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Insecticidal activity of some medicinal plant essential oils combined with Proteus® against greenhouse whitefly, *Trialeurodes vaporariorum* (Westwood) under greenhouse conditions

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Abstract

Trialeurodes vaporariorum (Westwood) is one of the most important pests of vegetable and ornamental crops in the world. Generally, control of T. vaporariorum is based on the application of insecticides, but they are resistant to many synthetic chemicals. There is a necessity to evaluate non-chemical and low-risk ways against the pest. Accordingly, medicinal plants are promising because they are safe to human, eco-friendly and biodegradable components. However, chemical insecticides are an inevitable component of agricultural products. Hence, in the present study, we examined the synergistic/antagonistic interactions between Agastache foeniculum (Pursh) Kuntze, Cuminum cyminum L., Ferulago angulata (Schlecht) Boiss, Foeniculum vulgare Mill., Origanum vulgare Mill., Satureja hortensis L., Trachyspermum ammi I. Sprague and Ziziphora tenuior L. essential oils with Proteus® against T. vaporariorum adults and eggs. Probit analysis of essential oils showed that O. vulgare and T. ammi were the most effective essential oils against T. vaporariorum adults that both exhibited LC_{50} values equivalent to 0.44 μ L/L air. F. vulgare essential oil had the highest mortality against the eggs of T. vaporariorum (LC₅₀= 30.60 μ L/L air). All essential oils synergized efficacy of Proteus® against adults of T. vaporariorum. The inclusion of Z. tenuior essential oil with Proteus® led to antagonistic interaction against T. vaporariorum eggs.

Key words: Trialeurodes vaporariorum, Medicinal plant, Essential oil, Proteus®, Synergistic effect

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Introduction

The greenhouse whitefly, Trialeurodes vaporariorum (Westwood) (Homoptera: Aleyrodidae) is an important and significant pest in greenhouses in the most parts of the world (Wang et al., 2003; Malais & Ravensberg, 2004). T. vaporariorum decreases the yield and quality of plants due to direct feeding and transmission of many plant viruses (Skaliac *et al.*, 2010). The pest sucks cell sap of the leaves and produces honeydews that adversely impacts plant photosynthesis (Ali et al., 2005). T. vaporariorum has a very vast host range consist of more than 300 types of 82 plant families that the most important ones are Cucurbitaceae, Solanaceae, Fabaceae and among the greenhouse vegetables, the most important ones are tomatoes, cucumbers and some ornamentals like chrysanthemum, poinsettia etc. It is well documented that chemical control is an effective method to control the pests (Pavela, 2009) and chemical insecticides are still important management tools of the greenhouse whitefly (Wang et al., 2003). Proteus® (the new commercial mixture of thiacloprid and deltamethrin) is a powerful and wide-spectrum insecticide (Sekeroglu et al., 2011). It is a contact and systemic insecticide. In recent decades, many new insecticides with different mode of actions have been developed for the management of whiteflies. Pesticides can cause serious risks to human health and the environment, so natural components such as plant materials could be useful, since they are easy to extract, safe and do not persist in water and soil (Isman, 2000). We selected Proteus® because it is a powerful and wide-spectrum insecticide with short pre-harvest interval and low effect on natural enemies (Elbert *et al.*, 2001). This insect has also developed immunity to many conventional insecticides. Several essential oils have been found to be effective against all insect stages, including eggs, nymphs and adults of T. vaporariorum without adverse effect on beneficial insects. Therefore, we examined some medicinal plant essential oils (Agastache foeniculum, Cuminum cyminum, Ferulago angulata, Foeniculum yulgare, Oreganum vulgare, Satureja hortensis, Trachysperum ammi and Ziziphora tenuior) against the pest. We also examined the antagonistic/synergistic interactions between mentioned essential oils combined with Proteus® against T. vaporariorum eggs and adults.

Materials and Methods

Insect rearing

Adults of *T. vaporariorum* were collected by an aspirator from the greenhouse of Department of horticulture at Urmia University, Urmia, Iran. Trials were carried out in the years 2015 and 2016 in greenhouse of Department of Plant Protection, Faculty of Agriculture, Urmia University, Urmia, Iran. The colony was mass reared on tomato (an important host of greenhouse whitefly). The collected adults were transferred on the tomatoes with 4-5 leaves. Experiments were conducted after three generations of *T. vaporariorum* rearing. Trials were carried out under greenhouse conditions at 27 ± 2 °C, $65 \pm 5\%$ RH and a photoperiod of 16 L: 8 D. In order to access a cohort, following Muniz and Nombela technique (2001), small cages with a few changes were used.

Insecticide

The insecticide used was Proteus[®] (110 OD, 100 g/l thiacloprid and 10 g/l deltamethrin, Bayer CropScience, New Zealand) that is a broad spectrum insecticide.

Essential oils

Essential oils were extracted from seeds of *T. ammi*, *C. cyminum* and *F. vulgare* and leaves of *S. hortensis*, *Z. tenuior*, *F. angulata* and *O. vulgare* and flowers of *A. foeniculum*. They were subjected to hydro distillation using a modified Clevenger-type apparatus.

Essential oils

Bioassays

Bioassay trials were carried out following Rahman and Schimdt technique (1999) and Negahban *et al.* (2007). The concentration ranges of essential oils were determined by preliminary dose setting experiments. In this respect, filter papers were placed in the cap of leaf cages and impregnated with the required concentration of essential oils, then they covered with parafilm. One tomato leaf was placed in each leaf cage, then 20 one to two-days-old adults of *T. vaporariorum* were transferred into the leaf cages by an aspirator. They were exposed to vapors of essential oils of *A. foeniculum, C. cyminum, F. angulata, F. vulgare, O. vulgare, S. hortensis, T. ammi* and *Z. tenuior* (before the cages were capped). These experiments were carried out under greenhouse conditions at 27 ± 2 °C, $65\pm5\%$ RH and a photoperiod of 16 L: 8 D. Each concentration had three replications and each experiment was replicated three times. Mortality was counted after 24 h. To examine the sensitivity of egg hatch of the pest to the essential oils, one tomato leaf containing 20 one-day-old eggs of *T. vaporariorum* was placed in leaf cages. They were exposed to vapors of essential oils of *A. foeniculum, C. cyminum, F. angulata, F. vulgare, O. vulgare, S. hortensis, T. ammi* and *Z. tenuior*. These trials were carried out similar to adults. Mortality was recorded after 10 days. Each experiment was replicated three times.

Insecticide

Trials were carried out following Prabhaker *et al.* (1988) method with a slight modification. Tomato leaves were dipped in determining concentrations of Proteus® that were determined by preliminary dose setting experiments. For controls, the leaves were dipped in distilled water. When the water evaporated and leaves were dried, 20 one to two-days-old adults of *T. vaporariorum* were transferred into leaf cages by an aspirator. These experiments were carried out under greenhouse conditions at 27 ± 2 °C, $65 \pm 5\%$ RH and a photoperiod of 16L : 8D. Mortality was counted after 24 h. Each experiment was replicated three times. In order to survey egg hatch of greenhouse whitefly, one tomato leaf containing 20 one-day-old eggs of *T. vaporariorum* was dipped in determining concentrations of Proteus® that were determined by preliminary dose setting experiments. After water evaporating and the leaves were dried, they were placed in leaf cages. These trials were carried out similar to adults. Mortality was recorded after 10 days. Each experiment was replicated three times.

Plant essential oils combined with Proteus®

In order to assay whether there was an antagonistic or synergistic interaction between essential oils combined with Proteus®, sub-lethal concentrations (LC₂₅) of essential oils were combined with LC₂₅ of Proteus®. Therefore, one tomato leaf was dipped in sub-lethal concentration of Proteus®. When the water evaporated and the leaf was dried, the leaf was placed in leaf cage and 20 one-day-old adults were transferred into leaf cages. Then LC₂₅ of essential oils were immersed with filter paper. Finally, the leaf cage was covered with parafilm. This trial was carried out for all essential oils separately. Mortality was recorded after 24 h. Each experiment was replicated three times. To examine whether the mentioned essential oils synergize performance of Proteus® against the eggs of *T. vaporariorum*, one tomato leaf containing 20 one-day-old eggs was dipped in sub-lethal concentration of Proteus®. When the water evaporated and the leaf was dried, the leaf was placed in leaf cage. Then filter paper was immersed in LC₂₅ of essential oils. Finally, the leaf cage was covered with Parafilm. This trial oils. Finally, the leaf cage was covered with Parafilm. This trial was carried out for all essential oils was dipped in sub-lethal concentration of Proteus®. When the water evaporated and the leaf was dried, the leaf was placed in leaf cage. Then filter paper was immersed in LC₂₅ of essential oils. Finally, the leaf cage was covered with Parafilm. This trial was carried out for all essential oils separately. Mortality was recorded after 10 days. Each experiment was replicated three times.

Data Analysis

In order to determine LC_{50} and LC_{25} values, the data were analyzed using the probit procedures with SPSS for Windows® release 16. The percentage data were transformed into $\arcsin\sqrt{x}$ before statistical analysis. To determine synergistic/antagonistic interactions, experiments were conducted following Gisi (1991). The relationship between data was assayed by analysis of variance (ANOVA) and correlation analysis. The means were separated by using the Tukey test.

Results

According to table 1, *T. ammi* and *O. vulgare* showed the highest efficacy against *T. vaporariorum* adults, which both exhibited LC_{50} value equivalent to 0.44 μ L/L air.

	Table 1- Pro	bit analysis of the toxi	city of some essential oils t	to T. vaporarium adults	
Compound	χ^2	Slope ± S.E	LC ₂₅ (µL/L air)	LC ₅₀ (µL/L air)	LC ₉₀ (µL/L air)
A. foeniculum	1.65	3.28 ± 0.85	1.76	2.12	2.96
C. cyminum	1.70	2.61 ± 0.70	2.80	3.52	5.08
F. angulata	1.93	2.26 ± 0.66	0.72	0.96	1.68
F. vulgare	1.77	2.00 ± 0.66	1.68	2.16	3.56
O. vulgare	2.68	1.78 ± 0.25	0.16	0.44	2.32
S. hortensis	2.78	2.97 ± 0.61	3.00	3.28	3.80
T. ammi	2.33	2.55 ± 0.51	0.28	0.44	1.04
Z. tenuior	3.02	3.22 ± 0.87	3.12	3.99	4.35

جدول ۱– تجزیه پروبیت سمیت برخی از اسانسهای گیاهی روی حشرات کامل سفیدبالک گلخانه

Probit analysis of Proteus® demonstrated that it was toxic for adults (LC_{50} = 1.11 ppm) (Table 2).

	Table 2- Prol	oit analysis of the toxicit	y of Proteus® to T. vap	porarium adults	
Compound	χ^2	Slope ± S.E	LC ₂₅ (ppm)	LC ₅₀ (ppm)	LC ₉₀ (ppm)
Proteus®	1.87	2.73 ± 0.51	0.73	1.11	2.44

Results presented in table 3 show that fennel had the highest toxicity on hatch eggs of greenhouse whitefly which exhibited LC_{50} value equivalent to 30.60 μ L/L air.

	Table 3- Probit a	analysis of the toxicity of	some essential oils to	T. vaporarium eggs	
Compound	χ^2	Slope ± S.E	LC ₂₅ (µL/L	LC ₅₀ (µL/L	LC ₉₀ (µL/L
			air)	air)	air)
A. foeniculum	1.76	5.21 ± 0.68	36.80	50.80	68.40
C. cyminum	2.57	2.08 ± 0.65	42.00	54.00	88
F. angulata	1.34	3.89 ± 0.71	39.90	51.04	93.24
F. vulgare	1.53	1.21 ± 0.20	2.40	3.60	6.44
O. vulgare	3.14	2.32 ± 0.31	34.40	50	100.48
S. hortensis	1.32	1.80 ± 0.09	70.40	75.6	86.36
T. ammi	0.97	2.17 ± 0.40	23.60	38.80	98
Z. tenuior	1.49	1.90 ± 0.11	67.60	74.40	89.76

جدول ۳- تجزیه پروبیت سمیت برخی از اسانس های گیاهی روی تخم سفیدبالک گلخانه

Albeit, essential oil of ammi was toxic for eggs of the pest (LC_{50} value = 38.80 μ L/L air). Proteus® exhibited LC_{50} value equivalent to 31.08 ppm against the eggs of *T. vaporariorum* (Table 4).

نخم سفيدبالک گلخانه	سمیت پروتئوس روی	جدول ۴- تجزیه پروبیت
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	Table 4- Pro	bit analysis of the toxici	ty of Proteus® to T. va	<i>porarium</i> eggs	
Compound	χ^2	Slope ± S.E	LC ₂₅ (ppm)	LC ₅₀ (ppm)	LC ₉₀ (ppm)
Proteus®	3.80	2.82 ± 0.35	20.01	31.08	88.39

The inclusion of A. foeniculum, C. cyminum, F. angulata, F. vulgare, O. vulgare, S. hortensis, T. ammi and Z. tenuior essential oils with Proteus® led to synergistic interaction against T. vaporariorum adults (Table 5).

Table 5- Synergistic	- Synergistic interactions between some essential oils and Proteus® against T. vaporarium adults			
P + essential oils	%mortality \pm S. E.		interaction	
	Expected	Observed		
P + EO of A. foeniculum	58.01 ± 1.80	73 ± 2.82	synergism	
P + EO of C. cyminum	57.69 ± 2.12	81 ± 3.03	synergism	
P + EO of F. angulata	76.62 ± 2.25	83 ± 2.98	synergism	
P + EO of F. vulgare	76.73 ± 2.56	80 ± 3.03	synergism	
P+ EO of S. hortensis	76.72 ± 2.00	80 ± 3.00	synergism	
P+ EO of T. ammi	56.69 ± 2.05	70 ± 2.03	synergism	
P+ EO of Z. tenuior	76.00 ± 2.45	80 ± 2.99	synergism	

جدول ۵– اثر سینرژیستی برخی از اسانس،های گیاهی و پروتئوس روی حشرات کامل سفیدبالک گلخانه

In the case of *T. vaporariorum* eggs exposed to a mixture of all essential oils (except for *Z. tenuior*) and Proteus[®], the synergistic interaction was recorded (Table 6). The inclusion of *Z. tenuior* essential oil with Proteus[®] led to antagonistic interaction against *T. vaporariorum* eggs (Table 6).

P + essential oils	$\%$ mortality \pm S. E.		interaction
	Expected	Observed	
P + EO of A. foeniculum	70.20 ± 2.01	75 ± 1.90	synergism
P + EO of C. cyminum	56.69 ± 2.05	80 ± 4.03	synergism
P + EO of F. angulata	56.69 ± 2.05	80 ± 4.03	synergism
P + EO of F. vulgare	70.00 ± 2.95	83 ± 3.13	synergism
P+ EO of S. hortensis	56.72 ± 1.35	70.69 ± 2.34	synergism
P+ EO of T. ammi	76.69 ± 2.35	86.69 ± 4.00	synergism
P+ EO of Z. tenuior	45.53 ± 1.25	40 ± 0.78	antagonism

جدول ۶– اثر سینرژیستی برخی از اسانسهای گیاهی و پروتئوس روی تخم سفیدبالک گلخانه

Discussion

T. vaporariorum adults were susceptible to essential oils combined with Proteus®. In the present study, *T. ammi* and *O. vulgare* essential oils exhibited high insecticidal activity against adults of *T. vaporariorum*. Proteus® had a high efficacy against *T. vaporariorum* adults. Essential oils are valuable secondary metabolites obtained through steam distillation of medicinal plants and herbs (Yatagai, 1997). Most of them have no persistence and are non-toxic to human and environment (Hjorther *et al.*, 1997). Khorrami and Soleymanzade (2016) investigated efficacy of some medicinal plant essential oils, extracts and Powders against adults of *C. allosobruchus maculatus* F. (Coleoptera: Bruchidae) and they found that essential oils of *T. ammi* and *F. vulgare* were effective on the pest (LC_{50} = 0.64 and 0.72 µL/L air, respectively). Regnault–Roger and Hamraoui (1993) evaluated the influence of aromatic essential oils of *O. vulgare* (L.), *Thymus vulgaris* (L.) and *Eucalyptus globules* Labill. on *Acanthoscelides obtectus* (Say) and they reported that *O. vulgare* resulted in more mortality of pest.

Our results showed that egg hatching was significantly reduced by *F. vulgare* essential oil. We demonstrated that in the case of *T. vaporariorum* eggs exposed to a mixture of Proteus® and essential oils, the synergistic interaction was recorded (except for *Z. tenuior*). Sampson *et al.* (2005) tested insecticidal activity of 23 essential oils on *Lipaphis pseudobrassicae* (Davis) and they showed that the most effective essential oils were *Rosmarinus officinalis* (L.), *S. hortensis, Mentha piperita* (L.) and *F. vulgare* (Miller), which elicited high fumigant toxicity. Choi *et al.* (2003) were tested 53 plant essential oils against eggs, nymphs and adults of *T. vaporariorum* and they reported that Pennyroyal and peppermint oils gave 89 and 83% mortality. They showed that caraway seed, lemongrass, oregano, and spearmint oils were the most effective compounds against eggs of the pest. Górski (2004) assayed the effectiveness of natural essential oils in the monitoring of greenhouse whitefly (*T. vaporariorum*). He

noted that greenhouse whitefly (T. vaporariorum) reacted most intensively to Ocimum basilicum oil. The addition of this oil on yellow sticky traps caused a significant increase in trapped insects in comparison with the control (with no aromatic substance). Dehghani and Ahmadi (2013) demonstrated that the highest anti-oviposition effect after 3 days caused by essential oil of A. millefolium (67.77%), while the lowest anti-oviposition index was for aqueous extract of C. cyminum (48.78%). Also the aqueous extract of C. cyminum showed the highest repellent effect on the greenhouse whitefly after 3 d (66.11%). Thielert and Hungenberg (2007) investigated the biological effect of O-D (oil dispersion) formulation insecticides such as Proteus®, Confidor® and biscayain and also EC, SC and SE formulations against important sucking-biting insects like peach and cotton aphid. Their results showed that all the insecticides with O-D formulations were more effective than the others and had a better knockdown effect and durability. Generally, based on our results, interaction of essential oils with Proteus® has insecticidal effectiveness against T. vaporariorum adults and eggs. The selection of Proteus® and medicinal plant essential oils was based on low side effects on natural enemies, bio-degrability, lower-risk to beneficial organisms, human and environment. Hence, A. foeniculum, C. cyminum, F. angulata, F. vulgare, O. vulgare, S. hortensis, T. ammi and Z. tenuior essential oils combined with Proteus® can be applied to control the pest infestations as a part of an integrated pest management strategy. Although, the combination of mentioned essential oils combined with Proteus® is very effective for controlling the pest, but additional research is needed.

References

- Ali, M. A., Rehman, R., Tatla, Y. H. and Ali, Z. 2005. Evaluation of different insecticides for the control of whitefly on cotton crop in karor district Layyah. Pakistan Entomologyist, 27(1): 5-8.
- Choi, W. I., Lee, E. H., Choi, B. R., Park, H. M. and Ahn, Y. J. 2003. Toxicity of Plant Essential Oils to *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae). Journal of Economic Entomology, 96 (5): 1479-1484.
- **Dehghani, M. and Ahmadi, K. 2013.** Anti-oviposition and repellence activities of essential oils and aqueous extracts from five aromatic plants against greenhouse whitefly *Trialeurodes vaporariorum* (Westwood) (Homoptera: Aleyrodidae). Bulgarian Journal of Agricultural Science, 19 (4): 691-696.
- Elbert, A., Ebbinghause-Kint, S. U., Erdelen, C., Nauen, R. and Schnorbach, H. J. 2001. The biological profile of thiacloprid, a new chloronicotinyl insecticide. Pslanzenschutz. Nach Bayer, 54(2): 185-208.
- Gisi, U. 1991. Synergism between fungicides for control of *Phytophthora*. In: *Phytophthora*. J. I. Lucas, R. C. Shattock, D. S. Shaw, and L. R. Cooke, eds. Cambridge University Press, Cambridge, pp: 361-372.
- **Górski, R. 2004.** Effectiveness of natural essential oils in the monitoring of greenhouse whitefly (*Trialeurodes vaporariorum* Westwood). Folia Horticulture, 16(1): 183-187.
- Hjorther, A. B., Christophersen, C., Hausen, B. M. and Menne, T. 1997. Occupational allergic contact dermatitis from carnosol, a naturally occurring compound present in rosemary. Contact Dermatitis, 37: 99-100.
- Isman, M. B. 2000. Plant essential oils for pest and disease management. Crop Protection, 19: 603-608.
- Khorrami, F. and Soleymanzade, A. 2016. Efficacy of Some Medicinal Plant Essential Oils, Extracts and Powders Combined with Diatomaceous Earth on Cowpea Weevil, *Callosobruchus maculatus* F. (Coleoptera: Bruchidae). International Journal of Science and Research, 5 (12): 1118-1123.
- Malais, M. H. and Ravensberg, W. j. 2004. Knowing and recognizing: the biology of glasshouse pests and their natural enemies. The Netherlands: Koppert B. V. Berkel en Rodenrijs, 288 pp.

- **Muniz, M. and Nombela, G. 2001.** Differential variation in development of the B- and Qbiotypes of *Bemisia tabaci* (Homoptera: Aleyrodidae) on sweet pepper at constant temperatures. Environmental Entomology, 30: 720-727.
- Negahban, M., Moharramipour, S. and Sefidkon, F. 2007. Fumigant toxicity of essential oil from *Artemisia sieberi* Besser against three insects. Journal of Stored Products Research, 43: 123-128.
- Pavela, R. 2009. Larvicidal effects of some Euro-Asiatic plants against *Culex quinquefasciatus* Say larvae (Diptera: Culicidae). Parasitology Research, 105: 887-892.
- Prabhaker, N., Coudirect, D. L. and Toscano, N. C. 1988. Effect of synergists on organophosphate and permethrin resistance in sweetpotato whitefly (Homoptera: Aleurodidae). Journal of Economic Entomology, 81(1): 34-39.
- Rahman, M. M. and Schimdt, G. H. 1999. Effect of Acorus calamus (L.) (Araceae) essential oil vapours from various origins on *Callosobruchus Phaseoli* (Gyllenhal) (Coleoptera; Bruchidae). Journal of Stored Products Research, 35: 285-295.
- **Regnault-Roger, C. and Hamraoui, A. 1993.** Influence d'huiles essentielles sur *Acanthoscelides obtectus* Say, bruche du haricot [Efficiency of plants from south of France used as traditional protectants of *Phaseolus vulgaris* L. against its bruchid *Acanthoscelides obtectus* (Say)]. Acta Bot Gallica [Journal of Stored Products Research], 29: 259-264.
- Sampson, B. J., Tabanca, N., Kirimer, N., Demirci, B., Baser, K. H., Khan, I. A., Spiers, J. M. and Wedge, D. E. 2005. Insecticidal activity of 23 essential oils and their major compounds against adult Lipaphis pseudobrassicae (Davis) (Aphididae: Homoptera). Pest Management Science, 61: 1122–1128.
- Sekeroglu, V., Sekeroglu, Z. and Kefelioglu, H. 2011. Cytogenetic effects of commercial formulations of Deltamethrin and or Thiacloprid on Wistar rat bone marrow cells. Environmental Toxicology DOI: 10.1002/tox.20746. pp: 524-531.
- Skaljac, M., Zanic, K., Goreta Ban, S., Kontsedalov, S. and Ghanim, M. 2010. Co-infection and localization of secondary symbionts in two whitefly species. BMC Microbiology, 10: 142-157.
- SPSS, 2007. Spss 16 for windows user, s guide release. Chicago Spss Inc.
- Thielert, W. and Hungengberg, H. 2007. Bioligical efficiacy of Q-TEQ insecticides under controlled conditions. Pflanzenschutz-Nachrichten Bayer, 60(1): 43- 58.
- Wang, K. Y., Kong, X. B., Jiang, X. Y., Yi, M. Q. and Liu, T. X. 2003. Susceptibility of immature and adult stages of *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae) to selected insecticides. Journal of Applied Entomology, 127: 527-533.
- Yatagai, M. 1997. Miticidal activities of tree terpenes. Current Topics in Phytochemistry, 1: 85-97.

فصلنامه تخصصي تحقيقات حشرهشناسي

(علمي- پژوهشي)

جلد ۹، شماره ۳، سال ۱۳۹۶، (۲۲–۲۰)

اثر حشره کشی تعدادی از گیاهان دارویی در ترکیب با پروتئوس روی سفیدبالک گلخانه تحت شرایط گلخانهای

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چکیدہ

سفیدبالک گلخانه (Westwood) و گیاهان (تعنی در معمترین آفات سبزیجات و گیاهان زینتی در جهان است. به طور کلی کنترل این آفت بر پایه استفاده از حشره کش ها است، اما آن ها به بسیاری از مواد شیمیایی مصنوعی مقاوم شده اند. ارزیابی روش های غیر شیمیایی و کم خطر برای سفیدبالک گلخانه ضروری به نظر می رسد. بر این اساس، گیاهان دارویی به دلیل بی خطر بودن برای انسان، سازگار بودن با محیط زیست و خاصیت تجزیه پذیری، ترکیباتی امیدوار کننده می باشند. با این حال، ترکیبات شیمیایی اجزای اجتناب ناپذیر در تولیدات کشاورزی هستند. از این رو در تحقیق حاضر ما اثر سینرژیستی/آنتاگونیستی اسانس گیاهان گل مکزیکی Agastache foeniculum در می و سبز Cuminum چویل Agastache foeniculum vulgare گیاهان گل مکزیکی Agastache foeniculum در مرزه معتند. از این رو در تحقیق ناصر ما اثر سینرژیستی/آنتاگونیستی اسانس گیاهان گل مکزیکی Foeniculum vulgare در مرزه و Satureja مونی ما شر مینرژیستی/آنتاگونیستی اسانس گیاهان گل مکزیکی مواند می مرزه مورک می دره مرزه مرزه می می اشد. کامل سفیدبالک گلخانه مورد بررسی قرار دادیم. تجزیه پروبیت اسانس ها نشان داد که مرزنجوش و زنیان روی حشره کامل و کامل سفیدبالک میزه می در این روی تخم می در این در اسانس برابر با ۲۰۴٬۰۰ میکرولیتر بر لیتر هوا بود. اسانس رازیانه بیشترین میزان مرگ و میر را روی تخم می در این دانده داشت (مقادار میکره می در در بین هو ابود. اسانس ترکیب تمامی اسانس ها با پروتئوس دارای اثر سینرژیستی روی مرحله حشره کامل می در باری گلخانه بود. ترکیب اسانس کاکوتی با پروتئوس روی مرحله تخم اثر آنتاگونیستی نشان داد.

واژههای کلیدی: سفیدبالک گلخانه، گیاه دارویی، اسانس گیاهی، پروتئوس، اثر سینرژیستی

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