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**Review Article** 

# Mechanisms of Antioxidant Actions and their Role in many Human Diseases: *A Review*

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**KEYWORDSABSTRACT:** Antioxidants are substances that are available in various natural food products, which play a vital role<br/>in reducing body cell damage caused by free radical formation. An imbalance between antioxidants and free radicals<br/>contributes to an oxidative stress in the human body. The electron acceptability of O2 produced Reactive Oxygen<br/>Species (ROS);<br/>Superoxide dismutase<br/>(SOD)Species (ROS). The imbalance equilibrium between the production of reactive oxygen species (ROS) and the<br/>purification supports a rise in the ROS levels, which is the key cause of disrupted cellular activity. A recent review of<br/>excessively mild antioxidants, processes of movement, and their role in many human illnesses.

#### INTRODUCTION

Oxidative stress is an imbalance situation in which the production of reactive oxygen species (ROS) exceeds the mobile antioxidant capacity. If a certain amount of oxidative damage is close to being below normal circumstances, except that is observed, an increase in the rate of damage due to aging and disease processes, provided that antioxidant and restoration mechanisms have decreased effectiveness [1, 2].

In the cycle of oxidative phosphorylation and the construction of ATP like the instantaneous and final energy supply [3, 4], free radicals may be additionally produced like various reactive species [5, 6]. Throughout oxidation, hydrogen atoms or just electrons are transmitted from one

\*Corresponding author: Shimarb@uomisan.edu.iq (Sh. Rabeea Banoon) DOI: 10.22034/jchr.2021.683158 molecule to another, the latter being used as an antioxidant. Antioxidants will also avoid the progress of interactions of free radicals, which would in any other case result in the death of the cell or impairment of it. Nevertheless, the procedure of oxidation performs the required protection of the body from pollution or impairment of tissue; how much is needed; and in which tissues are an exceptionally tuned slice of physiology [7]. Reactive species can also perform roles such as mobile minor messengers or signaling molecules such as nitric oxide [6, 8, and 9].

The cellular signaling network is frequently disrupted by immoderate ROS; besides, the shielding outcomes that greatest alimentary phytochemicals yield are expected to be the result of a variety of vital mechanisms [10]. Many experiments have dedicated themselves to the understanding and formulation of mechanistic pathways via which these certainly derivative components can fluctuate the cell's future. These antioxidant features of phytochemicals have been implicated as means of stressrelieving [11, 12].

The defense of endogenous antioxidants as opposed to the oxygen species reactive is supported via herbal antioxidants

that maintenance them and maintain the most excellent stability via the ROS counteracting [13].

#### Sources of free radicals

Free radicals and other ROS are derived either from the human body's enogenous metabolic processes or from external sources. While exposure to radiation, ozone, smoke, cigarettes, air pollution, and industrial chemicals are external sources, Figure 1 [14].

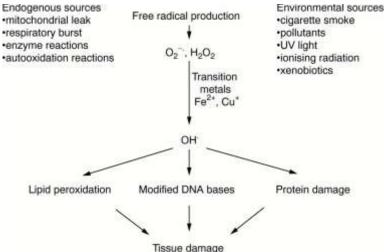


Figure 1. Main sources of free radicals [14].

#### Antioxidant classification

Antioxidants can be classified as enzymatic and nonenzymatic antioxidants (metabolic nutrient) and antioxidants. The major enzymatic antioxidants at once elaborated on the ROS counterbalancing are glutathione reductase (GRx), catalase (CAT), and superoxide dismutase (SOD) [16, 17, and 18]. SOD, the leading link of protection compared to these radicals, catalyzes the dismutation of the superoxide anion radical to (H<sub>2</sub>O<sub>2</sub>) via reduction means. The oxidant shaped  $(H_2O_2)$  is converted to  $O_2$  and  $H_2O$  via (GPx) or (CAT). The enzyme GPx selenoprotein destroys H<sub>2</sub>O<sub>2</sub> via utilization to oxidize reduced glutathione (GSH) to oxidized glutathione (GSSG). An enzyme flavoprotein, Glutathione reductase, revives GSH from GSSG, using NADPH as a low foundation of power. In addition to hydrogen peroxide, GPx decreases non-lipid or lipid hydroperoxides when oxidizing glutathione (GSH) [16]. Non-enzymatic antioxidants are additionally categorized as nutrient antioxidants and metabolic antioxidants. Metabolic

antioxidants, which belong to the endogenous antioxidants, are made by the assistance of metabolism in the body, such as bilirubin, coenzyme Q-10, glutathione, L-arginine, lipoid acid, melatonin, metal chelating protein, transferrin, uric acid, and so on [19]. Nutrient antioxidants, which belong to exogenous antioxidants, are composites that cannot be made in the body and should be provided by supplements or nutrition for instance carotenoids, diet C, diet E, flavonoids, trace metals (manganese, selenium, zinc), omega- 3, omega-6 and fatty acid. Nutrient antioxidants are connected by the purification of reactive oxygen species (ROS) [20] and play a critical character in supporting endogenous antioxidants for the oxidative tension equation [21]. The scarcity of nutrient antioxidants is one of the motives for several persistent and cancer and degenerative pathologies.

#### Antioxidant enzymes and their action mechanisms

The antioxidant is a chemical that can avoid or slowly oxidize macromolecules. The role of antioxidants is to stop or decrease these chain reactions by eliminating free radicals or constraining specific oxidation reactions by oxidizing themselves. Antioxidants are too frequently lessening producers, like thiols or polyphenols [20].

The antioxidant enzymes in entirely physical cells comprehend three primary orders of enzymes of antioxidants such as glutathione peroxidases, catalases, and superoxide dismutase (SOD), the entirety of which show indispensable turns in holding cells in their homeostasis. These enzymes' stimulations reveal a special response to contaminant oxidative tension [20]. The role of SOD is to subsist radicals of superoxide and convert them to H<sub>2</sub>O<sub>2</sub> [22, 23]. The site of GPx is accomplished with the support of the discount of different natural hydroperoxides, hydrogen peroxide, and lipid hydroperoxides [24]. Glutathione-S-transferases (GST) is a predominant category of enzymes of detoxifying [25]. A multifunctional protein family that is concerned with the detoxification of cytotoxic, genotoxic compounds and tissue protection as opposed to oxidative hazards [26,27]. In addition to other turns in the metabolism of self-evolution, the enzymes are implicated into the xenobiotic's purification, like carcinogens, medicines, and ecological contaminants in creatures, and with confrontation herbicides and insecticide in plants and pests [28].

Some studies have exposed the GPx, [30] and GST is incapacitated with the hydroperoxides support, which utilizes their poisonousness without delay, via oxidation of SH protein corporations or hydroxyl radical development. It is considered from the review that a large quantity of GST isoenzymes additionally exemplifies the GPx effort and catalyzes the markdown of natural hydroperoxides to their equivalent alcohols [31, 32]. Superoxide dismutase motivates the interruption of the superoxide anion to hydrogen peroxide and oxygen. It eliminates  $O_2$  by motivating the dismutation response. If there is a decrease in the levels of the enzyme, the reaction will be sluggish [33]. Catalase is the antioxidant enzyme which catalyzes the two-stage conversion of hydrogen peroxide to water and oxygen as in stage 1 and 2

Catalase-Fe (III) +  $H_2O_2 \rightarrow$  Compound I Stage 1

### Compound I + H<sub>2</sub>O<sub>2</sub> $\rightarrow$ Catalase-Fe (III) + 2H<sub>2</sub>O<sub>2</sub>+O<sub>2</sub> Stage 2

The above-mentioned response rate is extremely high (~107 M / s), which means that the saturation of the enzyme *in vivo* is practically impossible. Catalase is largely present in peroxisomal cells which also produce the most hydrogen peroxide-generating enzymes. Catalase consists of four chains of the polypeptide; every chain holds more than 500 acids of amino acids and encompasses 4 porphyrin heme and a molecule of NADPH. The turnover of catalase is the simplest of all the various enzymes of antioxidants. The disintegration of H<sub>2</sub>O<sub>2</sub> through the catalytic undertaking of catalase shadows the pattern of the first-order reaction and its success is set at the H<sub>2</sub>O<sub>2</sub> concentration [32, 35].

The glutathione mechanism consists of glutathione reductase, glutathione peroxidase, and glutathione S-transferases. Glutathione S-transferases are the group of enzymatic antioxidants that motivate lipid peroxide break. Glutathione peroxidase in the body results in an unnecessary use of hydrogen peroxide and human hydroperoxides [36].

Glutathione reductase motivates the decline of oxidized glutathione (GS SG) to condensed glutathione (GS H).

#### Non -enzymatic antioxidants and the mechanisms action

Non-protein thiol and Protein-enchained thiol represent a shielding agent and cell-decreasing contra most inorganic contaminants through the SH-group [37]. Consequently, the headline of the protection contra stress oxidation is the thiol. The thiol degrees can be extended through the appliance of adaptation to a mild oxidative tension by amplifying its production. Glutathione is the mobile antioxidant that achieves an essential part in the redox of the cells [37].

Ascorbic acid is an antioxidant that can only be obtained from the human diet as it cannot be synthesized by the body. Vitamin E has been shown to preserve mobile membranes from oxidation by intermediates free radical and reacting by lipid radicals [39]. Beta carotene has a solid antioxidant area through disposing of singlet oxygen to defend contra attacks of a free radical. It has been found in cabbage, cheese, grains, liver, tomatoes, and milk [40, 41]. Flavonoids play an important position in the safety of oxidative stress [42] with cancer in particular. Flavonoids are commonly present in tea, grains, berries, and cocoa [43]. They are found in high amounts in drinks and nutrients [44] of which the antioxidation has been studied in particular [45]. Natural antioxidants improve ROS' endogenous antioxidants and preserve the highest level of stability with the aid of neutralizing reactive species [46]. Antioxidant phenol-based products are linked to several different processes, such as hydrogen donation, single oxygen quenching, steel ion chelation, and radically hydroxylated and superoxide substrates [47].

The natural antioxidants have a range of biochemical activities, as free radical\_scavenging, and inhibition of ROS production. The literature suggested that the ingestion of curcumin, garlic, peppermint, pomegranate, rosemary, sea buckthorn seedcake, and sesame had proven protecting impacts in comparison to renal illnesses and nephrotoxic retailers, leading to renal dysfunction in humans and laboratory creatures [13, 47, 48 and 49].

#### Antioxidants function

The Administration of Food and Drug (FDA) describes antioxidants exclusively as nutritional additions to be used as well as daily food intake in an attempt to combat these illnesses [50]. Antioxidants are recognized to play a key role in the defense of the effects on the use of plant ingredients [51-54]. Regular consumption of greens and fruits has been described as a reduction in the risk of chronic disease [55]. Studies show that an antioxidant rich diet has an actual beneficial effect, fitness affects in the long run [56]. Recently, antioxidants have gained significant interest in radical and oxidative stress, most cancer prophylaxis and treatment, and robustness [57]. Skim milk can also be used as a dietary supplement as an antioxidant activity to minimize the risk of tobacco smoking or tobacco chewing [58]. Both antioxidants are involved as a system of the antioxidant, squad, accountable for avoiding the harmful properties of free radicals and the toxic merchandise of the metabolism. Though, the antioxidant works to exploit free radical development as a coordinated tool, leaving deficiencies in one problem affecting the effectiveness of others [59, 60].

#### Roles of antioxidants in the prevention of diseases cancer

Antioxidants preserve DNA by failing the impairment of oxidative DNA produced by the free radical and by managing the multiplied strange division of the cell, the central attribute of carcinogenesis. Using *in vitro* and systems of animal modeling, it has been experimentally observed that plant\_consequent phytochemicals, such as sulforaphanes, isothiocyanates and, allyl sulfides inhibit several pathogens <sup>[45]</sup> has stated that the aggregate of selenium, E-nutrition, pointedly decreases the risk of progression in many types of tumors, particularly in stomach cancer [61, 62].

#### Atherosclerosis

Atherosclerosis is a cardiovascular state that begins because of the oxidized fatty acids deposition in the arteries in a plaque formation. Around two-thirds of the serum LDL cholesterol pool is an insufficient density lipoproteincholesterol which is assumed to reveal a critical role in improving atherosclerosis <sup>[47]</sup>. Some plant-derived polyphenols and flavonoids, which occur in some vegetables and fruits, have been revealed to be effective antioxidants that are active in stopping LDL. Oxidation is persuaded via free radicals. The suggested allowance for flavonoids is one gram in a regular diet, which is appropriate for the system of antioxidant protection. Ironically, it has been determined that the antioxidant recreation of certain flavonoids synergistically upsurges when complemented via acid of a-ascorbic to avoid the oxidation of LDL. Furthermore, aspirin has been shown to prevent atherosclerosis in animal studies [63].

#### Alzheimer's disease

Alzheimer's disease is featured at an advanced loss of reminiscence as a significant clinical appearance. Research on free radicals recommends that the stress of oxidative triggers neurodegenerative diseases in tandem with AD. Besides, the metal ion plays a critical turn in the development of A D. Nutraceutical. Antioxidants like lycopene, turmeric, curcumin, etc. reportedly have the beneficial effects of a variety of neural disintegration varieties, oxidative tension, and dysfunction of mitochondria [64].

Decreased levels of the antioxidant enzyme, such as superoxide dismutase, is associated with the prevalence of Alzheimer's disease in humans [65]. Reference has been made to the fact that the supplementation of nutritional vitamins E and C to the affected person can substantially increase the values of nutritional vitamins in plasma and minimize the lipoprotein oxidation, although Vitamin E alone does not have any major effects at present. Strong nutraceutical intake is postponing the treatment of dementias such as Alzheimer's disease [66].

#### Parkinson's disease

Parkinson's disease outcomes from neuronal cell damage in some parts of the brain and is denoted weakness of muscles, trembling, and trouble walking [3]. In the study [68], it was argued that vitamin E in a diet can also protect contra Parkinson's disease. Also, Glutathione has exposed some encouraging results in initial studies to deal with Parkinson's disease, but side effects, the most beneficial technique of administration, and splendid long-term dosing are not yet clear.

#### Heart diseases

There are a variety of causes to heart diseases, such as high cholesterol, obesity, diabetes, smoking cigarettes that deliver a forum for the progress of illness of coronary heart. Oxidation of dipped density induces accumulation of fatty acids in the arteries leading to the progress of atherosclerosis, which eventually induces coronary heart disease, heart disease is obtained with age because oxidized fatty acid is more 'sticky' and more likely to bind to the artery walls. An excessive intake of ascorbic acid is thought to restore endothelial dysfunction and to protect circulating lipoprotein from free radicals [69].

#### Diabetes

Diabetes mellitus (DM) is characterized by hyperglycemia [70]. In addition, oxidative tension because of deficiency of antioxidant defenses can also cause [71, 72]. It is assumed that if ROS is complicated in diabetes genesis, antioxidants may also be an advantageous strategy for diabetes prevention, [73] published that vitamin E supplementation decreases the sensitivity of DL to in vitro oxidation and the accessibility of oxidized LDL in kind two diabetics [74]. It is assumed that the unevenness between technology and the subsisting of free radicals is the fundamental reason for diabetes. Insulin should improve the absorption of vitamin C from the cell however in hyperglycemic stipulations this system is disrupted ensuing in a circumstance acknowledged as 'tissue scurvy'. The accession of Vitamin C dominates the glucose of blood, advances endotheliumdependent vasodilation and increases the resistance of lipoprotein in the direction of oxidation in the affected person by both sort one or sort two diabetes mellitus [75].

#### Skin aging

In the collagen molecule, vitamin C as a cofactor is both needed in order to add hydroxy groupings to the amino acid proline and lysine via prolylhydroxylate and lysyl hydroxylate. Hydroxylation enables the three-fold structure of the collagen molecule to become important for the production and maintenance of scar, blood vessels, and cartilage. Moreover, the mRNA level of Collagen I and III, their treatment enzymes, and the tissue-inhibitor of matrix metalloproteinase 1 in the human derma will tend to increase topically to applied vitamin C [76-78].

#### Ocular disease

A major factor in the production of cataracts and the agerelated retina disease maculopathy is thought to be the oxidative processes. It is suspected that oxidation is a major cause of damage to the lens proteins, caused primarily by UV exposure. The oxidated protein reduces in amount and induces lens cloudiness. The antioxidants and antioxidant enzymes inactivate the deterioration and removal of hazardous free radicals and proteases from the lens; however, oxidative damage happens more quickly. Therefore, the oxidized protein will build up and the damage is permanent over time. Whether antioxidant lowers risks of cataract formation and development are important to evaluate. People that are more likely to be at lower risk for cataracts if those have higher plasma levels of different antioxidant nutrients [79].

#### Antioxidant biomarkers

Assessable hazard features, biomarkers intermediates have described the progress of evidence-based therapies for cardiovascular conditions such as blood tension and lipoproteins and diabetes such as physical obesity, and recent mainstream glycemic popularity. Nevertheless, there has been a biomarker proliferation that purports to point to food consumption, its metabolic penalties, and genetic predisposition to sickness and untimely mortality [80,81]. When it arises to antioxidants and its healthiness insinuations, the detected fluctuations in intermediates may also have extra side effects. Such biomarkers that symbolize harm and are analogs to what can also occur in tissues possibly will also have value. Examples may be the combination of HbA1c and AGEs for individuals with diabetes, lipid peroxides in lipoprotein conditions, and broken DNA [80,81] even though the data are contradictory.

#### Antioxidant toxicity

The greatest noticeable detriment to antioxidants is the elimination of the health defending tasks of oxidants consisting of antimicrobial phagocytosis and the purification of complicated and apoptotic functions of cytochrome P-450 for unwanted cells [81-87].

Some antioxidant nutrients, such as Vitamin C, should anticipate oxidant functions that can also be harmful, such as Fenton's reaction as it absorbs metal ions like iron [88-92]. Several medical studies have stated that antioxidants, such as beta-carotene, which is a precursor of Vitamin A, can escalate the danger of most cancers when managed as a remote supplement [89-93]. In the APPP (Australian Polyp Prevention Project) with involvements of a trivial fats diet, beta –carotene., whilst the mixture of low fats and bran of wheat avoided the repetition of massive polyps of adenomatous, beta-carotene expanded the danger of whichever polyp repetition in females [94-97].

#### CONCLUSIONS

The current study contributes the useful awareness of depicting the oxidative stress and free radicals causing diseases. The antioxidants have a high ability to be used as most traditional treatments of diseases are used. Hence, a diet with good supplementation of vegetables, fruits, and, nuts) that have adequate essential antioxidants such as vitamins A, E, C, etc. may be sufficient to strengthen our body's immune system to prevent many diseases.

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#### REFERENCES

1. Davari A., Solouki M., Fazeli-Nasab B., 2018. Effects of jasmonic acid and titanium dioxide nanoparticles on process of changes of phytochemical and antioxidant in genotypes of *Satureja hortensis* L. Eco-Phytochemical Journal of Medicinal Plants. 5(4), 1-20 [In Persian].

2. Baineni R., Gulati R., Delhi CK., 2017. Vitamin A toxicity presenting as bone pain. Archives of Disease in Childhood. 1;102(6), 556-8.

3. Speakman J.R. 2003. Oxidative phosphorylation, mitochondrial proton cycling, free-radical production, and aging. Advances in Cell Aging and Gerontology. 1; 14, 35-68.

4. PhD OI. 1999. Free radicals, antioxidants, and international nutrition. Asia Pacific Journal of clinical nutrition.8(1), 53-63.

5. Gruhlke M.C., Slusarenko A.J. 2012. The biology of reactive sulfur species (RSS). Plant Physiology and Biochemistry. 1; 59, 98-107.

6. Halliwell B., 2006. Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. Plant Physiology. 1;141(2), 312-22.

7. Herbert V., 2001. 21 Vitamin, Mineral, Antioxidant, and Herbal Supplements: Facts and Fictions. Behavioral Neurology in the Elderly. 23-27.

8. Kimura H., Nagai Y., Umemura K., Kimura Y., 2005. Physiological roles of hydrogen sulfide: synaptic modulation, neuroprotection, and smooth muscle relaxation. Antioxidants & Redox Signaling. 1;7(5-6), 795-803.

9. Liu D., Jin H., Tang C., Du J., 2010. Sulfur dioxide: a novel gaseous signal in the regulation of cardiovascular functions. Mini Reviews in Medicinal Chemistry. 1;10(11), 1039-45.

10. Calabrese V., Cornelius C., Mancuso C., Pennisi G., Calafato S., Bellia F., Bates T.E., Stella A.M., Schapira T., Kostova A.T., Rizzarelli E., 2008. Cellular stress response: a novel target for chemoprevention and nutritional neuroprotection in aging, neurodegenerative disorders and longevity. Neurochemical Research. 1;33(12), 2444-71.

11. Na H.K., Surh Y.J., 2008. Modulation of Nrf2mediated antioxidant and detoxifying enzyme induction by the green

tea polyphenol EGCG. Food and Chemical Toxicology. 1;46(4), 1271-8.

12. Tuzcu M., Sahin N., Karatepe M., Cikim G., Kilinc U., Sahin K., 2008. Epigallocatechin-3-gallate supplementation can improve antioxidant status in stressed quail. British Poultry Science. 1;49(5), 643-8.

13. Azab A.E., Albasha M.O., 2018. Hepatoprotective effect of some medicinal plants and herbs against hepatic disorders induced by hepatotoxic agents. J Biotechnol Bioeng. 2(1), 8-23.

14. Langseth L., 1996. Oxidants, antioxidants and disease prevention. ILSI. 16, 840–841.

15. Young I.S., Woodside J.V., 2001. Antioxidants in health and disease. Journal of Clinical Pathology. 1;54(3), 176-86.

16. Halliwell B., 2007. Biochemistry of oxidative stress. Biochem Soc Trans. 35, 1147-1150\_

17. Pacher P., Beckman J.S., Liaudet L., 2007. Nitric oxide and peroxynitrite in health and disease. Physiological Reviews. 87(1), 315-424.

18. Hagr T.E., Adam I.A., Mohammed E.H., El mageed M.A., 2021. GC/MS Analysis and Antioxidant Activity of Fixed Oil from Sudanese Safflower (*Carthamus tinctorius* L) Seeds.

International Journal of Advanced Biological and Biomedical Research. 9(2), 138-146.

19. Willcox J.K., Ash S.L., Catignani G.L., 2004. Antioxidants and prevention of chronic disease. Critical Reviews in Food Science and Nutrition. 1;44(4), 275-95.

20. Gupta R.K., Singh N., 2013. Morinda citrifolia (Noni) alters oxidative stress marker and antioxidant activity in cervical cancer cell lines. Asian Pac J Cancer Prev. 1;14(8), 4603-6.

21. Donaldson M.S., 2004. Nutrition and cancer: a review of the evidence for an anti-cancer diet. Nutrition Journal. 1;3(1), 19.

22. Cheung C.C., Zheng G.J., Li A.M., Richardson B.J., Lam P.K., 2001. Relationships between tissue concentrations of polycyclic aromatic hydrocarbons and antioxidative responses of marine mussels, Perna viridis. Aquatic Toxicology. 1;52(3-4), 189-203.

23. Tappel M.E., Chaudiere J., Tappel A.L. 1982. Glutathione peroxidase activities of animal tissues. Comparative Biochemistry and Physiology Part B: Comparative Biochemistry. 1;73(4), 945-9.

24. McCord J.M., Fridovich I., 1969. Superoxide dismutase an enzymic function for erythrocuprein (hemocuprein). Journal of Biological Chemistry. 25;244(22), 6049-55.

25. Hayes J.D., Pulford D.J., 1995. The glut athione S-transferase supergene family: regulation of GST and the contribution of the lsoenzymes to cancer chemoprotection and drug resistance part I. Critical Reviews in Biochemistry and Molecular Biology. 1;30(6), 445-520.

26. Mannervik B., Helena Danielson U., Ketterer B., 1988. Glutathione transferases—the structure and catalytic activity. Critical Reviews in Biochemistry. 1;23(3), 283-337. 27.25. Pickett C.B., Lu A.Y., 1989. Glutathione Stransferases: gene structure, regulation, and biological function. Annual review of Biochemistry. 58(1), 743-64.

28. Hayes P.C., Bouchier I.A., Beckett G.J., 1991. Glutathione S-transferase in humans in health and disease. Gut. 32(7), 813.

29. Pigeolet E., Corbisier P., Houbion A., Lambert D., Michiels C., Raes M., Zachary M.D., Remacle J., 1990. Glutathione peroxidase, superoxide dismutase, and catalase inactivation by peroxides and oxygen derived free radicals. Mechanisms of Aging and Development. 15;51(3), 283-97.

30. Prohaska J.R., 1991. Changes in Cu, Znsuperoxide dismutase, cytochrome c oxidase, glutathione peroxidase, and glutathione transferase activities in copper-deficient mice and rats. The Journal of Nutrition. 1;121(3), 355-63.

31. Zelko I.N., Mariani T.J., Folz R.J. 2002. Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. Free Radical Biology and Medicine. 1;33(3), 337-49.

32. Mosialou E., Ekström G., Adang A.E., Morgenstern R., 1993. Evidence that rat liver microsomal glutathione transferase is responsible for glutathione-dependent protection against lipid peroxidation. Biochemical Pharmacology. 22;45(8), 1645-51.

33. Nozik-Grayck E., Suliman H.B., Piantadosi C.A., 2005. Extracellular superoxide dismutase. The International Journal of Biochemistry & Cell Biology. 1;37(12), 2466-71. .Berg J.M., Tymoczko J.L., Stryer L., Clarke N.D., 2002. DNA replication, recombination, and repair. Biochemistry. 5th edition. New York, WH Freeman & Co. pp1119-1120. 35. Kirkman H.N., Galiano S., Gaetani G.F., 1987. The function of catalase-bound NADPH. Journal of Biological Chemistry. 15;262(2), 660-6.

36. Sharma R., Yang Y., Sharma A., Awasthi S., Awasthi Y.C., 2004. Antioxidant role of glutathione S-transferases: protection against oxidant toxicity and regulation of stress-mediated apoptosis. Antioxidants and Redox Signaling. 1;6(2), pp 289-300.

37. Mosialou E., Ekström G., Adang A.E., Morgenstern R., 1993. Evidence that rat liver microsomal glutathione transferase is responsible for glutathione-dependent protection against lipid peroxidation. Biochemical Pharmacology. 22;45(8), pp 1645-51.

38.36. Ulusu N.N., Tandoğan B., 2007. Purification and kinetic properties of glutathione reductase from bovine liver. Molecular and cellular biochemistry. 1;303(1-2), 45-51.

39. Babich H., Gold T., Gold R., 2005. Mediation of the *in vitro* cytotoxicity of green and black tea polyphenols by cobalt chloride. Toxicology Letters. 15;155(1), 195-205.

40. Linster C.L., Van Schaftingen E., 2007. Vitaminc. The FEBS Journal. 1;274(1), 1-22.

41. Abd El-Hameid A.R., 2021. *In vitro* Callus Induction of Tomato and Evaluation of Antioxidant Activity of Aqueous Extracts and Enzymatic Activities in Callus Cultures. International Journal of Advanced Biological and Biomedical Research. 9(1), 9-19.

42. Duarte T.L., Lunec J., 2005. When is an antioxidant not an antioxidant? A review of novel actions and reactions of vitamin C. Free Radical Research. 39(7), 671-86.

43.40. Matito C., Mastorakou F., Centelles J.J., Torres J.L., Cascante M., 2003. Antiproliferative effect of antioxidant polyphenols from grape in murine Hepa-1c1c7. European Journal of Nutrition. 1;42(1), 43-9.

44. Harborne J.B., Williams C.A., 2000. Advances in flavonoid research since 1992. Phytochemistry. 1;55(6), 481-504.

45. Vaya J., Mahmood S., Goldblum A., Aviram M., Volkova N., Shaalan A., Musa R., Tamir S., 2003. Inhibition of LDL oxidation by flavonoids in relation to their structure and calculated enthalpy. Phytochemistry. 1;62(1), 89-99.

46. Fetouh F.A., Azab A.E., 2014. Ameliorating effects of curcumin and propolis against the reproductive toxicity of gentamicin in adult male guinea pigs: Quantitative analysis and morphological study. American Journal of Life Sciences. 30;2(3), 138-49.

47. Mehta M., Kant V., Varshneya C., 2013. Screening of *in vitro* antioxidant potential of seabuckthorn seedcake extracts. Journal of Complementary Medicine Research. 2(2), 99-104.

48. Azab A.E., Albasha M.O., Elsayed A.S., 2017. Prevention of nephropathy by some natural sources of antioxidants. Yangtze Medicine. 13;1(04), 235.

49.46. Cyril D.G., Landry K.S., Francois K.Y., Abou B., Felix Y.H., Timothee O.A., 2016. Evaluation of nephroprotective activity of aqueous and hydroethanolic extracts of Trema guineensis leaves (Ulmaceae) against gentamicin-induced nephrotoxicity in rats. International Journal of Biochemistry Research & Review. 15(2), 1-10.

50. Ohlsson T., Bengtsson N., 2002. editors. Minimal processing technologies in the food industries. Elsevier; p. 85.

51. Gey K.F., Puska P., Jordan P., Moser U.K., 1991. Inverse correlation between plasma vitamin E and mortality from ischemic heart disease in crosscultural epidemiology. The American Journal of Clinical Nutrition. 1;53(1), 326S-34S.

52. Gey K.F., 1990. The antioxidant hypothesis of cardiovascular disease: epidemiology and mechanisms. Biochemical Society Transaction. 18(6), 1041-1045.

53. Liyana-Pathirana C.M., Shahidi F., Alasalvar C., 2006. Antioxidant activity of cherry laurel fruit (*Laurocerasus officinalis* Roem.) and its concentrated juice. Food Chemistry. 1;99(1), 121-8.

54. Fazeli-nasab B., Moshtaghi N., Forouzandeh M., 2019. Effect of Solvent Extraction on Phenol, Flavonoids and Antioxidant Activity of some Iranian Native Herbs. Scientific Journal of Ilam University of Medical Sciences. 27(3), 14-26.

55. Dembinska-Kiec A., Mykkänen O., Kiec-Wilk B., Mykkänen H., 2008. Antioxidant phytochemicals against type 2 diabetes. British Journal of Nutrition. 99(E-S1), ES109-17.

56. Sin H.P., Liu D.T., Lam D.S., 2013. Lifestyle modification, nutritional and vitamins supplements for age-related macular degeneration. Acta Ophthalmologica. 91(1), 6-11.

57. Kalcher K., Svancara I., Buzuk M., Vytras K., Walcarius A., 2009. Electrochemical sensors and biosensors based on heterogeneous carbon materials. Monatshefte für Chemie-Chemical Monthly. 1;140(8), 861-89.

58. Abdel-Wahhab D., Hassan M.A., Bakeer R.M., Mohammed H.E., 2020. Chemical composition, antioxidant activity and preventive role of milk byproducts against nicotine-induced alteration in sexual hormones and organs pathology in rats. Egyptian Journal of Chemistry. (Articles in Press)

59. Surai P.F., 2007. Natural antioxidants in poultry nutrition: new developments. In Proceedings of the

16th European symposium on poultry nutrition. World Poultry Science Association. 26-30.

60. Fazeli-Nasab B., Jafarvand N., Fooladvand Z., presented in part at the Internation Conference on Sustainable Development, Strategies and Challenges with a Focuse on Agriculture, Natural resources, Environment and tourism, tabriz, Iran, doi: 10.13140/RG.2.1.2876.2084, 2015.

61. Milner J.A., 1994. Reducing the risk of cancer. In Functional foods. Springer, Boston. pp. 39-70

62. Omenn G.S., Goodman G., Thornquist M., Grizzle J., Rosenstock L., Barnhart S., Balmes J., Cherniack M.G., Cullen M.R., Glass A., Keogh J., 1994. The  $\beta$ -carotene and retinol efficacy trial (CARET) for chemoprevention of lung cancer in high-risk populations: smokers and asbestos-exposed workers. Cancer Research. 1;54(7 Supplement), 2038s-43s.

63. Jaichander P., Selvarajan K., Garelnabi M., Parthasarathy S., 2008. Induction of paraoxonase 1 and apolipoprotein AI gene expression by aspirin. Journal of lipid research. 1;49(10), 2142-8.

64. Glenville M., 2006. Nutritional supplements in pregnancy: commercial push or evidence based? Current Opinion in Obstetrics and Gynecology. 1;18(6), 642-7.

65. Kontush A., Mann U., Arlt S., Ujeyl A., Lührs C., Müller-Thomsen T., Beisiegel U., 2001. Influence of vitamin E and C supplementation on lipoprotein oxidation in patients with Alzheimer's disease. Free Radical Biology and Medicine. 1;31(3), 345-54.

66. Haider B.A., Bhutta Z.A., 2017. Multiplemicronutrient supplementation for women during pregnancy. Cochrane Database of Systematic Reviews. (4), 1-5.

67. Anwar F., Latif S., Ashraf M., Gilani A.H. 2007. Moringa oleifera: a food plant with multiple medicinal uses. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives. 21(1), 17-25.

68. Losso J.N., 2003. Targeting excessive with functional angiogenesis foods and Trends nutraceuticals. in Food Science & Technology. 1;14(11), 455-68.

69. Ting H.H., Timimi F.K., Haley E.A., Roddy M.A., Ganz P., Creager M.A., 1997. Vitamin C improves endothelium-dependent vasodilation in forearm resistance vessels of humans with hypercholesterolemia. Circulation. 17;95(12), 17-22.

70. Grill V., Björklund A., 2000. Dysfunctional insulin secretion in type 2 diabetes: role of metabolic abnormalities. Cellular and Molecular Life Sciences CMLS. 1;57(3), 429-40.

71. Bonnefont-Rousselot D., Bastard J.P., Jaudon M.C., Delattre J., 2000. Consequences of the diabetic status on the oxidant/antioxidant balance. Diabetes and Metabolism. 1;26(3), 163-77.

72. West I.C., 2000. Radicals and oxidative stress in diabetes. Diabetic Medicine. 17(3), 171-80.

73. Reaven P., 1995, Dietary and pharmacologic regimens to reduce lipid peroxidation in non-insulindependent diabetes mellitus. The American Journal of Clinical Nutrition. 1;62(6), 1483S-9S.

74. Liao J.K., Shin W.S., Lee W.Y., Clark S.L., 1995. Oxidized low-density lipoprotein decreases the expression of endothelial nitric oxide synthase. Journal of Biological Chemistry. 6;270(1), 319-24.

75. Kawano H., Motoyama T., Hirashima O., Hirai N., Miyao Y., Sakamoto T., Kugiyama K., Ogawa H., Yasue H., 1999. Hyperglycemia rapidly suppresses flow-mediated endothelium-dependent vasodilation of brachial artery. Journal of the American College of Cardiology.1;34(1), 146-54.

76. Prockop D.J., Kivirikko K.I., 1995. Collagens: molecular biology, diseases, and potentials for therapy. Annual Review of Biochemistry. 64(1), 403-34.

77. Peterkofsky B., 1991. Ascorbate requirement for hydroxylation and secretion of procollagen: relationship to inhibition of collagen synthesis in scurvy. The American Journal of Clinical Nutrition.1;54(6), 1135S-40S.

78. Nusgens B.V., Colige A.C., Lambert C.A., Lapière C.M., Humbert P., Rougier A., Haftek M., Richard A., Creidi P., 2001, Topically applied vitamin C enhances the mRNA level of collagens I and III, their processing enzymes and tissue inhibitor of matrix metalloproteinase 1 in the human dermis. Journal of Investigative Dermatology.1;116(6), 853-9.

79. Taylor A., 1992. Role of nutrients in delaying cataracts. Annals of the New York Academy of Sciences. 30; 669:111.

80. Bjelakovic G., Nikolova D., Gluud L.L., Simonetti R.G., Gluud C., 2012, Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. Cochrane Database of Systematic Reviews. (3), 1-8.

81. Møller P., Vogel U., Pedersen A., Dragsted L.O., Sandström B., Loft S., 2003. No effect of 600 grams fruit and vegetables per day on oxidative DNA damage and repair in healthy nonsmokers. Cancer Epidemiology and Prevention Biomarkers. 1;12(10), 1016-22.

82. Naili M., Errayes A., Alghazeer R., Mohammed W. A., Darwish M., 2020. Evaluation of Antimicrobial and Antioxidant Activities of Psidium guajava L growing in Libya. International Journal of Advanced Biological and Biomedical Research. 8(4), 419-428.

83. Salganik R.I., 2001, The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population. Journal of the American College of Nutrition.1;20(sup5), 464S-72S.

84. Mahdieh G., Fazilati M., Izadi M., Pilehvarian A., Nazem H., 2020, Investigation of ACE Inhibitory Effect and Antioxidant Activity of Peptide Extracted from Spirulina Platensis, Chemical Methodologies. 4(2), 172-180.

85. Ataei Moghadam S., Rostami Charati F., Akbari R., Gholamalipour Alamdari E., Behmanesh B., 2020, Consideration antimicrobial and antioxidant properties of anbarnesa smoke ointment. Journal of Medicinal and Chemical Sciences. 3(3), 245-253.

86. Yousefi S., Nemati Karimooy F., Miyanbandi T., Esmaeilpour F., 2020, Dietary Supplementing with Resveratrol Improves Antioxidant Status in Hypercholestrolemic Rats. GMJ Medicine. 1(2), 72-78.

87. Gupta M., Panizai M., Tareen M., Ortega-Martinez S., Doreulee N., 2020. An Overview on Novel Antioxidant and Anti-Cancer Properties of Lycopene: A Comprehensive Revie'. GMJ Medicine. 1(2), 43-48.

88. Shahamatpour M., Tabatabaee Ghomsheh S.M., Maghsoudi S., Azizi S., 2021, Fenton Processes, Adsorption and Nano Filtration in Industrial Wastewater Treatment, Progress in Chemical and Biochemical Research. 4(1), 31-43.

89. El-Shahaby O., El-Zayat M., Rabei R., Aldesuquy H.S., 2019, Phytochemical constituents, antioxidant activity and antimicrobial potential of Pulicaria incisa (lam.) DC as a folk medicinal plant. Progress in Chemical and Biochemical Research. 2(4), 222-227. 90. El-Shahaby O.A., El-Zayat M., Abd El-Fattah, G., El-Hefny M.M., 2019. Evaluation of the biological activity of Capparis spinosa var. aegyptiaca essential oils and fatty constituents as Anticipated Antioxidant and Antimicrobial Agents. Progress in Chemical and Biochemical Research. 2(4), 211-221.

91. Hagr T., Adam I., 2020, Phytochemical Analysis, Antibacterial and antioxidant Activities of Essential Oil from *Hibiscus sabdariffa* (L.) Seeds, (Sudanese Karkadi). Progress in Chemical and Biochemical Research. 3(3), 194-201.

92. Thacker H., Ram V., Dave P.N., 2019. Plant mediated synthesis of Iron nanoparticles and their Applications: A Review. Progress in Chemical and Biochemical Research. 2(3), 84-91.

93. Fazeli-Nasab B., Khajeh H., Rahmani A.F., 2021. Effects of culture medium and plant hormones in organogenesis in olive (CV. Kroneiki). J Plant Bioinform Biotech. 1(1), 1-13.

94. Mursu J., Robien K., Harnack L.J., Park K., Jacobs D.R., 2011, Dietary supplements and mortality rate in older women: the Iowa Women's Health Study. Archives of Internal Medicine. 10;171(18), 1625-33.

95. MacLennan R., Macrae F., Bain C., Battistutta D., Chapuis P., Gratten H., Lambert J., Newland R.C., Ngu M., Russell A., Ward M., 1995. Randomized trial of intake of fat, fiber, and beta carotene to prevent colorectal adenomas. JNCI Journal of the National Cancer Institute. 6;87(23), 1760-6.

96. Terwanger Philip T., Asemave K., Obochi G.O., 2021. Comparative Assessment of Phytochemicals in Four (4) Varieties of *Ananas Comosus* (L.) Merr Peels. Progress in Chemical and Biochemical Research. 4(1), 1-10.

97. Asemave K., Anure T., 2019. The bioactivities of the neem (seeds and leaves) against Callosobruchus

maculatus on a *Vigna Subterranean* L. Progress in Chemical and Biochemical Research. 2(3), 92-98.