



## ORIGINAL ARTICLE

## Synthesis of New Glycine Cephalexin Condensed Polymer as Peptide Biopolymer for Controlled Release of Cephalexin

Aseel K. Al-Saffar<sup>1</sup>, Ahamed M. Abbas<sup>\*2</sup>, Dawood Salman<sup>2</sup>

<sup>1</sup>College of Pharmacy, Al-Mustansiriyah University, Baghdad, Iraq

<sup>2</sup>Department of Chemistry, College of Science, University of Misan, Maysan, Iraq

(Received: 25 May 2021

Accepted: 20 August 2021)

**KEYWORDS**

Glycine;  
Cephalexin;  
Biopolymer;  
characterization;  
Physical properties

**ABSTRACT:** A new peptide-based polymer was synthesized via polymerization of cephalexin acid chloride with glycine acid chloride with molar ratio 1:1 in condensed polymer solution. This Glycine Cephalexin peptide biopolymer was characterized by different analyses of UV, FT-IR and <sup>1</sup>H NMR spectroscopy. Also, physical properties of new synthesized Glycine Cephalexin peptide biopolymer were studied with measurement of its intrinsic viscosity at 30°C, swelling percentage in water and studying drug release in pH 4-10 at 37°C.

**INTRODUCTION**

Peptides are considering as a new class of bio-substances that have unique chemical, physical and biological properties [1]. Synthetic and natural biopolymers are with many applications in biointerface engineering, such as tissue engineering scaffolds, drug delivery, as detectors and transducers in biosensors. Contrasted to naturally occurring biopolymers, there are peptide-based biopolymer which is named under word "engineered". This peptide based biopolymers have attracted much attention many researchers as a new class of materials such as elastin-like polypeptides, poly-amino acids, silk-like proteins, tropoelastin-based peptides, coiled-coil domains, peptide amphiphiles, leucine zipper-based peptides, beta-hairpin peptides, and beta-sheet forming ionic oligopeptides [2]. For example, Spider-silk is a remarkably strong various materials that act as peptide based biopolymer [2], Another example of Peptide-based polymers is Collagen that has been found inside the human body, it forms major

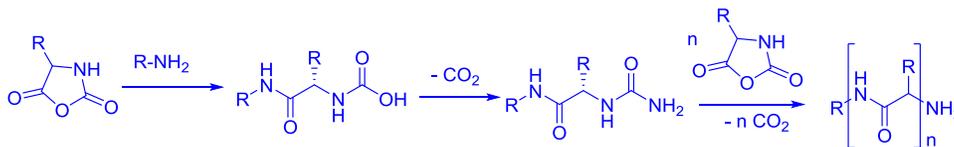
components of extracellular matrix in cell tissues to give mechanical strength to cells [3]. In the same manner, Elastin, peptide based biopolymer, has unique property of allowing body tissues to resume their shape after stretching or contracting [4-6]. Also, there are various peptide based biopolymer; poly-amino acid-based methacrylamide [7-9], biocompatible materials such as artificial skins and fibers [10].

Conventional synthesis of solid-phase peptide is not efficient in synthesis of large peptide-based polymer (>100 amino acid residues) together with conventional solution-phase peptide synthesis. These methods result in overall poor yield (10–40%) in 36–48 h. So, there are various methods that used in synthesis of polypeptides such as polymerization of  $\alpha$ -amino acid N-carboxyanhydrides as an economical and expedient process for the synthesis of relatively high molecular weight polypeptides [11] or rapid microwave-assisted solution-phase peptide

\*Corresponding author: ahmedjmn@uomisan.edu.iq (Ahamed M. Abbas)  
DOI: 10.22034/jchr.2021.685036

synthesis [12-14], for example, Chitosan hydrogels can be used as carriers for drug release and as bioactive

molecules as shown in Figure 1 [15-17].



**Figure 1.** Chitosan hydrogels as carriers for drug release and as bioactive molecules

Our research aims to synthesize new peptide based biopolymer that derived from glycine or cephalixin and determine some of its physical properties such as intrinsic viscosity, effect of pH and percentage of swelling.

## MATERIALS AND METHODS

### Instrumentation

Melting point were measured using Gallen Kamp M.F.B-600 melting point apparatus. Infrared spectrophotometer measurements were performed using Pyeunicam SP3-100. UV. Visible double beam scanning spectrophotometer-260 at room temperature. Differential Scanning calorimetry (DSC) and Thermo-ravimetric analysis (TGA) were recorded using (Pt- STA1500, Rheometric Sentic UK). All chemicals were purchased from Fluka and BDH. All analytical solvents were used without further purification.

### Preparation of Glycinoyl chloride $C_1$

In a round bottom flask equipped with a magnetic stirrer, a thermometer and a condenser, 5g glycine (1mmole) in 15 ml of Dioxane were added then 1 mmole thionyl chloride was added dropwise with rate 10 min at 0°C . Orange oily product was formed, isolated, washed with diethyl ether for several times, and dried at 50°C to give  $C_1$  with yield 75%.

### Preparation of Cephalixinoyl chloride $C_2$

Compound  $C_2$  was prepared by applying the same method of preparation of compound  $C_1$ , the yield was 80%.

### Polycondensation of [ $C_1$ ] with [ $C_2$ ] to prepared [ $C_3$ ]

A 100 mL round bottomed two necked flask equipped with a thermometer and a reflux condenser was charged

with (0.01mole) of dissolved cephalixinoyl chloride  $C_2$  the solvent was used (1ml) of 1:10 volume of DMF: Dioxane mixture were added to the flask dissolved Glycinoyl chloride  $C_1$ . Then stirred and refluxed continuously for 1h., the mixture was cooled to room temperature. The condensed polymer  $C_3$  was obtained then washed with ether and dried at 50°C the yield was 85% with  $tli_n=0.56$ .

### Swelling percentage

Swelling % of prepared polymer was determined in water for one day; swollen gels removed from the water at regular intervals were dried superficially with filter paper, weighed and placed in the same sample. S% was calculated according to the following relationship:-

$$S\% = \frac{M_1 - M_0}{M_0} \times 100$$

Where  $M_0$  is the mass of dry polymer at time 0

$M_1$  is the mass of swollen polymer at t time.

### Release studies

Condensed Cephalixin-glycine polymer [ $C_3$ J] (50mg) was kept in a cylinder containing 50:50 ml of buffer-dioxane in a water bath at 37 °C without stirring. A sample from the release medium was periodically with draws and analyzed by UV. At 300 nm to determine the amount of the released Cephalixin and glycine unites .A calibration curve was constructed with software built in the computerized UV. Spectrophotometer the pH4 and 10 were used at 37°C.

## RESULTS AND DISCUSSION

Thionyl chloride is reacted with glycine or cephalixin to afford their acid chloride derivatives ( $C_1$  and  $C_2$ ) to increase

reactivity of carboxylic acids because of presence of

chloride as good leaving group (Figure 2).

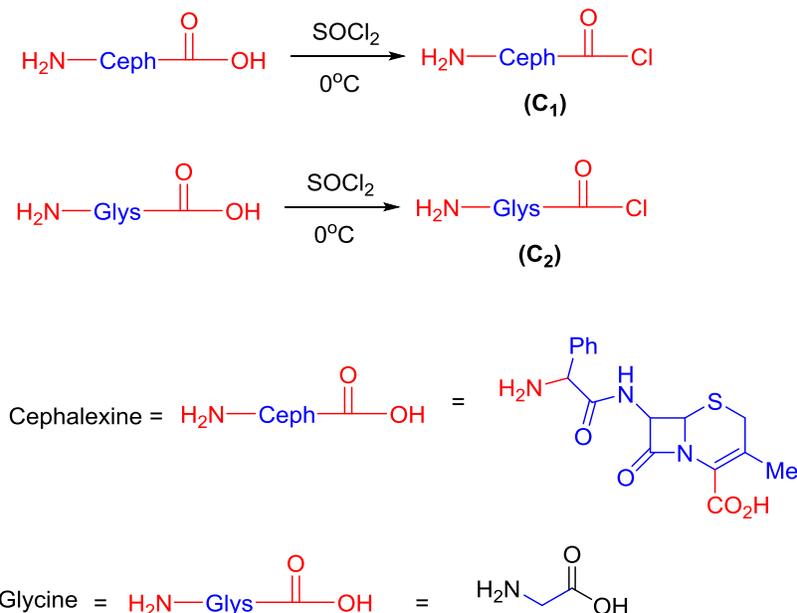


Figure 2. Mechanism reaction of Acid chloride derivatives (C<sub>1</sub> and C<sub>2</sub>) with glycine or cephalixin.

A direct condensation between glycine chloride (C<sub>1</sub>) and cephalixin chloride (C<sub>2</sub>) has been produced to afford

condensed polymer (C<sub>3</sub>) as illustrated in Figure 3.

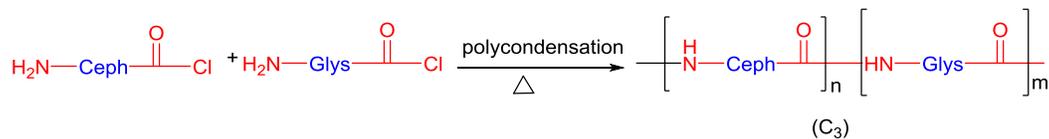


Figure 3. Mechanism condensation between (C<sub>1</sub>) and (C<sub>2</sub>) to produced condensed polymer (C<sub>3</sub>).

The mechanism of the reaction is depicted in Figure 4. A nucleophilic attack of amino group to the carbonyl carbon

with expulsion of HCl to afford the new Glycine-cephalexin based biopolymer (C<sub>3</sub>).

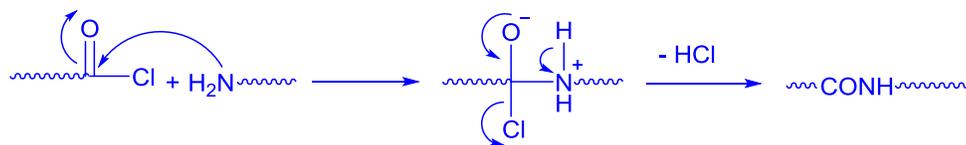


Figure 4. The mechanism of the reaction of A nucleophilic attack of amino group to the carbonyl carbon with expulsion of HCl.

Structure of new biopolymer is elucidated by using FT-IR, <sup>1</sup>H NMR as illustrated in Figures 5 and 6. IR spectrum of C<sub>3</sub> revealed presence of strong absorption band at 3200 (amide NH), 1740, 1732 (amide C=O) of cephalixin and

glycine, respectively. Also, CH aliphatic and aromatic has absorption bands at 2960-2620 and 3060. 1324 (C-N), 1114 (C-O) cm<sup>-1</sup> (Figure 5).

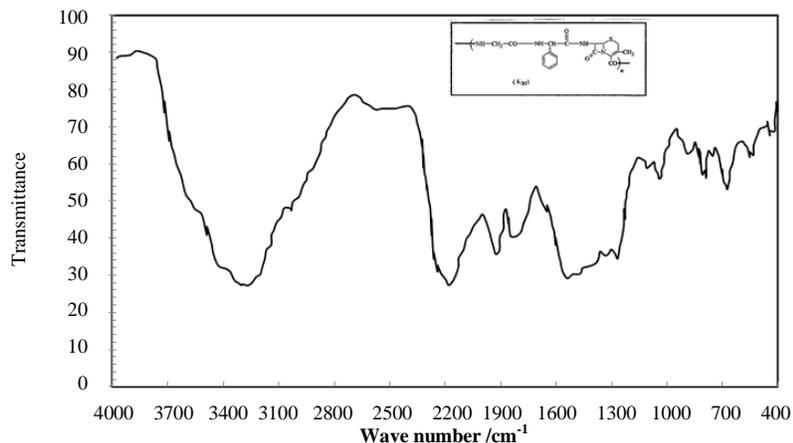


Figure 5. FTIR spectra of glycine-cephalexin polymer.

<sup>1</sup>H NMR spectrum of C<sub>3</sub> revealed presence of signals at 8.1-8.3ppm of aromatic protons, 3.2, 3.3 ppm (NH protons), and aliphatic protons of CH-Ph, CH<sub>2</sub>, CH, cyclic CH<sub>2</sub> and

CH<sub>3</sub> groups revealed strong signals at 2.8, 2.7, 1.9, 1.3, and 1.2 ppm, respectively (Figure 6).

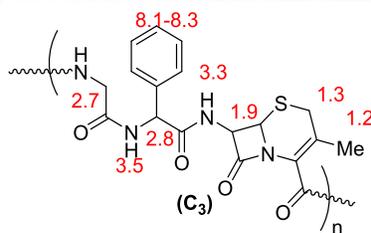
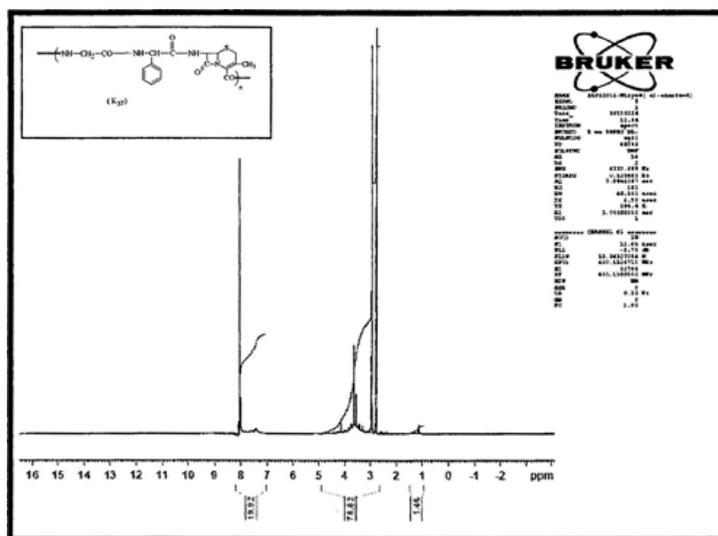


Figure 6. <sup>1</sup>H NMR spectrum of C<sub>3</sub>

**Physical Properties of C<sub>3</sub>**

**The intrinsic viscosity**

By using Ostwald viscometer, intrinsic viscosity of new prepared polymer (Figure 7) was calculated in dioxane [ $\eta_{in}$ ]

= 0.4dl/g. this result indicates that low molecular weight polymer is proportional with  $\eta_{in}$  (Table 1).

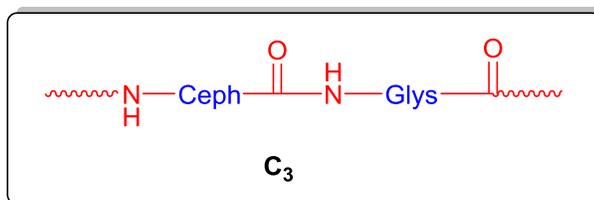


Figure 7. condensed polymer (C<sub>3</sub>)

Table 1. The Physical properties of condensed polymer C<sub>3</sub>

Poly. No.	Color	$\eta_{in}$ dl/g	Softening point °C	Yield %
C <sub>3</sub>	Yellow	0.56	120-130	85

### Effect of pH

Figure 8 shows the effect of pH values on the rate of release and profiles of mole fraction ratio to total moles present in

the sample versus time at pH4 and 10 at 37°C (Table 1).

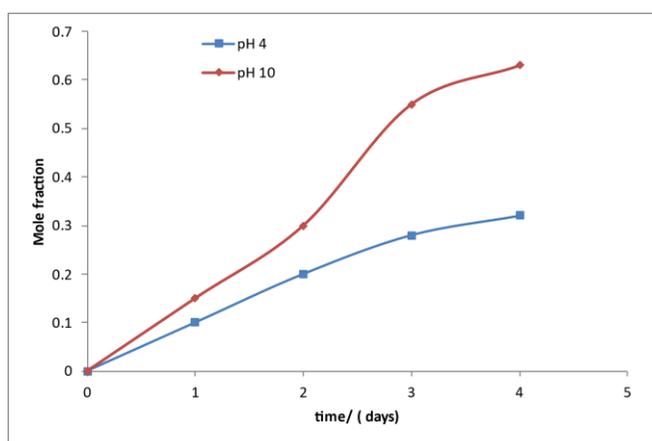


Figure 8. Effect of pH values and controlled drug release of condensed polymer (C<sub>3</sub>) in different medium

New formed amide group was hydrolyzed in basic medium to afford cephalixin and glycine units, this occurred with

sustained controlled release and to prolonged time, as shown in the following mechanism (Figure 9).

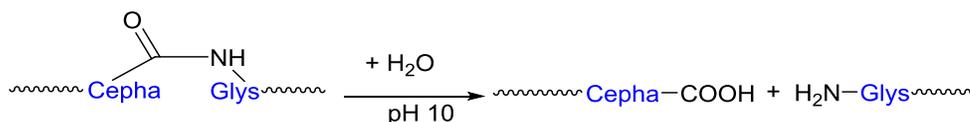


Figure 9. Mechanism reaction to hydrolyzed amid group in basic medium to afford cephalixin and glycine units.

### Swelling percentage

Swelling % was measured for C<sub>3</sub> polymer in water was high S% = 25% (Table 1).

swelling. The gradually hydrolyzed in different pH values, could enhanced the controlled release of bioactive units.

### CONCLUSIONS

New synthesized peptide-based biopolymer was characterized with measuring of some physical properties such as intrinsic viscosity, effect of pH and percentage of

### ACKNOWLEDGEMENTS

The researchers are grateful to the University of Misan and Al-Mustansiriyah University for their unlimited support for scientific work

### Conflict of interests

The author declares no conflict of interest.

### REFERENCES

1. Chow D., Nunalee M.L., Lim D.W., Simnick A.J., Chilkoti A., 2008. Peptide-based biopolymers in biomedicine and biotechnology. *Materials Science and Engineering: R: Reports*. 62(4), 125-155.
2. Shao Z., Vollrath F., 2002. Surprising strength of silkworm silk. *Nature*. 418(6899), 741-741.
3. Jenkins C.L., Raines R.T., 2002. Insights on the conformational stability of collagen. *Natural Product Reports*. 19(1), 49-59.
4. Mithieux S.M., Weiss A.S., 2005. Advances in protein chemistry. *Elastin*. 70, 437-461.
5. Hovee C., Flory P., 1974. The elastic properties of elastin. *Biopolymers: Original Research on Biomolecules*. 13(4), 677-686.
6. Davidson J.M. 2021. *Elastin: structure and biology, Connective Tissue Disease*. CRC Press
7. Duro-Castano A., Conejos-Sánchez I., Vicent M.J., 2014. Peptide-based polymer therapeutics. *Polymers*, 6(2), 515-551.
8. Hatton F.L., 2020. Recent advances in RAFT polymerization of monomers derived from renewable resources. *Polymer Chemistry*. 11(2), 220-229.
9. Namvari M., Biswas C.S., Wang Q., Liang W., Stadler F.J., 2017. Crosslinking hydroxylated reduced graphene oxide with RAFT-CTA: A nano-initiator for preparation of well-defined amino acid-based polymer nanohybrids. *Journal of Colloid and Interface Science*. 504, 731-740.
10. Cui T., Yu J., Li Q., Wang C.F., Chen S., Li W., Wang G., 2020. Large-scale fabrication of robust artificial skins from a biodegradable sealant-loaded nanofiber scaffold to skin tissue via microfluidic blow-spinning. *Advanced Materials*. 32(32), 2000982.
11. Kirschning A., Monenschein H., Wittenberg R., 2001. Functionalized polymers—emerging versatile tools for solution-phase chemistry and automated parallel synthesis. *Angewandte Chemie International Edition*. 40(4), 650-679.
12. Miao W., Chan T.H., 2005. Ionic-liquid-supported peptide synthesis demonstrated by the synthesis of Leu5-enkephalin. *The Journal of Organic Chemistry*. 70(8), 3251-3255.
13. Fuse S., Otake Y., Nakamura H., 2018. Peptide synthesis utilizing micro-flow technology. *Chemistry—An Asian Journal*. 13 (24), 3818-3832.
14. Mahindra A., Sharma K.K., Jain R., 2012. Rapid microwave-assisted solution-phase peptide synthesis. *Tetrahedron Letters*. 53(51), 6931-6935.
15. Berger J., Reist M., Mayer J.M., Felt O., Gurny R., 2004. Structure and interactions in chitosan hydrogels formed by complexation or aggregation for biomedical applications,. *European Journal of Pharmaceutics and Biopharmaceutics*. 57(1), 35-52.
16. Rajabi M., McConnell M., Cabral J., Ali M.A., 2021. Chitosan hydrogels in 3D printing for biomedical applications. *Carbohydrate Polymers*. 117768.
17. Pei M., Mao J., Xu W., Zhou Y., Xiao P., 2019. Photocrosslinkable chitosan hydrogels and their biomedical applications. *Journal of Polymer Science Part A: Polymer Chemistry*. 57(18), 1862-1871.