

# Fe<sub>3</sub>O<sub>4</sub>-MNPs promoted green synthesis of thiazole derivatives using alkyl bromides

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Received: August 2021; Revised: September 2021; Accepted: October 2021

**Abstract:** In this research, magnetic iron oxide nanoparticles (Fe<sub>3</sub>O<sub>4</sub>-MNPs) were produced using green biosynthetic method by reduction of ferric chloride solution with *Clover Leaf* water extract. The nanoparticles generated using this procedure can potentially important in different purposes such as organic synthesis. A one-pot, efficient and high yielding procedure for the synthesis of thiazole derivatives is investigated. The procedure carried out *via* multicomponent reaction of isothiocyanate, alkyl bromides, *N*-methylthiazole and triphenylphosphine in the presence of bio-Fe<sub>3</sub>O<sub>4</sub>-MNPs as catalyst under solvent-free conditions at 50 °C. Easy, simple, rapid and clean procedure for the synthesis of thiazole derivatives are the advantages of this study.

Keywords: Isothiocyanate, Fe<sub>3</sub>O<sub>4</sub>-MNPs, Alkyl bromides, Green synthesis, Thiazole.

### Introduction

In multicomponent reactions (MCRs), three or more reactants combine in a one-pot and generate one product. These reactions are interesting in last decade of economically and environmentally because advantageous towards multi-step methods [1-7]. Multistep synthesis produce large amounts of waste due to separation steps of intermediate that commonly include employing of expensive, toxic and dangerous solvents in each step. MCRs are absolutely studied about combinatorial synthesis, and used in the discovery method for new drugs and agrochemicals [8]. MCRs have a great instrument toward the one-pot synthesis of different and complex compounds with small heterocycles.19approach to improve their synthetic utility. The nano catalyst display good catalytic activity compared to their bulk sized samples [9, 10] for the synthesis of some organic compounds. Nanoparticles are particles with very small sized along.

These compounds because of its large surface area to volume ratio have good catalytic reactivity, thermal conductivity, non-linear optical presentation and chemical steadiness [11]. Nanoparticles due to their antimicrobial activities [12] could be considered as nano antibiotics. Using plants with biomedical applications for biosynthesis of nanoparticles is a procedure of generating nanoparticles. This method is a cheap, bio friendly, safe, green procedure [13]. Green synthesis contains synthesis using plants, bacteria, fungi, algae etc. These methods investigated generation of bio-Fe3O4-MNPs with big scale without additional impurities [14]. Nanoparticles produced from plant show more catalytic activity without employing of expensive and toxic chemicals. These plants extract displaye some phytochemicals that play in both decreasing capping and stabilization agent. Currently, a great number of physical, chemical and biological procedure are available to generated different classes of nanoparticles [15]. The generated nanoparticles by using each methods display specific properties.

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Conversely, at this time biosynthesis of metal

Green nanotechnology has many important and involves a wide scale of procedures that decrease or remove toxic materials to restore the environment. Green chemistry methods are significant due to aim to protect resources and reducing costs [16–18]. In this research we investigate the synthesis of thiazole nanoparticles by using of plants is under expansion.

derivatives 5 via three-component reaction of thiazole 1, isothiocyanates 2, alkyl bromides 3 and triphenylphosphine 4 in the presence of catalytic amount of  $Fe_3O_4$ -MNPs nanoparticles under solvent-free conditions in good yield (Scheme 1).

N	+ R-N=C=S +	Br	R' + PPh <sub>3</sub>	Fe <sub>3</sub> O <sub>4</sub> -MNPs(	(15 mol%) $\sqrt{15 mol}$	R'
1	2	3	Ö	solvent-free,	30°C., 3 II S	н́ 5
		2, 3, 5	R	R'	Yield % of 5	
		a	Ph	CO <sub>2</sub> Et	92	
		b	4-MeO-C <sub>6</sub> H <sub>4</sub>	$CO_2Et$	95	
		с	4-Me-C <sub>6</sub> H <sub>4</sub>	$CO_2Et$	92	
		d	$4-NO_2-C_6H_4$	CO <sub>2</sub> Et	85	
		e	4-Br-C <sub>6</sub> H <sub>4</sub>	4-MeO-C <sub>6</sub> H <sub>4</sub>	90	
		f	4-Me-C6H4	4-MeO-C <sub>6</sub> H <sub>4</sub>	87	
		g	4-Me-C <sub>6</sub> H <sub>4</sub>	$4-NO_2-C_6H_4$	80	
		h	<sup>t</sup> Bu	CO <sub>2</sub> Et	97	

Scheme 1. Synthesis of compound 5.

Optimization the reaction conditions is the first step in these reactions. The reaction of *N*-methylthiazole 1, phenylisothiocyanate 2a, ethyl bromopyruvate 3a and triphenylphosphine 4 was selected as the model reaction (Scheme1). This reaction has low yield without catalyst and mixture of reaction is very busy. We employed some nano catalysts such as ZnO-NPs, TiO<sub>2</sub>-NPs, KF/CP NPs and CuO-NPs in the model reaction. Among them bio-Fe<sub>3</sub>O<sub>4</sub>-MNPs is the best because of green synthesis, easy separation of catalyst from the mixture of reaction and easy purification of product. Also several solvents and solvent-free conditions are employed for this reaction. The yield of reaction under solvent-free conditions and 50 °C is the good. The results of our optimization are showed in Table **1**.

Table 1. Optimization of reaction condition for the synthesis of compound 5a.

Entry	Catalyst	Temperature °C	Solvent	Yield, % of <b>5a</b>	
1	None	r.t.	None		
2	None	50	None	15	
3	KF/CP NPs	r.t.	$CH_2Cl_2$	45	
4	KF/CP NPs	50	CH <sub>3</sub> CN	55	
5	TiO <sub>2</sub> -NPs	50	CH <sub>3</sub> CN	65	
6	TiO <sub>2</sub> -NPs	50	None	68	
7	TiO <sub>2</sub> -NPs	80	Toluene	72	
8	Fe <sub>3</sub> O <sub>4</sub> -MNPs	r.t.	$CH_2Cl_2$	54	
9	Fe <sub>3</sub> O <sub>4</sub> -MNPs	50	CH <sub>3</sub> CN	87	
10	Fe <sub>3</sub> O <sub>4</sub> -MNPs	80	CH <sub>3</sub> CN	87	
11	Fe <sub>3</sub> O <sub>4</sub> -MNPs	50	None	92	
12	CuO-NPs	r.t.	$CH_2Cl_2$	35	

Bio-Fe<sub>3</sub>O<sub>4</sub>-MNPs were prepared by using the aqueous extract of *Clover Leaf*. For confirming the structure of bio-Fe<sub>3</sub>O<sub>4</sub>MNPs, field emission scanning electron microscopy (FESEM) (Figure 1) and XRD

(Figure 2) spectra are given for nanostructure. FESEM analysis of the magnetic nanoparticles showed uniform-sized particles with spherical morphology.



Figure 1. FESEM images of Fe<sub>3</sub>O<sub>4</sub>-MNPs.

Figure 2 shows the XRD spectra of the Fe<sub>3</sub>O<sub>4</sub>-MNPs. The diffraction peaks at  $2\theta = 29.7^{\circ}$ ,  $35.6^{\circ}$ ,  $42.9^{\circ}$ ,  $57.3^{\circ}$  and  $62.7^{\circ}$  can be indexed to (220), (311), (400), (511) and (440) planes of cubic Fe<sub>3</sub>O<sub>4</sub> (JCPDS 19-0629). The average size of the Fe<sub>3</sub>O<sub>4</sub>-MNPs is calculated 14 nm.



The UV-visible spectrum of the green synthesized  $Fe_3O_4$ -MNPs employing as a capping agent has been shown in Figure 3. This spectrum shows a broad bond at 365 nm which is characteristic of the Surface Plasmon Resonance (SPR) of  $Fe_3O_4$ -MNPs in solution.



Figure 3. UV-visible spectrum of Fe<sub>3</sub>O<sub>4</sub>-MNPs

Figure 4 shows the FT-IR spectra of the *Clover Leaf* extract and Fe<sub>3</sub>O<sub>4</sub> nanoparticles. The FT-IR spectrum of the extract (a) represented some peaks at 3500 to 3000, 1634, 1412, 1300 to 1100 cm<sup>-1</sup>. These peaks could be attributed to the functional groups of flavonoids and other phenolic compounds in the *Clover Leaf* extract that are answerable for the decrease of the metal ions and capping of the bio-decreased nanoparticles. The characteristic band of Fe–O in the FT-IR spectra of the Fe<sub>3</sub>O<sub>4</sub>-MNPs (b) is located at 588 cm<sup>-1</sup>.



Figure 4. FT-IR spectra of (a) Clover Leaf extract (b) Fe<sub>3</sub>O<sub>4</sub>-MNPs

The amount of catalyst is 15 mol% in optimized condition. We used 10-30 mol% catalyst for selecting best amount of catalyst. By increasing the amount of catalyst from 15 mol%, isn't seen any change in the yield of reaction. After each run, the catalyst was separated from the mixture of reaction and washed with ethylacetate, then dried under vacuum at room temperature.

The structure of compounds **5a-h** was confirmed by <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, IR and mass spectroscopy. For example, the <sup>1</sup>H-NMR of compound **5a** displays one singlet for NMe protons at 2.85 ppm, one doublet at 5.26 (d, <sup>3</sup>*J* = 6.8 Hz) ppm for -CH protone, three doublets for three =CH proton at 6.56 (d, <sup>3</sup>*J* = 6.5 Hz), 6.75 (d, <sup>3</sup>*J* = 6.5 Hz), 7.03 (d, <sup>3</sup>*J* = 6.8 Hz) ppm along with signals for aromatic moiety. The <sup>13</sup>C NMR spectra

of **5a** show resonance of C=N group and carbonyl at 157.2 (C=N), 165.3 (C=O) ppm respectively.

Although we have not confirmed the mechanism of the reaction in the presence of Fe<sub>3</sub>O<sub>4</sub>-MNPs, a possible examination is proposed in Scheme 2. In one pot the reaction of *N*-methylthiazole 1 and isothiocyanate 2 produce intermediate 6. In another pot, the reaction of  $\alpha$ -bromo ketones 3 and triphenylphosphine 4 produce intermediate 7 that in the presence of Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles (MNPs) is converted to intermediate 8 by elimination of HBr. The intermediate 6 react with intermediate 8 to produce zwitterionic species 9. Finally, intermolecular cyclization of intermediate 9 generates intermediate 10 that followed by elimination of triphenyl phosphin oxide produce compounds 5 (Scheme 2).



Scheme 2. Proposed mechanism for the synthesis of 5.

#### Conclusion

In summary, thiazole derivatives were prepared *via* multicomponent reaction of *N*-methylthiazole,

isothicyanates, alkyl bromides and triphenylphosphine in the presence of catalytic amounts of Fe<sub>3</sub>O<sub>4</sub>-MNPs under solvent-free condition at 50°C in good yield. These procedures have some advantages such as easy separation of product, green conditions, and reusability and easy separation of catalyst.

## Experimental

All chemicals used in this work were prepared from Fluka and were employed without further purification. IR spectra (KBr medium) were recorded on a Shimadzu IR-460 spectrometer. <sup>1</sup>H and <sup>13</sup>C NMR spectra were obtained on a Bruker DRX-500 AVANCE spectrometer at 500 and 125 MHz, respectively, in CDCl<sub>3</sub> using TMS as internal standard. Mass spectra (electron impact ionization) were recorded on a Finnigan MAT 8430 spectrometer operating at an ionization potential 70 eV. Elemental analyses were performed using a Heraeus CHN–O-Rapid analyzer.

## Preparation of Fe<sub>3</sub>O<sub>4</sub> MNPs:

Dried *Clover Leaf* (10gr) was drench in water (200 mL) at 80°C. After 2 h, the mixture was filtered and water essential oil was used for preparation of  $Fe_3O_4$ -MNPs as following.

100 ml water extract of *Clover Leaf* was taken in a 250 two-neck round bottom flask, and FeCl<sub>3</sub> (2 mmol), FeCl<sub>2</sub> (1 mmol) were added. Then, the solution of NH<sub>4</sub>OH (9 M, 10 ml) was then injected drop wise into the mixture with vigorous stirring under N<sub>2</sub> atmosphere for 1 h at room temperature. The resultant solution was a black color precipitate. The precipitate was separated by applying external magnetic field and washed with water for several times as well as dried in oven at 80 °C for 24 h.

#### Preparation of compounds 5a-h:

To a stirred mixture of *N*-methylthiazole **1** (2 mmol) mixed with isothiocyanate **2** (2 mmol) under solvent-free conditions was added mixture of alkyl bromides **3** (2 mmol) and triphenylphosphine **4** in the presence of Fe<sub>3</sub>O<sub>4</sub>-MNPs (15 mol%) after 45 min at 50 °C. After completion of the reaction (5 h; TLC control (hexane–AcOEt,1:3), 15 mL H2O was poured to mixture of reaction. Fe<sub>3</sub>O<sub>4</sub>-MNPs were separated by using external magnetic. After removing solvent, the residue was purified by column chromatography (3:1 hexane/EtOAc) to afforded pure title compounds.

# *Ethyl* 1-methyl-5-(phenylimino)-1,8a-dihydroimidazo [1,2-c][1,3]thiazine-7-carboxylate (5a):

Yellow oil; Yield: 0.58 g (92%). IR (KBr) ( $v_{max}$ /cm<sup>-1</sup>): 1735, 1694, 1525, 1454, 1337 and 1229 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  1.28 (3 H, t, <sup>3</sup>*J* = 7.3 Hz,

Me), 2.85 (3 H, s, NMe), 4.25 (2 H, q,  ${}^{3}J$  = 7.3 Hz, CH<sub>2</sub>O), 5.26 (1 H, d,  ${}^{3}J$  = 6.8 Hz, CH), 6.56 (1 H, d,  ${}^{3}J$  = 6.5 Hz, CH), 6.75 (1 H, d,  ${}^{3}J$  = 6.5 Hz, CH), 7.03 (1 H, d,  ${}^{3}J$  = 6.8 Hz, CH), 7.22 (2 H, t,  ${}^{3}J$  = 7.5 Hz, 2 CH), 7.34 (1 H, t,  ${}^{3}J$  = 7.5 Hz, CH), 7.43 (2 H, d,  ${}^{3}J$  = 7.5 Hz, 2 CH) ppm.  ${}^{13}$ C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  14.2 (Me), 38.3 (NMe), 62.4 (CH<sub>2</sub>O), 81.2 (CH), 107.2 (CH), 121.6 (CH), 124.3 (C), 125.3 (2 CH), 130.4 (2 CH), 131.2 (CH), 133.4 (CH), 152.4 (C), 157.2 (C=N), 165.3 (C=O) ppm. MS, m/z (%): 315 (M<sup>+</sup>, 15), 269 (82), 77 (68), 45 (100). Anal.Calcd for C<sub>16</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub>S (315.39): C, 60.93; H, 5.43; N, 13.32. Found: C, 61.12; H, 5.62; N, 13.54.

# *Ethyl* 1-methyl-5-(4-methoxyphenylimino)-1,8adihydroimidazo[1,2-c][1,3]thiazine-7-carboxylate (5b):

Yellow oil; Yield: 0.66 g (95%). IR (KBr) (v<sub>max</sub>/cm<sup>-</sup> <sup>1</sup>): 1737, 1695, 1547, 1478, 1357 and 1286 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  1.25 (3 H, t,  ${}^{3}J$  = 7.4 Hz, Me), 2.86 (3 H, s, NMe), 3.87 (3 H, s, MeO), 4.27 (2 H, q,  ${}^{3}J = 7.4$  Hz, CH<sub>2</sub>O), 5.26 (1 H, d,  ${}^{3}J = 6.3$  Hz, CH), 6.48 (1 H, d,  ${}^{3}J = 6.2$  Hz, CH), 6.67 (1 H, d,  ${}^{3}J =$ 6.2 Hz, CH), 6.95 (2 H, d,  ${}^{3}J = 7.6$  Hz, 2 CH), 7.14 (2 H, d,  ${}^{3}J = 7.6$  Hz, 2 CH), 7.18 (1 H, d,  ${}^{3}J = 6.3$  Hz, CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  13.8 (Me), 38.5 (NMe), 55.6 (MeO), 62.2 (CH<sub>2</sub>O), 81.5 (CH), 106.8 (CH), 114.5 (2 CH), 124.5 (C), 126.7 (2 CH), 130.5 (CH), 133.4 (CH), 145.2 (C), 155.3 (C), 157.5 (C=N), 165.7 (C=O) ppm. MS, m/z (%): 345 (M<sup>+</sup>, 10), 314 (84), 107 (88), 45 (100). Anal.Calcd for C<sub>17</sub>H<sub>19</sub>N<sub>3</sub>O<sub>3</sub>S (345.42): C, 59.11; H, 5.54; N, 12.17. Found: C, 59.25; H, 5.72; N, 12.32.

## *Ethyl* 1-methyl-5-(4-methylphenylimino)-1,8adihydroimidazo[1,2-c][1,3]-7-carboxylate (5c):

Yellow oil; Yield: 0.61 g (92%). IR (KBr) ( $v_{max}$ /cm<sup>-1</sup>): 1735, 1698, 1562, 1486, 1378 and 1295 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  1.32 (3 H, t, <sup>3</sup>*J* = 7.4 Hz, Me), 2.23 (3 H, s, Me), 2.95 (3 H, s, NMe), 4.26 (2 H, q, <sup>3</sup>*J* = 7.4 Hz, CH<sub>2</sub>O), 5.23 (1 H, d, <sup>3</sup>*J* = 5.8 Hz, CH), 6.63 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.73 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.73 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.74 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 7.15 (2 H, d, <sup>3</sup>*J* = 7.6 Hz, 2 CH), 7.18 (1 H, d, <sup>3</sup>*J* = 5.8 Hz, CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  13.5 (Me), 21.5 (Me), 38.7 (NMe), 62.5 (CH<sub>2</sub>O), 79.6 (CH), 107.2 (CH), 127.3 (2 CH), 129.5 (2 CH), 130.4 (CH), 131.4 (C), 132.5 (CH), 149.5 (C), 158.2 (C=N), 166.2 (C=O) ppm. MS, m/z (%): 329 (M<sup>+</sup>, 15), 284 (82), 91 (68), 45 (100). Anal.Calcd for C<sub>17</sub>H<sub>19</sub>N<sub>3</sub>O<sub>2</sub>S (329.42): C, 61.98; H, 5.81; N, 12.76. Found: C, 62.18; H, 5.96; N, 12.92.

## *Ethyl* 1-methyl-5-(4-nitrophenylimino)-1,8a-dihydro imidazo[1,2-c][1,3]-7-carboxylate (5d):

Yellow oil; Yield: 0.61 g (85%). IR (KBr) ( $v_{max}$ /cm<sup>-1</sup>): 1737, 1696, 1573, 1494, 1389 and 1295 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  1.26 (3 H, t, <sup>3</sup>*J* = 7.4 Hz, Me), 2.92 (3 H, s, NMe), 4.22 (2 H, q, <sup>3</sup>*J* = 7.4 Hz, CH<sub>2</sub>O), 5.53 (1 H, d, <sup>3</sup>*J* = 6.3 Hz, CH), 6.58 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.64 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 7.16 (2 H, d, <sup>3</sup>*J* = 7.8 Hz, 2 CH), 7.22 (1 H, d, <sup>3</sup>*J* = 6.3 Hz, CH), 8.12 (2 H, d, <sup>3</sup>*J* = 7.8 Hz, 2 CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  14.6 (Me), 38.5 (NMe), 61.8 (CH<sub>2</sub>O), 81.4 (CH), 107.4 (CH), 124.3 (C), 125.2 (2 CH), 127.5 (2 CH), 130.5 (CH), 133.6 (C), 142.6 (C), 154.7 (C), 159.4 (C=N), 165.8 (C=O) ppm. MS, m/z (%): 360 (M<sup>+</sup>, 10), 315 (76), 45 (100). Anal.Calcd for C<sub>16</sub>H<sub>16</sub>N<sub>4</sub>O<sub>4</sub>S (360.39): C, 53.32; H, 4.47; N, 15.55. Found: C, 53.46; H, 4.64; N, 15.76.

## *N-[7-(4-methoxyphenyl)-1-methyl-1,8a-dihydro imidazo[1,2-c][1,3]thiazine-5-ylidene]-N-(4methylphenyl) amine (5e):*

Yellow oil; Yield: 0.65 g (90%). IR (KBr) ( $v_{max}/cm^{-1}$ <sup>1</sup>): 1698, 1589, 1494, 1386 and 1296 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 2.24 (3 H, s, Me), 2.93 (3 H, s, NMe), 3.85 (3 H, s, MeO), 5.23 (1 H, d,  ${}^{3}J = 5.9$  Hz, CH), 6.65 (1 H, d,  ${}^{3}J = 6.5$  Hz, CH), 6.73 (1 H, d,  ${}^{3}J =$ 6.5 Hz, CH), 6.83 (1 H, d,  ${}^{3}J = 5.9$  Hz, CH), 7.03 (2 H, d,  ${}^{3}J$  = 7.6 Hz, 2 CH), 7.16 (2 H, d,  ${}^{3}J$  = 7.6 Hz, 2 CH), 7.28 (2 H, d,  ${}^{3}J$  = 7.6 Hz, 2 CH), 7.43 (2 H, d,  ${}^{3}J$  = 7.6 Hz, 2 CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$ 21.8 (Me), 39.2 (NMe), 55.3 (MeO), 82.6 (CH), 108.4 (CH), 113.8 (2 CH), 123.2 (CH), 126.2 (2 CH), 128.4 (2 CH), 130.7 (2 CH), 131.2 (CH), 131.8 (C), 133.4 (C), 134.6 (C), 149.3 (C), 154.8 (C=N), 161.4 (C) ppm. MS, m/z (%): 363 (M<sup>+</sup>, 15), 272 (86), 107 (100). Anal.Calcd for C<sub>21</sub>H<sub>21</sub>N<sub>3</sub>OS (363.48): C, 69.39; H, 5.82; N, 11.56. Found: C, 69.52; H, 5.97; N, 11.72.

## *N-[7-(4-methoxyphenyl)-1-methyl-1,8a-dihydro imidazo[1,2-c][1,3]thiazine-5-ylidene]-N-(4bromophenyl) amine* (5*f*):

Yellow oil; Yield: 0.74 g (87%). IR (KBr) ( $v_{max}$ /cm<sup>-1</sup>): 1696, 1585, 1497, 1389 and 1295 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 2.87 (3 H, s, NMe), 3.87 (3 H, s, MeO), 5.56 (1 H, d, <sup>3</sup>*J* = 6.3 Hz, CH), 6.62 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.74 (1 H, d, <sup>3</sup>*J* = 6.8 Hz, CH), 6.96 (1 H, d, <sup>3</sup>*J* = 6.3 Hz, CH), 7.12 (2 H, d, <sup>3</sup>*J* = 7.5 Hz, 2 CH), 7.23 (2 H, d, <sup>3</sup>*J* = 7.6 Hz, 2 CH), 7.45 (2 H, d, <sup>3</sup>*J* = 7.6 Hz, 2 CH), 7.57 (2 H, d, <sup>3</sup>*J* = 7.6 Hz, 2 CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  38.5 (NMe), 55.6 (MeO), 81.8 (CH), 106.8 (CH), 113.2 (2 CH), 114.6

(C), 123.5 (CH), 126.2 (2 CH), 128.4 (2 CH), 130.2 (CH), 132.4 (2 CH), 133.4 (C), 134.2 (C), 151.2 (C), 155.3 (C=N), 160.4 (C) ppm. MS, m/z (%): 428 (M<sup>+</sup>, 15), 321 (58), 107 (100). Anal.Calcd for  $C_{20}H_{18}BrN_{3}OS$  (428.35): C, 56.08; H, 4.24; N, 9.81. Found: C, 56.23; H, 4.42; N, 9.95.

# *N-[7-(4-methylphenyl)-1-methyl-1,8a-dihydro imidazo[1,2-c][1,3]thiazine-5-ylidene]-N-(4nitrophenyl) amine* (5g):

Yellow oil; Yield: 0.60 g (80%). IR (KBr) ( $v_{max}/cm^{-1}$ ): 1695, 1589, 1496, 1387 and 1292 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 2.25 (3 H, s, Me), 2.93 (3 H, s, NMe), 5.52 (1 H, d,  ${}^{3}J = 5.9$  Hz, CH), 6.67 (1 H, d,  ${}^{3}J = 6.5$  Hz, CH), 6.78 (1 H, d,  ${}^{3}J = 6.5$  Hz, CH), 6.92 (1 H, d,  ${}^{3}J = 5.9$  Hz, CH), 7.15 (2 H, d,  ${}^{3}J = 7.5$  Hz, 2 CH), 7.24 (2 H, d,  ${}^{3}J = 7.6$  Hz, 2 CH), 7.82 (2 H, d,  ${}^{3}J = 7.6$  Hz, 2 CH), 7.93 (2 H, d,  ${}^{3}J = 7.6$  Hz, 2 CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  21.4 (Me), 38.9 (NMe), 81.6 (CH), 107.4 (CH), 123.5 (CH), 125.3 (2 CH), 127.3 (2 CH), 128.6 (2 CH), 130.4 (2 CH), 131.2 (CH), 134.3 (C), 146.3 (C), 148.4 (C), 150.4 (C), 156.4 (C=N) ppm. MS, m/z (%): 378 (M<sup>+</sup>, 15), 287 (62), 91 (100). Anal.Calcd for C<sub>20</sub>H<sub>18</sub>N<sub>4</sub>O<sub>2</sub>S (378.45): C, 63.47; H, 4.79; N, 14.80. Found: C, 63.62; H, 93; N, 14.94.

# *Ethyl* 1-methyl-5-(tert-butylimino)-1,8a-dihydro imidazo[1,2-c][1,3]thiazine-7-carboxylate (5h):

Yellow oil; Yield: 0.57 g (97%). IR (KBr) ( $v_{max}/cm^{-1}$ ): 1734, 1563, 1476, 1345 and 1242 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  1.25 (3 H, t, <sup>3</sup>J = 7.3 Hz, Me), 1.28 (9 H, s, *Me*<sub>3</sub>C), 2.87 (3 H, s, NMe), 4.23 (2 H, q, <sup>3</sup>J = 7.3 Hz, CH<sub>2</sub>O), 5.27 (1 H, d, <sup>3</sup>J = 6.4 Hz, CH), 6.63 (1 H, d, <sup>3</sup>J = 6.7 Hz, CH), 6.82 (1 H, d, <sup>3</sup>J = 6.7 Hz, CH), 7.15 (1 H, d, <sup>3</sup>J = 6.4 Hz, CH) ppm. <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>):  $\delta$  14.3 (Me), 30.2 (*Me*<sub>3</sub>C), 38.2 (NMe), 62.5 (CH<sub>2</sub>O), 80.6 (CH), 107.8 (CH), 123.4 (C), 130.2 (CH), 133.4 (CH), 156.2 (C=N), 165.6 (C=O) ppm. MS, m/z (%): 295 (M<sup>+</sup>, 20), 238 (86), 57(100), 45 (88). Anal.Calcd for C<sub>14</sub>H<sub>21</sub>N<sub>3</sub>O<sub>2</sub>S (295.40): C, 56.92; H, 7.17; N, 14.22. Found: C, 57.12; H, 7.35; N, 14.40.

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