley and Sons.
- Nevas, M., Korhonen, A. R., Lindstrom, M., Turkki, P. and Korkeala, H. 2004. Antibacterial efficiency of finnish spice essential oils against pathogenic and spoilage bacteria. *Journal of Food Protection*, 67: 199-202.
- Nollet, L. 2005. AGP alternatives-part I. EU close to a future without antibiotic growth promoters. *World Poultry*, 21: 14-15.
- Patterson, J.A., Burkholder, K.M. 2003. Application of prebiotics and probiotics in poultry production. *Poultry Science*, 82: 627–631.

- Rahimi, S., Teymouri Zadeh, Z., Karimi Torshizi, M. A., Omidbaigi, R. and Rokni, H. 2011. Effect of the Three Herbal Extracts on Growth Performance, Immune System, Blood Factors and Intestinal Selected Bacterial Population in Broiler Chickens. *Journal of Agricultural Science and Technology*, 13: 527-539
- Ricke, S.C. 2003. Perspectives on the use of organic acids and short chain fatty acids as antimicrobials. *Poultry Science*, 82: 632–639.
- Suppakul, P., Miltz, J., Sonneveld, K. and Bigger, S. W. 2003. Antimicrobial properties of basil and its possible application in food packaging. *Journal of Agricultural and Food Chemistry*, 51: 3197- 3207.
- Tabak, M., Armon, R., Potasman, I. and Neeman, I. 1996. *In vitro* inhibition of Helicobacter pylori by extracts of thyme. *Journal of Applied Bacteriology*, 80: 667-672.
- Teymouri Zadeh, Z., Rahimi, S.H., Karimi Torshizi, M.A., Omidbaigi, R. 2009. *Iranian Journal of Medicinal and Aromatic Plants*, Vol 25, No. 1.
- Thakare, M. 2004. Pharmacological screening of some medicinal plants as antimicrobial and feed additive, M.S Thesis, Virginia Polytechnic Institute and State University, Blacksburg, Virginia USA.

- Toghyani, M., Tohidi M,Gheisari, A., Tabeidian, S. 2010. Performance, immunity, serum biochemical and hematological parameters in broiler chicks fed dietary thyme as alternative for an antibiotic growth promoter. *African Journal of Biotechnology*, 9(40): 6819-6825.
- Ultee, A., Slump. R. A., Steging, G. and Smid, E. J. 2000. Antimicrobial activity of carvacrol toward *Bacillus cereus* on rice. *Journal of Food Protection*, 63: 620-624.
- Vahdatpour, T., Nikpiran, H., Babazadeh, D., Vahdatpourand, S. & Jafargholipour, M.A. 2011. Effects of Protexin ®, Fermacto ® and combination of them on blood enzymes and performance of Japanese quails (Coturnix Japonica). Annals of Biological Researches, 2: 283-291.
- Wakeman, G.W. 2005. AGP alternatives- Part II. Dietary strategies to influence bacterial microflora. *World Poultry*, 21: 28-29.
- Vincent, H.V. 2002. Carvacrol and thymol reduce swine waste odour and pathogens stability of oils. *Current Microbiology*, 44: 38-43.