



REVIEW ARTICLE

Potential Harmful Effects of Aflatoxin M1 in Milk and Milk Products and Novel Methods to Reduction of Aflatoxin M1: A Review

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ABSTRACT: Dairy products are rich sources of vitamins, proteins and calcium that are vital to the human body. Aflatoxin M1 is a hydroxylated metabolite of aflatoxin B1. The presence of this toxin in milk has caused serious concern among nutritionists. Consumption of aflatoxin-contaminated milk (AFM1) may lead to serious health problems in humans. AFM1 causes various cancers such as liver cancer, damage to the nervous and DNA, as well as mutagenicity and teratogenicity. In reducing the amount of aflatoxin in the product, with does not affect the milk quality is of particular importance. Absorption and restructuring methods such as yeast, lactic acid (bacteria), enzymes, ozon, and cold plasma have been used for this purpose. In this study, they have been studied on aflatoxin M1 and toxicology besides reducing method for aflatoxin M1.

INTRODUCTION

Mycotoxins are substances that are produced naturally by poisonous fungi that grow on a number of agricultural products and are often responsible for human and animal health. Hundreds of know types of mycotoxins, aflatoxins (AF) are the most studied and the most public health concern worldwide [1]. Food contamination with mycotoxins (aflatoxin) is a particular importance and the World Health Organization [2], Relevant international organizations (FAO and Codex) have set the maximum

amount of aflatoxin contamination for different foods [3]. Aflatoxins are products of *Aspergillus* species including *A. flavus*, *A. parasiticus* and *A. nomius*. Aflatoxins G1, G2, B1 and B2 are the most important aflatoxins. AFM1 has been reported worldwide in dairy products [4]. When aflatoxin-contaminated feed is ingested with highly variable levels of AFB. However, milk has been shown to have the greatest potential for introducing AFM1 into the human diet [5, 6]. In terms of toxicity, aflatoxin M1 is

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approximately ten times less toxic than aflatoxin B1, but AFM1 is exposed to diet consumption of milk and dairy products can cause human disease such as liver cancer [7, 8]. Studies show that AFM1 is relatively stable during pasteurization. In anticipation of potential hazards, several national and international regulators have set a maximum AFM1 limit for milk and dairy products. AFM1 in milk should not exceed $0.05 \mu\text{g kg}^{-1}$ for adults and $0.025 \mu\text{g kg}^{-1}$ for foods used for infants and young children, while the amount of AFM1 in milk has been reported by the Codex Commission $0.05 \mu\text{g kg}^{-1}$ and by the FDA $0.5 \mu\text{g kg}^{-1}$ [9,10]. The national limit of AFM1 milk in Iran is $0.1 \mu\text{g kg}^{-1}$ [10].

Studies show that the existence of AFM1 in milk is a public health solicitude because milk plays a very important role in human diets across countries [11]. Aflatoxin causes poisoning (acute and chronic) and cancers in humans. The results indicated that aflatoxin poisoning varies with age, sex, and condition, and symptoms such as central nervous system, immunity system disorders, liver and kidney, brain injury, and death [12,13]. Given that milk is a valuable and ideal food for all age groups and is rich in essential nutrients, vitamins and amino acids and plays an important role in human health, so ensuring the quality of milk is very important. In recent years, the presence of dangerous and toxic chemicals in food, especially in milk, has increased. In this review, we discuss the presence of aflatoxin M1 in dairy products (milk) and its adverse effects on human health as well as new reducing methods that assessed in recent years.

Aflatoxin M1 in dairy products (milk and milk products)

AFM1 appearance in the milk 48-72 hours after the ingestion of AFB1. In one study, adding 1.7 to 2 micrograms of aflatoxin to milk and production of cheese, it was observed that 40% of aflatoxin remained in cheese and 60% in whey [15]. Studies show that about 0.3% - 6.2% of AFM1 can be transferred to milk. The amount of infection depends on the season, winter and genetic conditions of the animal [16-18]. Almost all samples of dry matter, baby food and yogurt in Italy and Kuwait are

contaminated with aflatoxin M1, which causes health problems for Italians [15]. Detection of AFM1 derivative in milk is from 12 to 24 hours after AFB1 consumption, but there is a decrease in concentration after 72 hours of contact [19, 20]. Measurement of AFM1 in Turkey showed that high-fat cheeses containing AFM1 exceeded the standard in Turkey [21]. Researchers show that only 33% of Spanish raw milk samples contain contamination [15]. A study (in Brazil) showed that aflatoxin M1 content has been $24\text{--}50 \text{ ng l}^{-1}$ [22]. The AFM1 in raw milk samples (cow) on northern, northeastern and western China was $69 \pm 52 \text{ ng kg}^{-1}$ [23]. The level of AFM1 in milk in Iran from 0.03 for Hamadan to 1293.29 for Kermanshah and from 0.03 for East Azerbaijan to 66.00 ng kg^{-1} for Qazvin were, respectively [6]. The average estimated level of AFM1 in Italian milk samples using valid ELISA method was about 35 ng kg^{-1} . Studies show that the average level of AFM1 in the milk of Middle Eastern countries was low. Meanwhile, the results of one study showed that the average level of AFM1 in Iranian raw milk (55.74 ng kg^{-1}), pasteurized (47.02 ng kg^{-1}), UHT milk (93.61 ng kg^{-1}) [24]. Dairy products made from raw milk contaminated with AFM1, so contain AFM1. Mixing bulk milk consignments with different levels of AFM1 contamination together is the only current process applied to reduce AFM1 content in milk [25]. The mean concentration of AFM1 in raw milk samples of Alborz province was from 0.0024 to $0.231 \mu\text{g kg}^{-1}$, the mean concentration $0.027 \pm 0.018 \mu\text{g kg}^{-1}$. According to FAO, in 2002 the annual milk consumption in Iran was 26.5 kg. Meanwhile, in the years 2010 to 2017, due to advertising and educational activities, the per capita annual consumption increased by 90 to 100 kg per person. Finally, for reasons such as the increase in dairy prices, the annual per capita consumption was reported to be 66.12 kg per Iranian. While the per capita annual milk consumption in the United States and Europe was reported to be 300-400 kg per person [12]. All these results indicate the importance of paying attention to milk and its products and aflatoxin contamination. With the advancement of the world and the acquisition of new scientific methods to identify and reduce food toxins, more global attention should be paid to this issue. Because milk, in addition to being the main food of

many sections of society, has also become the raw ingredient for other process products.

Effects of AFM1 on human health

Exposure to AFM1 is problematic for health on human and animal. The most commonly used methods recently are ELISA and fluorescence detection (FLD) liquid chromatography (LLD) [26-28]. However, other methods include thin layer chromatography (TLC), lateral flow immunoassay, and gel-based safety assays. TLC is an old technique and using HPLC with FLD is most effective. It is widely used in the diagnosis of AFM1 these days [29-31]. Daily AFM1 intake, as estimated by JECFA for Africa, the Middle East, Latin America, Europe, and the Far East, was 0.002 ng kg⁻¹ bw/day, 0.1, 0.058, 0.11, and 0.20, respectively [3]. Therefore, dairy foods are particularly

vulnerable to contamination by AFM1 and there are certain risks to human health [32]. Acute aflatoxin poisoning can have serious effects on human health, including as different types of kidney and liver disturbance. Studies show that people with hepatitis B are 30 times more likely to develop liver cancer, while in chronic poisoning leading to depression, neurodegenerative diseases and deficiency of vitamins A and D. Studies show that aflatoxicosis causes poisoning, disease and death in humans and animals [32]. Numerous studies on living creatures exposed to aflatoxin have shown DNA damage, with AFM1 being the destructive agent Figure 1. Therefore, acute and chronic toxicity research on living creatures, it is obvious that AFM1 is carcinogenic to living creatures and is classified in group B2 as carcinogenic according to the IARC [12].

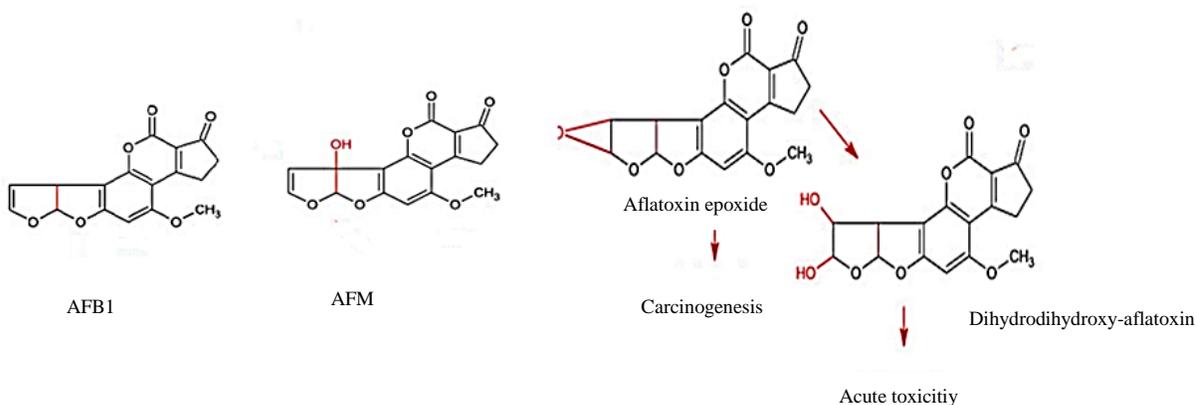


Figure 1. Aflatoxin M1 and its toxicity ways

Reduction of aflatoxin M1

Most countries have strict regulations for aflatoxins in milk, which vary from country to country. While the maximum residual rate (MRL) of AFM1 in dairy products is presented in Table 1. There are different ways to control aflatoxin levels, such as traditional methods of keeping

feed dry to prevent the growth of toxin-producing fungi. However, humidity may be present in some countries based on climatic conditions. Therefore, it is necessary to study and find alternative methods.

Table 1. Regulation of aflatoxin M1 in dairy products in some countries [29]

Country	product	MRL ($\mu\text{g L}^{-1}$)
USA	Milk and milk products	0.5
	cheese	0.25
EU	milk	0.05
	infants and young children	0.25
France	children <3 years	0.03
	pasteurized infant milk	0.01
Austria	butter	0.020
	cheese	0.25
	milk powder	0.4
Iran	milk	0.1
	milk powder	0.5
	butter and butter	0.02
	milk	0.2
Turkey and Switzerland	milk	0.05
	cheese	0.25

Aflatoxin reduction occurs in absorption or restructuring ways Figure 2. AFM1 was reduced by up to 100% after 60 min of treatment using *S. cerevisiae* in combination with lactic acid bacteria. *S. cerevisiae* alone reduced AFM1 by 90.3% during 30 minutes and 92.7%[33]. Heat-killed cells are used to prevent fermentation, which may affect the binding capacity of the yeast cell wall and also increase the ability of the microorganism to change its cell wall

structure [33]. The use of biological filters for *S. cerevisiae* immobilized on perlite is also used [34]. In this study, the initial level of 0.23 ppb AFM1 decreased by 81.3% at 80 min. The effect of different levels of AFM1 on the binding potential of microorganisms was reduced by 0.1% AFM1 in milk (0.05 $\mu\text{g/L}$) using 0.1 $\mu\text{g l}^{-1}$ *S. cerevisiae* alone or in combination with lactic acid bacteria [34].

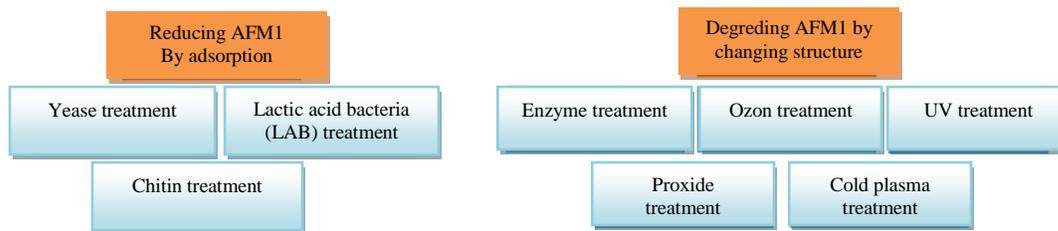


Figure 2. Reduction methods of Aflatoxin M1.

Chitin was used to bind AFM1 in milk [35]. Chitin binding capacity depends on the time of treatment, the level of chitin used, and the level of AFM1 contamination in the milk. The longer (24 hr) the binding between chitin and AFM1 was more stable compared to 30 min. Also, reducing the level of contamination, long duration and high level of chitin by using and low level of contamination of AFM1 increases the binding capacity of chitin.

The reduction of AFM1 in milk by LAB when using one strain was less than 90% [36, 37]. In other environments, such as yogurt, the binding capacity of AFM1 and lactic acid bacteria, which are also used as starters, include *L. delbrueckii* and *S. thermophilus* are less common [38]. Aflatoxin binding decreased by approximately 9% after yogurt production, which is probably related to the pH of the acid. However, Pierides et al. [39] It has been found that there is an increase in the binding ability of phosphate buffered saline at the acid pH level.

The ability of non-living microorganisms to bind has been more adaptable than that of living organisms. Therefore, to reduce the effects of fermentation, the use of heat in milk can be useful for deactivation, as has been done in yeasts [40]. In this regard, Kuharic et al. [41] isolated a higher percentage of AFM1 compared to living cells by examining non-living cells of *Lactobacillus plantarum*.

The link between aflatoxins and microbes is said to be weak, which is the reason for the connection. Although the binding mechanism is not fully understood, the binding mechanism is hydrogen bonds and vanderwaals bonds. Polysaccharides and peptidoglycans (Bacterial cell wall) are responsible for the uptake of AFM1 [42]. Different strains of bacteria have different ability to absorb aflatoxins due to changes in cell wall tissue [39,43].

The ability of bacteria and yeasts to bind aflatoxins in the short term [34]. This method suggests as a potential method for controlling aflatoxins in milk. Investigating the ability of microorganisms and environmental factors that may increase or decrease the connection capacity. Binding between the bacterial/yeast cell wall and aflatoxins is associated with AFM1 as a reversible bond, which does not degrade aflatoxins [44,45]. So evaluating the taste of milk

after treatment will be necessary for future studies (Table 2).

The effect of enzymes on microbial species can be another way to control and of course the degradation of aflatoxin. In fact, instead of using the microorganisms, their enzymes are used for extraction. Degradation of aflatoxin using microbial extracts is said to depend on pH and temperature [46,47], and if destroyed, its mutagenicity is reduced [48]. In many studies, ozone has been used to reduce AFB1 in different food [49,50]. Ozone concentration, sample, time as well as moisture level inactivate aflatoxin by ozone [51]. Report the efficiency of this method has been around 50% [52]. Although ozone treatment for 5 min at 16 mg/l reduced AFM1 in milk containing 0.56 µg/kg by 50%, but it did decrease in β-carotene content and total microbial count have seen [52].

Formation of aldehydes, organic acids, and ketones are other reactions that may occur [53]. Monozone derivatives through ozone treatment need to evaluate their effect on human health. This treatment causes reactions such as bonding between the furan and ozone. This may also be associated with reducing toxins and thus reduce the carcinogenicity and teratogenicity of aflatoxin [54].

Peroxide treatments are effective for inactivating AFM1 at a level of 0.7-1.7 µg L⁻¹ in naturally contaminated raw milk of hydrogen peroxide (H₂O₂), riboflavin (Rib) or H₂O₂ and lactoperoxidase (LOP). The effectiveness of these treatments was up to 100%, but using too much H₂O₂ in food can be a risk to human health. Excessive consumption of hydrogen peroxide can cause serious gastrointestinal problems [55]. Food and Drug Administration (FDA) regulations for H₂O₂ 0.5 mg L⁻¹ in processed food and 0.05% by weight of milk [56]. Using ultraviolet light (UV) is another way to reduce aflatoxin. The use of high-intensity ultraviolet radiation can alter the organoleptic properties of milk in taste [34].

Cold plasma, argon, nitrogen, nitrogen containing oxygen, helium and air can also be used to degrade aflatoxins [27]. AFB1 degradation is believed to be caused by cold plasma OH radicals, which are strong oxidizing agents, increase

the reaction between ozone and the aflatoxin eruption ring [57].

Mutagenicity and toxicity of aflatoxin due to olefin double bonding in the aflatoxin catalyzed by the epoxy group by cytochrome P450. The disappearance of this double bond can prevent the formation of AFB1-exo-8,9-epoxide [58, 59] ultimately lead to a reduction in aflatoxin toxicity. Changes in this process have resulted in changes in the concentrations of butyric acid, caprylic acid [60] and stearic acid [34]. Concentrations of milk fatty acids, ketones

and alcohols did not change significantly while an increase in total aldehyde composition was observed [61]. Aldehyde content has also been reported to increase with increasing cold plasma exposure time [62]. Increased aldehydes can be associated with increased residual reactions. Studies have shown that medicinal plants have active compounds of medicinal and antioxidant substances can eliminate contaminants in food [63, 64] and have therapeutic effects in humans [65-74].

Table 2. Bacterial strains used to reduce Aflatoxin M1 in milk [33, 35-37, 43]

Species	Sample	Contamination level (ppb) of aflatoxin M1	Temperature (°C) and time	Reduction (%)
<i>L. plantarum</i>				
<i>L. rhamnosus</i>				
<i>L. plantarum</i>	Skim milk/PBS	10	37°C- 24 h	4.13 –64.16
<i>L. plantarum</i>				
<i>L. plantarum</i>				
<i>L. plantarum</i>				
<i>B. animalis</i>	Yoghurt/PBS	0.1	42°C – 4h	49 -60
<i>B. bifidum</i>				
<i>L. plantarum</i>				
<i>B. pumilus</i>	-	40	37°C- 12 h	89.55
<i>L. Plantarum</i>				
<i>L. acidophilus</i>				
<i>B. bifidum</i>	Milk	50	37°C- 12 h	80 - 90
<i>Kl. lactis</i>				
<i>S. cerevisiae</i>				
<i>L. rhamnosus</i>	-	50	37°C- 18 h	63
<i>L. helveticus</i>	Milk	0.1	-	85
<i>L.rhamnosus</i>	UHT			
<i>B. lactis</i>	skimmed	0.5	4°C- 15 min	13-37
<i>L. bulgaricus</i>	milk			

CONCLUSIONS

AFM1 in dairy foods has adverse effects on alive creature (Human, infants and children) who consume large amounts of milk. Given that AFM1 has irreversible effects on human health and the highest transmission of AFM1 is through contaminated animal feed, it is concluded that to reduce the side effects of this dangerous toxin, the World

Agricultural Health Organization should provide the necessary training to provide livestock feed to ranchers. Among the proposed methods, some methods are not able to completely reduce AFM1 and may pose an additional risk to food safety. Peroxide treatment requires high doses of hydrogen peroxide, which may create residues that are of

concern to human health. Microbial enzymes degrade aflatoxins and no harmful effects have been observed. But it is not used directly in milk and could be a new study.

CONFLICT OF INTERESTS

The authors declare no conflict of interest

REFERENCES

1. Milićević D.R., Spirić D., Radičević T., Velebit B., Stefanović S., Milojević L., Janković S., 2017. A review of the current situation of aflatoxin M1 in cow's milk in Serbia: risk assessment and regulatory aspects. *Food Addit Contaminants: Part A*, 34, 1617-1631.
2. WHO, 2002. Evaluation of certain mycotoxins in food: fifty-sixth report of the Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain mycotoxins in food: fifty-sixth report of the Joint FAO/WHO expert committee on food additives. <https://apps.who.int/iris/handle/10665/42448>
3. Joint F., 2002. Evaluation of certain mycotoxins in food: fifty-sixth report of the Joint FAO/WHO Expert Committee on Food Additives, World Health Organ Tech Rep Ser. 2002;906:i-viii, 1-62.
4. Assi S.H.A., 2019. Determination of Aflatoxin M1 Levels in Milk and Milk-based Products in Gaza Strip, Palestine. *Al-Azhar University-Gaza*, 1-45.
5. Fallah A.A., Fazlollahi R., Emami A., 2016. Seasonal study of aflatoxin M1 contamination in milk of four dairy species in Yazd, Iran. *Food Cont.* 68, 77-82.
6. Khaneghahi Abyaneh H., Bahonar A., Noori N., Yazdanpanah H., Shojaee Aliabadi M.H., 2019. Aflatoxin M1 in raw, pasteurized and UHT milk marketed in Iran. *Food Addit Contaminants: Part B*, 12, 236-244.
7. Ruangwises N., Ruangwises S., 2010. Aflatoxin M1 contamination in raw milk within the central region of Thailand. *Bulletin Environ Contamination Toxicol.* 85, 195-198.
8. Anttila A., Bhat R V., Bond J.A., Borghoff S J., Bosch F.X., Carlson G.P., Castegnaro M., Cruzan G., Gelderblom W. C., Hass U., 2002. IARC monographs on the evaluation of carcinogenic risks to humans: Some traditional herbal

medicines, some mycotoxins, naphthalene and styrene. IARC monographs on the evaluation of carcinogenic risks to humans, 82.

9. Kamkar A., Jahed K G.R., Alavi S., 2011. Occurrence of aflatoxin M1 in raw milk produced in Ardebil of Iran. *Iranian J Environm Health Sci Engin.* 8(2), 123-128
10. Jaiswal P., Jha S.N., Kaur J., Borah A., Ramya H., 2018. Detection of aflatoxin M1 in milk using spectroscopy and multivariate analyses. *Food Chem.* 238, 209-214.
11. Bukari N., Kwofie M.K., Adeboye O., 2020. Aflatoxin M1 (*Aspergillus parasiticus, flavus*) Occurrences in Milk and Milk Products and Its Possible Health Effects. *Adv Microbiol.* 10, 509.
12. Pardakhti A., Maleki S., 2019. Risk assessment of Aflatoxin M1 contamination of milk in Iran. *Int J Environmental Res.* 13, 265-271.
13. Li H, Xing L., Zhang M., Wang J., Zheng N., 2018. The toxic effects of aflatoxin B1 and aflatoxin M1 on kidney through regulating L-proline and downstream apoptosis. *Bio Med Res Int.* 12, 9074861.
14. Duarte S., Almeida A., Teixeira A., Pereira A., Falcão A., Pena A., Lino C., 2013. Aflatoxin M1 in marketed milk in Portugal: Assessment of human and animal exposure. *Food Cont.* 30, 411-417.
15. Salari N., Kazeminia M., Vaisi-Raygani A., Jalali R., Mohammadi M., 2020. Aflatoxin M1 in Milk Worldwide from 1988 to 2020: A Systematic Review and Meta-Analysis. *J Food Quality.* Article ID 8862738, 14 pages <https://doi.org/10.1155/2020/8862738>
16. Palizban M., Jahed G., Pakbin B., 2016. Aflatoxin M1 contamination of cow's raw milk in different seasons from Qazvin province, Iran. *J Biol Today's World.* 5, 173-176.
17. Akbar N., Nasir M., Naeem N., Ahmad M.U.D., Iqbal S., Rashid A., Imran M., Aslam Gondal T., Atif M., Salehi B., 2019. Occurrence And Seasonal Variations of Aflatoxin M1 In Milk From Punjab, Pakistan. *Toxins.* 11, 573-574.
18. Panariti E., 2001. Seasonal variations of aflatoxin M1 in the farm milk in Albania. *Arhiv za Higijenu Rada Toksiko.* 52, 37-41.
19. Yosef T., Al-Julaifi M., Salah-El-Dein W., Al-Rizqi A., 2013. Assessment of aflatoxin M1 residues in raw cow

milk at Al-Riyadh area with reference to some detoxification applications. *Life Sci J.* 10(1), 4365-4369.

20. Fallah A.A., Rahnama M., Jafari T., Saei-Dehkordi S.S., 2011. Seasonal variation of aflatoxin M1 contamination in industrial and traditional Iranian dairy products. *Food Cont.* 22, 1653-1656.

21. Oruc H., Sonal T., 2001. Determination of aflatoxin M1 levels in cheese and milk consumed in Bursa, Turkey. *Vet Human Toxicol.* 43, 292-293.

22. Garrido N., Iha M., Santos Ortolani M., Duarte Favaro R., 2003. Occurrence of aflatoxins M1 and M2 in milk commercialized in Ribeirão Preto-SP, Brazil. *Food Addit Contaminants.* 20, 70-73.

23. Bilandžić N., Varenina I., Kolanović B.S., Božić Đ., Đokić M., Sedak M., Tanković S., Potočnjak D., Cvetnić Ž., 2015. Monitoring of aflatoxin M1 in raw milk during four seasons in Croatia. *Food Cont.* 54, 331-337.

24. Rahmani J., Alipour S., Miri A., Fakhri Y., Riahi S.M., Keramati H., Moradi M., Amanidaz N., Pouya R.H., Bahmani Z., 2018. The prevalence of aflatoxin M1 in milk of Middle East region: a systematic review, meta-analysis and probabilistic health risk assessment. *Food Chem Toxicol.* 118, 653-666.

25. Sarvar Taherabadi M., Gharavi M.J., Javadi I., Alimohammadi M., Moghadamnia S.H., Mosleh N.M., Farajollahi M., Sharif M., 2016. The level of Aflatoxin M1 in raw and pasteurized milk produced in Alborz Province, Iran. *Jundishapur J Nat Pharm Prod.* 11 (4), 31708.

26. Liu B.H., Chu K.C., Yu F.Y., 2016. Novel monoclonal antibody-based sensitive enzyme-linked immunosorbent assay and rapid immunochromatographic strip for detecting aflatoxin M1 in milk. *Food Cont.* 66, 1-7.

27. Wang J.J., Liu B.H., Hsu Y.T., Yu F.Y., 2011. Sensitive competitive direct enzyme-linked immunosorbent assay and gold nanoparticle immunochromatographic strip for detecting aflatoxin M1 in milk. *Food Cont.* 22, 964-969.

28. Lee D., Lee K.G., 2015. Analysis of aflatoxin M1 and M2 in commercial dairy products using high-performance liquid chromatography with a fluorescence detector. *Food Cont.* 50, 467-471.

29. Iqbal S.Z., Jinap S., Pirouz A., Faizal A.A., 2015. Aflatoxin M1 in milk and dairy products, occurrence and

recent challenges: A review. *Trends Food Sci Technol.* 46, 110-119.

30. Carvalho K., Goncalves G., Lopes A., Santos E., Vargas E., Magalhães W., 2012. Modelling uncertainty estimation for the determination of aflatoxin M1 in milk by visual and densitometric thin-layer chromatography with immunoaffinity column clean-up. *Food Addit Contaminants: Part A.* 29, 679-693.

31. OSSA D.E.H., Hincapie D.A., Peñuela G.A., 2015. Determination of aflatoxin M1 in ice cream samples using immunoaffinity columns and ultra-high performance liquid chromatography coupled to tandem mass spectrometry. *Food Cont.* 56, 34-40.

32. Pour S.H., Mahmoudi S., Masoumi S., Rezaie S., Barac A., Ranjbaran M., Oliya S., Mehravar F., Sasani E., Noorbakhsh F., 2020. Aflatoxin M1 contamination level in Iranian milk and dairy products: A systematic review and meta-analysis. *World Mycotoxin J.* 13, 67-82.

33. Corassin C., Bovo F., Rosim R., Oliveira C., 2013. Efficiency of *Saccharomyces cerevisiae* and lactic acid bacteria strains to bind aflatoxin M1 in UHT skim milk. *Food Cont.* 31, 80-83.

34. Nguyen T., Flint S., Palmer J., 2020. Control of aflatoxin M1 in milk by novel methods: A review. *Food Chem.* 311, 125984.

35. Assaf J.C., Atoui A., Khoury A.E., Chokr A., Louka N., 2018. A comparative study of procedures for binding of aflatoxin M1 to *Lactobacillus rhamnosus* GG. *Brazilian J Microbiol.* 49, 120-127.

36. Abdelmotilib N.M., Hamad G.M., Elderea H.B., Salem E.G., Sohaimy S.A., 2018. Aflatoxin M1 reduction in milk by a novel combination of probiotic bacterial and yeast strains. *Eur J Nutr Food Safety.* 83-99.

37. Gu X., Sun J., Cui Y., Wang X., Sang Y., 2019. Biological degradation of aflatoxin M1 by *Bacillus pumilus* E-1-1-1. *Microbiol Open.* 8, 00663.

38. Sarimehmetoğlu B., Kuplulu Ö., 2004. Binding Ability of Aflatoxin M1 To Yoghurt Bacteria. *Ankara Üniv Vet Fak Derg.* 51, 195-198.

39. Pierides M., El-Nezami H., Peltonen K., Salminen S., Ahokas J., 2000. Ability of dairy strains of lactic acid

- bacteria to bind aflatoxin M1 in a food model. *J Food Prot.* 63, 645-650.
40. Elsanhoty R.M., Salam S.A., Ramadan M.F., Badr F.H., 2014. Detoxification of aflatoxin M1 in yoghurt using probiotics and lactic acid bacteria. *Food Contr.* 43, 129-134.
41. Kuharić Ž., Jakopović Ž., Čanak, I., Frece, J., Bošnjir, J., Pavlek, Ž., Ivešić, M., Markov K., 2018. Removing aflatoxin M1 from milk with native lactic acid bacteria, centrifugation, and filtration. *Arhiv za Higijenu Rada Toksikolo.* 69, 334-339.
42. Shetty P.H., Jespersen L., 2006. *Saccharomyces cerevisiae* and lactic acid bacteria as potential mycotoxin decontaminating agents. *Trends Food Sci Technol.* 17, 48-55.
43. Ismail A., Levin R.E., Riaz M., Akhtar S., Gong Y.Y., De Oliveira C.A., 2017. Effect of different microbial concentrations on binding of aflatoxin M1 and stability testing. *Food Contr.* 73, 492-496.
44. Guan S., Zhao L., Ma Q., Zhou T., Wang N., Hu X., Ji C., 2010. In vitro efficacy of *Myxococcus fulvus* ANSM068 to biotransform aflatoxin B1. *Int J Molec Sci.* 11, 4063-4079.
45. Rao K.R., Vipin A., Hariprasad P., Appaiah K.A., Venkateswaran G., 2017. Biological detoxification of Aflatoxin B1 by *Bacillus licheniformis* CFR1. *Food Control.* 71, 234-241.
46. Zhao L., Guan S., Gao X., Ma Q., Lei Y., Bai X., Ji C., 2011. Preparation, purification and characteristics of an aflatoxin degradation enzyme from *Myxococcus fulvus* ANSM068. *J Appl Microbiol.* 110, 147-155.
47. Motomura M., Toyomasu T., Mizuno K., Shinozawa T., 2003. Purification and characterization of an aflatoxin degradation enzyme from *Pleurotus ostreatus*. *Microbiolog Res.* 158, 237-242.
48. Adebo O.A., Njobeh P.B., Mavumengwana V., 2016. Degradation and detoxification of Afb1 by *Staphylococcus warneri*, *Sporosarcina* Sp. and *Lysinibacillus fusiformis*. *Food Contr.* 68, 92-96.
49. Akbas M.Y., Ozdemir M., 2006. Effect of different ozone treatments on aflatoxin degradation and physicochemical properties of pistachios. *J Sci Food Agri.* 86, 2099-2104.
50. Luo X., Wang R., Wang L., Li Y., Bian Y., Chen Z., 2014. Effect of ozone treatment on aflatoxin B1 and safety evaluation of ozonized corn. *Food Contr.* 37, 171-176.
51. Agriopoulou S., Koliadima A., Karaiskakis G., Kapolos J., 2016. Kinetic study of aflatoxins' degradation in the presence of ozone. *Food Contr.* 61, 221-226.
52. De Jesus Benevides C.M., Da Cunha Veloso M.C., De Paula Pereira P.A., De Andrade J.B., 2011. A chemical study of β -carotene oxidation by ozone in an organic model system and the identification of the resulting products. *Food Chem.* 126, 927-934.
53. Diao E., Hou H., Chen B., Shan C., Dong H., 2013. Ozonolysis efficiency and safety evaluation of aflatoxin B1 in peanuts. *Food Chem Toxicol.* 55, 519-525.
54. De Alencar E.R., Faroni L.R.D.A., Soares N.D.F.F., Da Silva W.A., Da Silva Carvalho M.C., 2012. Efficacy of ozone as a fungicidal and detoxifying agent of aflatoxins in peanuts. *J Sci Food Agri.* 92, 899-905.
55. Silva R.A., Montes R.H., Richter E.M., Munoz R.A., 2012. Rapid and selective determination of hydrogen peroxide residues in milk by batch injection analysis with amperometric detection. *Food Chem.* 133, 200-204.
56. Özkan M., Kirca A., Cemerolu B., 2004. Effects of hydrogen peroxide on the stability of ascorbic acid during storage in various fruit juices. *Food Chem.* 88, 591-597.
57. Diao E., Hou H., Dong H., 2013. Ozonolysis mechanism and influencing factors of aflatoxin B1: A review. *Trends Food Sci Technol.* 33, 21-26.
58. Diaz G.J.M., 2011. Biotransformation of Aflatoxin B1 and Its Relationship with the Differential Toxicological Response to Aflatoxin in Commercial Poultry Species. In *Aflatoxin. Biochem Molecular Biol.* 11(3), 4-23.
59. Samuel M.S., Sivaramakrishna A., Mehta A., 2014. Degradation and detoxification of aflatoxin B1 by *Pseudomonas putida*. *Int Biodeterio Biodegrad.* 86, 202-209.
60. Kim H.J., Yong H.I., Park S., Kim K., Choe W., Jo C., 2015. Microbial safety and quality attributes of milk following treatment with atmospheric pressure

- encapsulated dielectric barrier discharge plasma. Food Contr. 47, 451-456.
61. Korachi M., Ozen F., Aslan N., Vannini L., Guerzoni M.E., Gottardi D., Ekinici F.Y., 2015. Biochemical changes to milk following treatment by a novel, cold atmospheric plasma system. *Int Dairy J.* 42, 64-69.
62. Ragni L., Berardinelli A., Vannini L., Montanari C., Sirri F., Guerzoni M.E., Guarnieri A., 2010. Non-thermal atmospheric gas plasma device for surface decontamination of shell eggs. *J Food Engin.* 100, 125-132.
63. Pirhadi M., Shariatifar N., Bahmani M., Manouchehri A.A., 2021. Heavy metals in wheat grain and its impact on human health: A review. *J Chem Health Risks.* 10.22034/jchr.2021.1924307.1269
64. Bahmani M., 2019. A new method for promoting biologic synthesis and reducing the size of titanium dioxide nanoparticles (TiO₂ NPs) synthesized by *Origanum vulgare*. *Plant Biotechnol Persa.* 1(1), 10-12.
65. Farzan B., Shahsavari S., Abbaszadeh S., Teimouri H., 2019. Phytotherapy for seizure: An overview of the most important indigenous Iranian medicinal plants with anticonvulsant properties. *Plant Sci Today.* 6(4), 367-372.
66. Manouchehri A., Shakib P., Biglaryan F., Nazer M., Darvishi M., 2021. The most important medicinal plants affecting bee stings: A systematic review study. *Uludag Aricilik Dergisi.* 21(1), 91-103.
67. Farzan B., Abbaszadeh S., Teimouri H., 2019. Ethnobotanical treatments for earache and sore throat. *Int J Res Pharma Sci.* 10(2), 1354-1360.
68. Esmaili A., Parsaei P., Nazer M.R., Bakhtiari R., Mirbehresi H., Safian Boldaji H., 2021. Phytotherapy in Burn Wound Healing: A Review of Native Iranian Medicinal Plants. *J Chem Health Risks.* 11(0), doi: 10.22034/jchr.2021.1932188.1322
69. Farzan B., Abbaszadeh S., Basati G., Teimouri H., 2019. An overview of the most important medicinal plants effective on the strength of memory and mind in Iranian ethnobotany. *J Pharmacy Pharmacog Res.* 7(3), 156-162.
70. Abbasi N., Khalighi Z., Eftekhari Z., Bahmani M., 2020. Extraction and phytoanalysis of chemical compounds of *Eucalyptus globulus* leaf native to Dehloran, Ilam province, Iran by HS-SPME and GC-MS. *Adv Animal Vet Sci.* 8(6), 647-652.
71. Aidy A., Karimi E., Ghaneialvar H., Mohammadpour S., Abbasi N., 2020. Protective effect of *Nectaroscordum tripedale* extract and its bioactive component tetramethylpyrazine against acetaminophen-induced hepatotoxicity in rats. *Adv Trad Med.* 20(3), 471-477.
72. Karimi E., Abbasi S., Abbasi N., 2019. Thymol polymeric nanoparticle synthesis and its effects on the toxicity of high glucose on OEC cells: Involvement of growth factors and integrin-linked kinase. *Drug Des, Develop Thera.* 13, 2513-2532.
73. Abbasi N., Khosravi A., Aidy A., Shafiei M., 2016. Biphasic response to luteolin in MG-63 osteoblast-like cells under high glucose-induced oxidative stress. *Iran J Med Sci.* 41(2), 118-125.
74. Nouri A., Heidarian E, Amini-Khoei H, Abbaszadeh S., Basati G., 2019. Quercetin through mitigation of inflammatory response and oxidative stress exerts protective effects in rat model of diclofenac-induced liver toxicity. *J Pharmacy Pharmacog Res.* 7(3), 200-212.