



ORIGINAL ARTICLE

Enhancing the Antimicrobial Properties Copper Oxide Shell with the Magnetic Mesoporous Core- Shell

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KEYWORDS

Mesoporous core-shell;
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ABSTRACT: In this work, Magnetic $\text{Fe}_3\text{O}_4@\text{MCM-41}/\text{CuO}$ nanocomposite was preparation of iron oxide magnetite nanoparticles and development of MCM-41 mesoporous shells on the surface of iron oxide magnetite after $\text{Fe}_3\text{O}_4@\text{MCM-41}$ was organofunctionalized and finally formation CuO shells with thickness ~ 25-30 nm in the surface of $\text{Fe}_3\text{O}_4@\text{MCM-41-NH}_2$ core-shell. The properties of prepared magnetic core-shell were characterized by transmission electron microscopy (TEM), X-ray diffraction (XRD), nitrogen adsorption-desorption measurement and vibration sample magnetometer (VSM). The applicability of the synthesized core-shell was tested as an antimicrobial agent against gram-positive and gram-negative bacteria. It was showed that the $\text{Fe}_3\text{O}_4@\text{MCM-41}@\text{CuO}$ act as an ideal antimicrobial agent in compared with that of the pure copper oxide and $\text{Fe}_3\text{O}_4@\text{CuO}$.

INTRODUCTION

Various metal and metal oxide materials are widely used such as magnetic storage media, solar energy conversion, electronics, sensors, batteries and catalytic converters, which include copper nanoparticles and copper oxides [1, 2]. Nanoparticles (NP) have emerged as a novel alternative to overcome bacterial multidrug resistance encountered globally due to misuse of antibiotics [3]. Use of nanoparticles as antimicrobial agents could overcome mechanisms of bacterial resistance as the microbicidal nature of nanoparticles result from direct contact with the bacterial cell wall, without the need to penetrate into the cell [3]. The development of antibacterial resistance to NPs is therefore less likely when compared to antibiotics. Nanoparticles therefore have potential to be developed into antimicrobial theranostics in medicine [3, 4]. Several methods for the synthesis of copper oxide with different

sizes and morphologies, such as vacuum vapor fermentation, solid-phase growth, micro-emulsion and hydrothermal synthesis have been developed [5-7]. However, the synthetic routes that are reported in many sources require plentiful and complex materials, reducing agents and special additives. [1, 2]. The intrinsic and super-antibacterial properties of copper oxide, compared to other antimicrobial agents, are due to longer and longer shelf life, low toxicity, low prices and good environmental environments [1]. Sometimes copper is a selective option used as an agent for water purification and the deactivation of some microorganisms and bacteria [8]. The main problem of copper oxide nanoparticles in industrial applications is the re-collection of refined water that increases industrial costs and contaminates the treated water [9]. To solve the problem of antibacterial magnetic

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nanocomposites used [9]. Several advantages of using metal oxides as antimicrobial agents can be seen in improving safety and stability in comparison with other antibacterial agents [10]. Day by day, the use of copper and its complexes increases as the effective materials in water treatment, textiles and etc. [11]. Microorganisms are affected by low amounts of copper and their complexes. Due to the insignificant sensitivity of human tissues to copper and its complexes, microorganisms exhibit sensitivity to efficiency even at low concentrations [11].

Considering that pure MCM-41 silicone materials with high surface area, high pore volume, parallel pore structure without network complexity and biocompatibility have been successfully used for such systems as drug delivery control systems and antibacterial carriers [12-17]. Many previous researches have been documented in the introduction of transition metal elements in the mesoporous silica materials to prepare redox catalysts [18]. Two preparative methods are generally available to introduce catalytically active species into the MCM-41 support, as-synthesized and post synthesized [18-20].

Some of the applications of silicon-magnetic nanoparticles include photocatalytic [12, 21 and 22], catalytic [23], and antibacterial [24] properties.

Research shows that due to its unique magnetic properties, low toxicity, good biocompatibility, the combination of magnetic nanoparticles with silicone mesoporous can be a very good option for drug delivery systems [25, 26]. In recent research, the synthesis of mesoporous silicon nanocomposites with magnetic properties, either in the form of core-shell or as a non-magnetic magnetic material [24-27] also copper deposition on mesoporous silicon oxide and other metal oxides [12] has been reported. To the best of our knowledge, there is no report on the controlled synthesis of $\text{Fe}_3\text{O}_4@\text{MCM-41}@\text{CuO}$ core-shell as an antibacterial agent. Herein, in this study, a hybrid of a magnetic Fe_3O_4 cores along with MCM-41 shells and CuO shells was constructed [24]. However, $\text{Fe}_3\text{O}_4@\text{MCM-41}@\text{CuO}$ not only shows high efficiency and selectivity towards the antimicrobial activity but also can be separated

and recovered efficiently from the reaction solution by applying external magnetic fields .

MATERIALS AND METHODS

Materials

Tetraethylorthosilicate (TEOS, 98%), ammonia (25 wt.%), the surfactant Cetyltrimethylammonium bromide (CTAB), 3-aminopropyltriethoxysilane (APTES), copper (II) nitrate ($\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$), sodium hydroxide (NaOH), sodium fluoride (NaF), Ferric trichloride hexahydrate, ethylene glycol (EG), ammonium acetate, and ethanol were purchased from Fluka and Merck (Darmstadt, Germany).

Synthesis of magnetic $\text{Fe}_3\text{O}_4@\text{MCM-41}@\text{CuO}$ core-shell

The synthesis of $\text{Fe}_3\text{O}_4@\text{CuO}$ core-shell, Fe_3O_4 core and CuO NPs was done according to the previous literatures [1, 28 and 29]. To synthesize $\text{Fe}_3\text{O}_4@\text{MCM-41}$ [29], 3 g of the synthesized Fe_3O_4 NPs was dispersed in a mixture of ammonia solution (10 mL, ammonia 25% and distilled water 100 mL) in a beaker at 40 °C. Then, 20 mL TEOS, 0.9 g NaOH and 0.2 g sodium fluoride and 14 g CTAB were added to the mixture and stirred for 2 h. Then the mixture was stirred at 40 °C for 2 h. Finally the magnetic composite in an autoclave was treated at 120 °C for 48 h. The resultant solid was filtered, washed with distilled water and dried at 60. After, the synthesized particles calcined for 3 h at 500°C.

Organofunctionalization of $\text{Fe}_3\text{O}_4@\text{MCM-41}$ was carried out according to literature [18]. In brief, 4.0 g synthesized $\text{Fe}_3\text{O}_4@\text{MCM-41}$ was charged into a round flask containing 120 mL toluene and then 6.0 mL of 3-aminopropyltriethoxysilane was added drop wise. The reflux was performed for 8 hours. The resulting solid was separated by magnet, washed with ethanol and distilled water several times, and dried at room temperature for 12h. To prepare $\text{Fe}_3\text{O}_4@\text{MCM-41}@\text{CuO}$ core-shell, 120 ml of copper (II) nitrate solutions was mixed with (0.02 M) with 2.0 g NH_2 functionalization $\text{Fe}_3\text{O}_4@\text{MCM-41}$ [12, 18]. The mixture was stirred constantly at room temperature for 24h. The solid product was washed with distilled water several

times and dried at 50°C. Eventually dried solid product calcined in a furnace at 550°C in air for 6h.

Evaluation of antibacterial properties

Fe₃O₄, CuO, Fe₃O₄@CuO, Fe₃O₄@MCM-41 and Fe₃O₄@MCM-41@CuO were diluted in different concentrations (2 to 200 µg/ml) using ethanol and then added to well microplates [24]. 0.5 McFarland of *K. pneumoniae* ESBL producer, non-ESBL producing *K. pneumoniae*, ESBL producing *E. coli*, *E. coli* b121 (free plasmid), MRSA, MSSA, MBL producing *P. aeruginosa* and non MBL producing *P. aeruginosa* were prepared and added to each well. Then different values (2-200 µg/mL) of antimicrobial candidate in each well added to 0.5 McFarland of each bacterium and subjected to evaluation for Minimum inhibitory concentration (MIC) and Minimum bactericidal concentration (MBC) [30]. Bacteria in presence and absence of antimicrobial candidate were positive and negative control respectively. Microplates incubate conditions were at 37°C for 24h. The turbidity was evaluated at 570nm by ELISA reader (enzyme-linked immune sorbent assay). To determine the MBC, the MIC value and higher concentration were cultivated on nutrient agar. CFU/ml of each bacterium was obtained in comparison with McFarland standard, [30].

Characterizations

The Magnetic Mesoporous materials for investigation constructed properties were characterized using a variety of techniques. See Electronic Supplementary information (ESI) for more details.

RESULTS AND DISCUSSION

Characterization of core-shell

The x-ray diffraction patterns of the fabricated core-shells, copper oxide (CuO) and magnetite nanoparticles were presented in Figure 1. The XRD pattern of synthesized Fe₃O₄ cores showed (Figure 1a) that Diffraction peaks 35.058° (311), 43.293° (400), 55.312° (422), 57.589° (411), 63.190° (440) and 74.081° (533) correspond to different

planes of cubic phase of Fe₃O₄ [31]. According by Scherer's formula [29, 32], the average crystal size of Fe₃O₄ core was 12 nm. The XRD pattern for copper oxide can be showed (Figure 1b) by the characteristic 32.67°, 35.63°, 38.90°, 48.83°, 53.60°, 58.24°, 61.59°, 66.5° and 68.19° refer to (111), (002), (-111), (-202), (020), (202), (-113), (022), (220) respectively that illustrates of monoclinic phase of copper oxide [24, 29]. The XRD pattern of synthesized Fe₃O₄@MCM-41 core-shell indicated (Figure 1c) Diffraction peaks 30.23°, 35.73°, 43.31°, 57.21° and 62.47° corresponds to(220), (311), (400), (411) and (440) planes of cubic phase of Fe₃O₄ and one strong diffraction peak at 2.75° refer to (100) plane of hexagonal structure from MCM-41 [12]. The XRD pattern of Fe₃O₄@MCM-41@CuO core-shell (Figure 1d) showed peaks that could be indexed to MCM-41 structure, copper oxide and Fe₃O₄. The broadening of core-shell peaks in comparison with magnetite core and copper oxide alone could be related to the quantum size effect of core-shell. Using Scherer's formula [12, 29], calculated average crystal size of MCM-41 and CuO shells and obtained 27.72 and 14.00 nm, respectively.

In Figure 2, Spectrums FTIR very well were introduced structure of Fe₃O₄@MCM-41/CuO core-shell and other perpetrated nanocomposite. The bands at 3430 cm⁻¹ and 557 cm⁻¹ in Figure 2a and Figure 2b, are corresponding to O-H stretching and Fe-O bond respectively. Also band 1625 cm⁻¹ was attributed to the bending vibration of water in the Fe₃O₄ nanoparticles [33, 12, and 34]. In Figure 2b, Figure 2c, Figure 2d, peaks 1072.33, 800.76 and 450.95 cm⁻¹ can be owned to Si-O-Si bending and stretching vibrations [12, 18]. The weak peaks at 2338.92 cm⁻¹, 2854.46 cm⁻¹, and 2925.63 cm⁻¹ were assigned to the vibrations of atmospheric CO₂ and the symmetry and asymmetry stretching vibrations of -CH₂-, respectively [12, 34]. In addition, in Figure 2c the peaks 2935, 2837.5 cm⁻¹ and 1567 cm⁻¹ can be attributed to asymmetric and symmetric vibrations of -CH₂ group of APTES molecules and amine groups, respectively. Accordingly, they show surface of Fe₃O₄@MCM-41 by APTES was successfully functionalized [18, 35]. Usually, three peaks in the range of

400–700 cm^{-1} indicate CuO nanostructures [36]. In Figure 2d, peaks 469.04 and 586.47 cm^{-1} can be corresponded to the Fe-O and Cu-O stretching vibration of Fe_3O_4 and CuO, respectively. All bands in Fe_3O_4 @MCM-41/CuO sample shift to higher wave numbers and increased intensity (except peak at 3328.55 cm^{-1}). This can be related on core-shell structure [12]. The particle sizes and the morphologies of the samples were illustrated by using SEM and TEM. SEM images for Fe_3O_4 @MCM-41 core-shell

indicated a smooth surface and slightly rough one for Fe_3O_4 @MCM-41@CuO core-shell due to copper oxide shells (Figure 3). The EDX spectra (Figure 3) of the samples explained loading of copper oxide in Fe_3O_4 @MCM-41 core-shell. Based on peaks Cu, Fe, Si and O, results EDX showed which weight percentage (wt. %) of magnetite cores, Mesoporous and copper oxide shells were calculated 70.48, 21.05 and 10.16%, respectively.

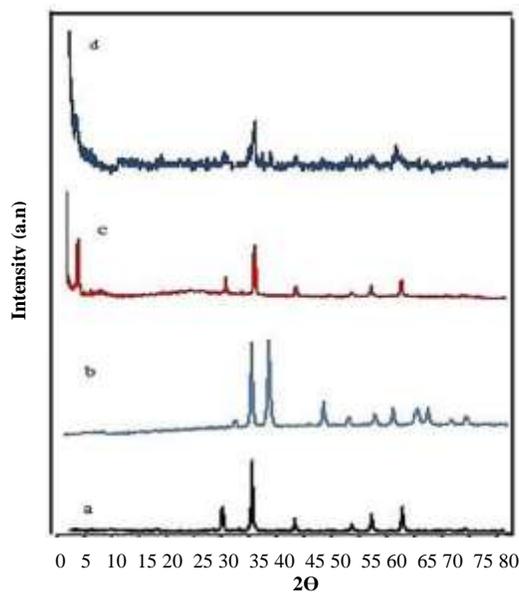


Figure 1. XRD patterns for Fe_3O_4 (a), CuO (b), Fe_3O_4 @MCM41 (c) and Fe_3O_4 @MCM41@CuO nanoparticles (d).

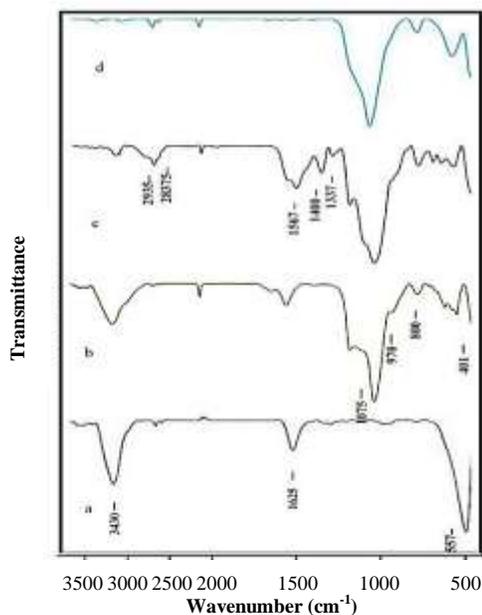


Figure 2. FTIR spectra of the Fe_3O_4 (a), Fe_3O_4 @MCM41 (b), Fe_3O_4 @MCM41- NH_2 (c), Fe_3O_4 @MCM41/CuO (d) nanocomposite.

The TEM images for core-shells were showed, presented the Fe_3O_4 cores (the black part) with diameter size between 15-20 nm and surrounded shells of MCM-41 material (~ 5-10 nm) and copper oxide average with thickness ~25-30 nm (Figure 4). The porous properties of magnetic core-shells samples were examined by measuring the nitrogen adsorption isotherms (Figure S5). The Fe_3O_4 @MCM-41 core-shell isotherm is of type IV according to the IUPAC classification of adsorption isotherms [12, 36], typical of mesoporous solids. The Brunauer–Emmett–Teller surface area (S_{BET}) Fe_3O_4 @MCM-41 core-shell was $895\text{m}^2\text{g}^{-1}$. Also the pore diameter and pore volume were 2.7 nm and 0.71 mL g^{-1} , respectively. These results indicated the presence of MCM-41 mesoporous in the new core-shell [12, 36]. The BET surface area of the copper oxide, magnetic cores according with literature [36] were 13.8 and $0.32\text{ m}^2\text{ g}^{-1}$ respectively and Fe_3O_4 @MCM-41@CuO core-shell [12] was calculated $750\text{ m}^2\text{ g}^{-1}$. These results were in agreement

with other articles [12, 33- 34]. To appraise the magnetic behavior of the magnetic core-shells, the magnetic measurements were carried out at 300 K. The saturation magnetization values (M_s) of the Fe_3O_4 MNPs, Fe_3O_4 @MCM-41 and Fe_3O_4 @MCM-41@CuO core-shells were measured to be 73.65, 23.20 and 1.84 emu g^{-1} , respectively (Figure 6). The decrease of the saturation magnetization is most probably related to the the growth of the CuO shells and MCM-41 shells on the surface on the surface of Fe_3O_4 nanoparticles [37]. Moreover, coercive force (H_c) of 50.0, 52.0 and 90.0 Oe and remnant magnetization (M_r) of 4.89, 2.40 and 0.70 emu g^{-1} were obtained for Fe_3O_4 , Fe_3O_4 @MCM-41 and Fe_3O_4 @MCM-41@CuO core-shells, respectively. These results reflect ferromagnetic character [38]. The Fe_3O_4 @MCM-41@CuO core-shell can be easily dispersed in aqueous solution and remains in suspension until exposed to a magnetic field, because it's the low coercively and remanence.

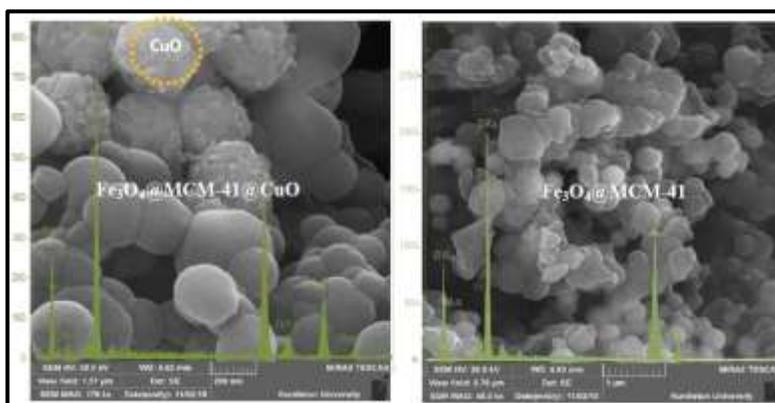


Figure 3. SEM images and EDX spectra of core-shells.

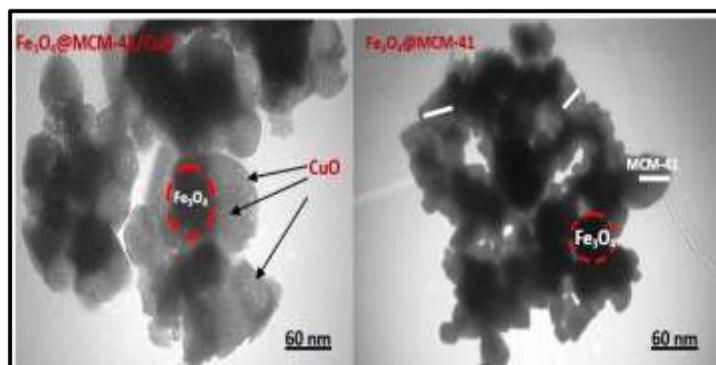


Figure 4. TEM images of core-shells

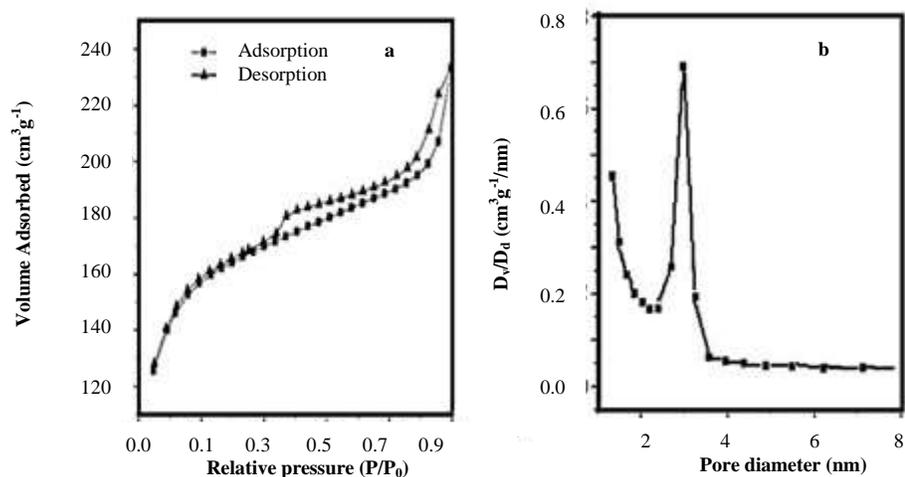


Figure 5. (a) Nitrogen adsorption-desorption isotherms and (b) Pore size distribution of the $\text{Fe}_3\text{O}_4\text{@MCM-41}$ [25].

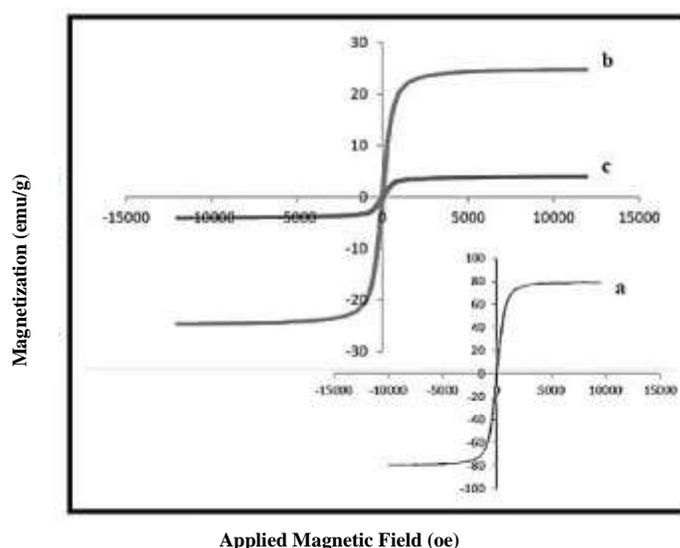


Figure 6. Hysteresis loops recorded of (a) Fe_3O_4 , (b) $\text{Fe}_3\text{O}_4\text{@MCM-41}$ and $\text{Fe}_3\text{O}_4\text{@MCM41/CuO}$ from solution under an external magnetic field.

Antibacterial mechanism

The results showed that antimicrobial activities of samples and core-shells against studied bacteria were $\text{Fe}_3\text{O}_4\text{@MCM-41@CuO}$, $\text{Fe}_3\text{O}_4\text{@CuO}$ and CuO NPs respectively. Based on the results, magnetite cores and $\text{Fe}_3\text{O}_4\text{@MCM-41}$ core-shell did not display antimicrobial activity (Table 1). Several possible antimicrobial mechanisms of metal and metal oxide NPs have been summarized in the latest studies [39]. It has been reported that, copper (II) ions released from copper oxide surface in aqueous phase are responsible for antimicrobial activity of the materials [40]. According to physiologic conditions,

bacterial cell walls are negatively charged by functional groups such as carboxyl's, phosphate and hydroxyl in lipoproteins at the surface [24, 41]. By binding the Cu^{2+} ions with the functional groups of proteins and enzymes the cell process gets inactive and inhibited. This is the main reason for the antimicrobial property of the synthesized copper oxide and $\text{Fe}_3\text{O}_4\text{@MCM-41@CuO}$ core-shell. The magnetic $\text{Fe}_3\text{O}_4\text{@MCM-41}$ core-shell together with copper oxide shell showed extraordinary antimicrobial properties against studied bacteria. This increase in antimicrobial activity can be attributed to two factors:

1. MCM-41 as a Porous surface not only prevents copper oxide aggregation which in turn barricades the release of large amount of copper (II) ions and the decrease in antimicrobial activity CuO NPs but also causes uniformity of distribution[24, 42].

2. Copper oxide particles in the magnetic mesoporous with their smaller sizes (average crystal size ~14 nm) in comparison to other copper oxide nanoparticles reported in Table 1 can release large amounts of copper (II) ions in shells [24, 42]. Findings

Table 1. Investigation antibacterial activity of samples.

Samples	Fe ₃ O ₄	CuO	Fe ₃ O ₄ @CuO	Fe ₃ O ₄ @MCM-41	Fe ₃ O ₄ @MCM-41@CuO
Bacteria	MIC (MBC)	MIC (MBC)	MIC (MBC)	MIC (MBC)	MIC (MBC)
	µg.mL ⁻¹	µg.mL ⁻¹	µg.mL ⁻¹	µg.mL ⁻¹	µg.mL ⁻¹
<i>E.coli</i> b121	-(-)	100(100)	30(30)	-(-)	5(5)
<i>E.coli</i> producing ESBL	-(-)	150(155)	55(60)	-(-)	6(10)
<i>K. pneumoniae</i> ESBL producer	-(-)	90(100)	60(65)	-(-)	8(8)
non-ESBL producing <i>K. pneumoniae</i>	-(-)	75(80)	50(55)	-(-)	8(8)
MSSA	-(-)	85(90)	40(40)	-(-)	5(5)
MRSA	-(-)	140(150)	65(70)	-(-)	6(6)
MBL producing <i>P. aeruginosa</i>	-(-)	180(200)	80(80)	-(-)	10(10)
non MBL producing <i>P. aeruginosa</i>	-(-)	185(200)	75(80)	-(-)	7(7)

Demonstrate that Fe₃O₄@MCM-41/CuO core-shell indicated strong antimicrobial activity at an early stage in comparison to other antimicrobial materials including Fe₃O₄@CuO core-shell (Table 2). According to the recent researches [1, 24] crystal size of copper oxide in new core-shells is smaller than Fe₃O₄@CuO core-shell (~22 nm) [1]. Figure 7 shows the results of tests MBC. In Figure 7A, two plates are seen; one contains Staphylococcus aureus and Fe₃O₄@MCM-41 core-shell and the other one contains

Fe₃O₄ @ MCM-41 / CuO and Staphylococcus aureus. The sign of bacterial growth; turbidity in the first plate was seen while no bacterial growths was observed in the second plate. In Figure 7B, the plate containing E. coli bacteria and Fe₃O₄ @ MCM-41 got turbid whereas the other plate containing Fe₃O₄ @ MCM-41 @ CuO samples and E. coli bacteria showed no bacterial growth which is the sign of Fe₃O₄ @ MCM-41 @ CuO positive response to antibacterial activity.

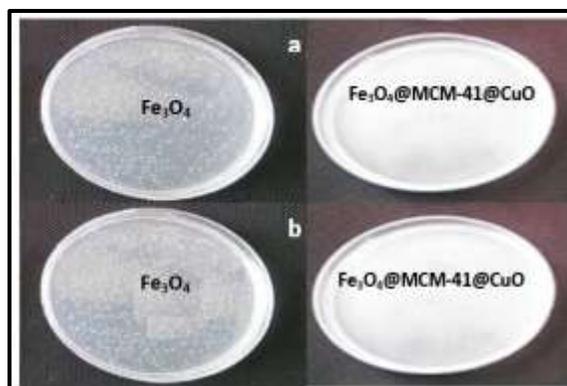


Figure 7. MBC result: Staphylococcus aureus bacteria (a), E. coli bacteria (b).

Table 2. The comparison of antimicrobial activities different

Antimicrobial agent	MIC	Reference
CuO microspheres	500 mg/L	[43]
CuO/TiO ₂ nano fibers	45000 mg/L	[10]
Fe ₃ O ₄ @CuO	40 mg/L	[1]
Fe ₃ O ₄ @MOR@CuO	17.5 mg/L	[24]
Fe ₃ O ₄ @MCM-41@CuO	5 mg/L	<i>Current work</i>

CONCLUSIONS

In this work, we have suggested a new protocol for the efficient synthesis of Fe₃O₄@MCM-41@CuO core-shell for antimicrobial activity. To the best of our knowledge, this is the first report on antimicrobial activity Fe₃O₄@MCM-41@CuO core-shell against Gram positive (G+) and Gram negative (G-) bacteria. Highlight results in this study include, phenomenal antimicrobial properties of Fe₃O₄@MCM-41@CuO core-shell and easy separation of them using an external magnetic field due to the magnetism of the Fe₃O₄. Nanocomposite characterized by TEM, SEM, XRD, FTIR, BET, VSM and EDX. TEM images confirm XRD spectrums and illustrate the magnetite cores covered with MCM-41 and copper oxide shells.

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