

## Toxicity of pesticides to plants and non-target organism: a comprehensive review

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### Abstract

Pesticides are agrochemicals used to increase agronomic production and higher economic profit. However, these chemicals could be persistent in soil and water, collective in silt or bio-aggregates in biota relying upon their dissolvability, prompting various kinds of natural contamination. These pesticides have also shown detrimental effects such as phytotoxicity, genotoxicity, and cytotoxicity on target plants along with alteration in their antioxidant system. The demerits continue as they also affect non-target species such as humans, birds, animal, and other aquatic organisms. The higher accumulations of pesticides are also responsible for the generation of ROS that leads to oxidative stress and finally cell demise. Thus, in this review the toxicity of pesticides will be discussed in detail at a cellular and molecular level linked with the response of the plant defense system. Besides, various strategies that have been commonly used worldwide to remediate the toxicity have also been highlighted in the later section. The study will help plant researchers and chemical engineers to understand the gap between the research and a novel, innovative, and cost-effective technique/ procedure will be outlined for healthy environments.

Keywords: pesticides; cytotoxicity; antioxidant system; phytohormones

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#### Introduction

In ancient times, the introduction of natural compounds or synthetic pesticides to increase agricultural crop productivity for the rapidly growing global population had been proved an effective strategy against the adverse climatic condition, coupled with biotic and abiotic stress (Kumar and Verma, 2018). Beside this, rapid population growth also led to introduction of new component of pesticides in modern agricultural system viz organochlorine, organophosphate,

\*Corresponding author *E-mail address*: aishakamal04@gmail.com Received: December, 2019 Accepted: June,2020 carbamates, pyrethroids, phenoxy herbicides, benzoic acid herbicides, triazines, and urea as insecticides, fungicides, herbicides, rodenticides, and nematicides for improved agricultural output and productivity yield by protecting crop from various pest, insect, and other possible threats (Mieldazys et al., 2015; Fatma et al., 2018). Ideally, a pesticide must have a deadly effect on only unwanted growth or pest that affect productivity but no lethal effect on non-target species, including man. But disappointingly, this is not the situation, as it affects nearly all non-specific targets and causes other serious consequences along with major environmental issues thus, the

uses of pesticides are always contradictory (Aktar, 2009). The adjustment and resistance developed by pests to these synthetic substances further increases the demand for a new formulation of pesticides to protect the crop and further increases the cost of food production (Parween et al., 2016). It was also presumed that only a small part of pesticides (<0.1%) of total applied pesticide concentrations reached to the active site while major portion run-off or accumulated on offtarget sites. These excess pesticides should also be removed from the environment with modern equipment and techniques and these practices also rise the expenditure on crop production cost (Parween et al., 2016). However, after making the maximum effort to remove or minimize the level of residual pesticides, only a little or nonsignificant result is achieved. As pesticides degradation is the sequential process of chemical reaction, arrangement of chemical moieties and metabolic product of pesticides is subject to abiotic natural conditions (temperature, dampness, soil pH, and so forth.) and pesticide chemical composition (hydrophilicity, pKa/b, Kow, etc.) (Ahemad and Khan, 2012). The other most unacceptable fact is that pesticide application (over or higher doses/ repeated uses) shows a detrimental effect on a host plant that is being protected by these pesticides. The end product or pesticide residual incorporate into the host plant consequently and damage important biomolecules in the plant when enters into the metabolic pathway and thus leads to ROS production (Faize et al. 2011; Mahmood et al., 2014). Sufficient literature is available revealing the negative effects at cellular/genetic level, cytotoxic, genotoxicity, chromosomal abnormalities, DNA damage, and increased rates of mutation induction associated with pesticides usage (Anitha and Savitha 2013; Fatma et al., 2018) but their detailed mechanism of action is still unclear. Pesticide toxicity has been usually assessed by means of phytotoxicity symptoms in plants, changes in photosynthetic pigment, plant growth and development, alteration in the antioxidant system, and specific molecular changes (Chris et al. 2011; Horbowicz et al., 2013). The detrimental effects of pesticides are not only limited to plant but their excessive application also contaminates soil and water, kills beneficial

insects and microorganism, birds, fish, and nontarget plants, and negatively affect the environment and human health (Nicolopoulou-Stamati et al., 2016). These chemical pesticides are also associated with dermatological, gastrointestinal, neurological, cancer-causing, respiratory, regenerative, endocrine, and other health issues in humans (Sanborn et al., 2007; Hutter and Moshammer, 2018). As the application of harmful chemical pesticides affect the entire environment, there is an urgent need to develop innovative ideas/ approaches in the farming sector with the eco-friendly modern technique used to avoid these lethal chemical moieties. Also, the exploitation of some biologically active components extracted with natural resources should also be taken in the active mode of agricultural practices. Thus, this review will critically discuss different pesticides and their effect on the plant from the early stage of seed germination to maturity and highlight the suitable methods or strategies to nullify the exceeded concentration than permissible limits. Before discussing the alternative approach, it is worth to discuss the correlation of different pesticides in growth, development, and physiological response in plants.

### Morphotoxicity, phytotoxicity, cytotoxic, and genotoxicity effect of pesticides

Pesticides, above permissible dose severely affect cell development, photosynthesis, biosynthesis, and molecular responses at various stages of plant life (DeLorenzo et al., 2001; Shakir et al., 2016). Excessive and repeated application of these chemical pesticides exhibit reduced germination, retarded growth of vegetative and reproductive organs, and severely affect various morphological and physiological efficiencies in several important crops (Tort et al., 2005; Rio et al., 2012; Aksoy et al., 2013; Kilic et al., 2015; Shakir et al., 2016). The visible morphological symptoms that are frequently used to assess the effect of pesticide usages on crops are poor germination, retarded growth, yellowing and curling of leaves, poor reproductivity and yield. Common herbicide acifluorfen, a diphenyl ether utilized on numerous harvests, such as soybean, bean, pea, wide bean, cotton, and spinach causes leave injury (Saladin and Clément, 2005) and in some cases chlorosis, necrosis, wilting, and shrinking of leaves have also been reported. In another study, exogenous application of insecticides such as dimethon-S-methyl (organophosphate), dimethoate (organophosphate), pirimicarb (carbamate), dicofol (diphenylethane) to wild tamarind plant affect dry matter, leaf area, and leaf number and decrease or inhibit photosynthetic machinery (Dalzell and Mullen, 2004, Saladin and Clément, 2005). Furthermore, in vitro developed grapevines with the treated fungicides pyrimethanil (anilinopyrimidine) and fludioxonil (phenylpyrrole) at field focus or at a lower rate showed a decreased rate of photosynthesis, low carotenoid and chlorophyll contents, sucrose, and hexose content along with overproduction of secondary metabolites (Scarponi et al., 2002; Saladin et al., 2003). Besides this, net CO<sub>2</sub> absorption also reduced along with changes in stomatal conductance and intercellular CO<sub>2</sub> concentration in Malusdomestica and Cucumissativus after fungicide application (Untiedt and Blanke; 2004; Xia et al., 2006). The use of a non-systemic fungicide, fludioxonil, in Vitis vinifera caused a reduction in net CO<sub>2</sub> absorption and in intercellular CO<sub>2</sub>; however, stomatal conductance was not influenced (Petit et al., 2008). Another research carried out by Saladin and Clement, (2003), revealed that use of a similar fungicide, fludioxonil, and pyrimethanil with in vitro developed plants and fruiting cuttings of Vitis vinifera showed advanced distinctive physiological reactions: in vitro plants, the two fungicides diminished net CO<sub>2</sub> absorption, transpiration rate, stomatal conductance, and intercellular CO<sub>2</sub> focus; however in the fruiting cuttings, the fungicides did not influence CO<sub>2</sub> trade either transpiration rates (Dias, 2012). The herbicide glufosinate (phosphinic acid) applied to tomato plants caused ammonium accumulation and ethylene generation as a marker of both stress and bright senescence (You and Barker, 1997). The treatment of pea plants with herbicide imazethapyr (imidazolinone) enhanced absolute free amino acid content but decreased protein content demonstrating protein hydrolysis (Royuela et al., 2003). Besides herbicides, fungicides (phenylpyrrole anilino pyrimidine) also reported to decrease amino acid

content; enhance protein hydrolysis, and stress protein accumulation in grapevine (Llorens et al., 2000). Hence, it was reported that both carbon and nitrogen metabolism showed abnormal pathway regulation but the alteration in metabolism depends upon the type of crop and duration of pesticide exposure. Extensive studies about the deleterious effect of pesticides on plant growth, development, seed germination, flowering, pollen germination, and pollen tube growth have already been done very precisely. Recently, it was found that application of mancozeb, imidacloprid, and sulfentrazone, on model plant Allium cepa showed cytotoxic and genotoxic effects by inducing different types of chromosomal abnormalities, likely sticky, disoriented and fragmented chromosomes, abnormal DNA condensation, and chromosome coiling by spindle inactivation and thus reduced mitotic index (Bianchi et al., 2016; Fatma et al., 2018). The detailed list of commercially available pesticide and their mode of action in various crops are listed in Table 1. In addition to these parameters, the literature is very scary on the pesticides' effect on antioxidant modulation, plant defense mechanisms, pathogenesis-related proteins (PRP), and alteration of secondary metabolites and phenolic compounds. Thus, an attempt has been made to explore the modulation antioxidant system, and of the relation component to detoxify the negative effect of pesticides in the next section.

### Pesticides-induced oxidative stress and modulation of the antioxidant system

Highly regulated mechanisms are involved in maintaining the equilibrium between ROS formation and their detoxification during normal growth and metabolism. But unfavorable such as water scarcity, higher condition accumulation of salts, ions, and toxic metal coupled with climatic changes are responsible for excessive ROS production that disturbs the equilibrium and causes oxidative stress (Khan et al., 2015; Ahmad et al., 2017). Although, ROS are short-lived, toxic molecules, they degrade important biomolecules such as lipids and

| SI.<br>No | Plants/ species         | Pesticides        | Nature/<br>Chemical Family       | Toxicity  | References                  |
|-----------|-------------------------|-------------------|----------------------------------|---|-----------------------------|
| 1.        | Allium cepa L           | Endosulfan        | Insecticide/<br>Organophosphate  | Irreparable chromosomal<br>damage   | (Khan and Damalas,2013)     |
| 2.        | Lens culinaris L        | Mancozeb          | Fungicide/<br>Dithiocarbamate    | Morphological and anatomical  | (Bashir et al., 2007)       |
| 3.        | Zea mays L              | Omethoate<br>(OM) | Insecticide/<br>Organophosphorus | Yellowing of leaf,<br>distortion of stomata and,<br>loss of chloroplast<br>mitochondrial structure<br>and function  | (Wen et al.,2017)           |
| 4.        | Phaseolus vulgaris L.   | Butachlore        | Herbicide/<br>Acetanilide        | Inhibit photosynthesis,<br>protein synthesis, RNA<br>synthesis, lipid Synthesis   | (Chang et al 1985)          |
| 5.        | Vigna radiata L.        | Chlorpyrifos      | Insecticide/<br>Organophosphate  | Decreasing root and shoot<br>length   | (Parween, et al. 2011)      |
| 6.        | Vicia faba              | Fenthion          | Insecticide/<br>Organophosphate  | DNA damage, program<br>cell death including<br>chromatin condensation,<br>cytoplasmic vacuolization,<br>nuclear shrinkage,<br>condensation of the<br>protoplast, fragmentation<br>with apoptotic-like corpse<br>formation | (Cortés-Eslava et al. 2018) |
| 7.        | Phaseolus vulgaris L.   | Atrazine          | Herbicide/<br>Triazine           | Effects on fine Structure<br>of Chloroplasts  | (Ashton et al., 1963)       |
| 8.        | Cucumis sativus L.      | Carbendazim       | Benzimidazoles/<br>Fungicide     | It affects the anti-oxidant system in different plant tissues   | (Zhang et al., 2007)        |
| 9.        | Triticum aestivum L.    | Malathion         | Insecticide/<br>Organophosphate  | Higher concentration of<br>malathion inhibits the<br>production of primary<br>metabolites, seed<br>germination and<br>photosynthetic pigments.  | (Kumar& Sharma, 2017)       |
| 10.       | Solanum<br>lycopersicum | Pirimicarb        | Insecticide/<br>carbamate        | Morphological parameters<br>are affected such as<br>germination and growth  | (Chahid et al., 2013)       |

| Table 1  |
|--|
| Inhibitory effects of pesticides and their mode of action on several important crops |

proteins, membrane disruption, along with denaturation of nucleic acids, leading to cell death (Anjum et al., 2011; Ahmad et al., 2018). A similar effect is also caused by pesticide-induced stress in plants that are being protected by the application of pesticides and also to non-target plants by altering cell metabolism, biochemical and other

physiological responses. In a recent study, application of emamectin benzoate, alphacypermethrin, and imidacloprid pesticides at prescribed level or high level of dosage to tomato seedlings, disturbed cell viability, cell injury coupled with alteration in SOD, CAT, POD activity, increment of GR antioxidant, accumulation of

proline, and increased content of MDA level (Shakir et al., 2018). Besides this, the toxicity of mancozeb and chlorpyrifosin, a dose-dependent manner, enhanced the antioxidant system (CAT, POD, and SOD) in Allium to tolerate morphotoxicity of the pesticides but failed beyond the precise limit (Fatma et al., 2018). In addition to this, histochemical detection by NBT methods carried out by Kamal and co-worker pointed out that application of different fungicides and insecticides on mung bean showed an enhanced level of ROS generation and disturbance in the equilibrium of redox hemostasis pathway (unpublished data). The pesticides also interfere with the photosynthetic pathway and bind to (QA) D1 protein and inhibit electron transport by acting as a non-reducible analog of plastoquinone and block the PQH<sub>2</sub> for example. Paraquat, a commonly known herbicide, interferes with photosynthesis via the generation of free radical O<sup>2.-</sup>. Free radicals disturb the electron gradient system, thus, affecting the transport of electron to the final molecule and hindering the process of energy generation (Laskay and Lokas, 2011). To overcome pesticides prompted oxidative stress, plants themselves have created viable safeguard systems, for the most part, alluded to as the inner tolerant component through which plants get by against oxidative strain by means of expanding the action of antioxidant enzymes (Weisany et al., 2012). The antioxidant enzyme activities in plants determine their scavenging ability (Xu et al., 2011). A wide range of antioxidant enzyme response has been discussed in the literature in the context of the application of the pesticide. For instance, enhanced activity of catalases and isoenzymes was reported to minimize the level of H<sub>2</sub>O<sub>2</sub> in wheat and tomato plants when exposed to a higher level of insecticides pentachlorophenol 2, 4-dichlorophenol (Michałowicz et al., 2009; Shakir et al., 2018). However, different parts of the plant showed a slight variation in antioxidant responses e.g., root and shoot showed differential responses in POD activity upon application of chlorotoluron herbicide (Song et al., 2007). Besides this, GST is a multifunctional enzyme that activates the ascorbate-glutathione cycle which regulates the conversion of herbicide/ ROS to GSH thus providing tolerance to these chemical pesticides (Edwards et al., 2000). The induced level of GST

activity was reported in Arachis hypogaea and Triticum aestivum plants in response to glyphosate and prometryn, respectively (Singh and Prasad, 2018). Moreover, the most important osmolytes, proline, has been considered as the signal of biotic or abiotic stress as its concentration elevated in stress conditions (Ahmad et al., 2017). Proline is the only organic solute that has been shown to protect plants against singlet oxygen and free radical-induced damage that is why proline is able to stabilize proteins, DNA, and membranes (Matysik et al., 2002). Almost every treatment of pesticides enhances the proline level in plants indicating the level of stress in crop plants. To understand the proper mechanism induces by pesticides, extensive scientific research should be carried out and the most appropriate and possible pathway should be given. In the next section, various outcomes of pesticide effect or more specifically, insecticides on non-target organisms will be critically discussed.

# Mode of collateral toxicity induced by pesticides to human and non-target organisms

Despite all the sufficient scientific information collected and the facts revealed in this area, little effort has been made to give a complete assessment on the danger dimensions of a wide range of pesticides to the different nontarget taxa. The lethality and toxicity of pesticides are a result of their biochemical strategy for cell or physiological responses in life forms. The toxicity caused by pesticides is associated with various human disease such as astrointestinal, respiratory, endocrines. well reproductive as as neurodegenerative diseases (Mostafalou and Abdollahi, 2013). The major chronic diseases are Parkinson (Van Maele-Fabry et al., 2012), Alzheimer (Parronet al. 2011), diabetes (Thayer et al., 2012), cardiovascular diseases (Zamzila et al., 2011) and chronic nephropathies (Siddharth et al., 2012) and most lethal cancer disease are more prone to population where excessive pesticide has been used and either most of these diseases are not curable or the available drugs show severe

| SI.No | Pesticides                               | Chemical Nature                              | Site of action                           | Symptoms  | Chronic disease                                  |
|-------|--|--|--|---|--|
| 1.    | Chlorpyrifos,<br>Lindane,<br>Methoxchlor | Chlorinated<br>Hydrocarbon                   | Kidney, Liver                            | Aplastic anemia,<br>hyperesthesias  | Leukemia, Parkinson<br>disease, Prostrate cancer |
| 2.    | Phenoxy derivative                       | 2,4-D and 2,4,5-T                            | Respiratory and GI tract                 | Diarrhoea, metabolic<br>acidosis  | Lung cancer, gall bladder<br>cancer              |
| 3.    | Diquat, paraquat                         | Dipyridyls                                   | Kidney, Liver, GI<br>tract               | Nausea, vomiting,<br>Jaundice   | Stomach cancer                                   |
| 4.    | Captofol                                 | Fungicides                                   | CNS, lungs, liver                        | Chronic cough,<br>Pulmonary edema   | Pancreatic and lung<br>cancer                    |
| 5.    | Chlorpyrifos                             | Organophosphate                              | Neurotoxins                              | Muscular cramps, fatigue,<br>asthma   | Leukemia, and colorectal cancer                  |
| 6     | Methylbromide                            | Organobromine                                | Neurotoxins                              | Headache, memory loss   | Neurological effects                             |
| 7     | Toxaphene                                | <u>bicyclic</u><br>chloroorganic<br>compound | lungs, nervous<br>system, and<br>kidneys | Not specified   | <u>bronchial carcinoma</u> in                    |
| 8     | Pendimethalin                            | Dinitroaniline                               | Skin, Eye                                | Skin, eye and respiratory irritations                                       | Rectum cancer                                    |
| 9.    | Oxychlordane                             | Organochlorine                               | adipose tissues,<br>Liver                | migraines, respiratory<br>infections, anxiety,<br>depression, blurry vision | Non-Hodgkin's<br>Lymphoma                        |

Table 2 Pesticides responsible for human disease

side effects (Table 2). The molecular machinery related to the disease is very specific and involves different modes of action. However, the oxidative generation has been considered as the primary reason for all these chronic diseases which involve various caspases, suppression of immune and autophagy system, and formation of defective protein (Kaur and Kaur, 2018). Besides, this epigenetic alteration involves the acetylation, methylation, and also involves modifications to histone protein at the gene level and these alterations are heritable to the next generation (Mostafalou and Abdollahi, 2013). Sometimes, these modifications mimic DNA repairs system and also induce lethal mutation.

The pesticide toxicity is not limited to humans but influences other terrestrial and aquatic invertebrate species. well-known effects of pesticides range from organismal toxicology and behavioral effects at population level. While poisonous quality is constrained by the internal bit required to cause the end of a living thing (paracelsus), the distinction depends upon the biochemical or physiological frameworks centered by the pesticides, which can either change essentially among taxa (for instance explicit bug sprays showers) or be similar for every single living being (for instance wide range creepy-crawly splashes) (Jayaraj et al., 2016). Obviously, broad range insects' splashes are risky to a wide scope of life forms, paying little heed to whether their destructive doses are in a general sense higher for humans and other creatures than for little bugs (Katagi, 2010). Life shapes weak to this kind of poisons join all arthropods, vertebrates, night crawlers, marine worms, mollusks, and distinctive living creatures with a made neuronal structure. Acetylcholinesterase For example, (AChE)

inhibitors include the organophosphorus (for instance chlorpyrifos, dimethoate and 60 others) and carbamate (for instance aldicarb, methomyl, pirimicarb, and 25 others) pest showers are the sweeping extents and risky poisonous substances, especially to bumblebees, warm-blooded creatures and feathered creatures (Sanchez-Bayo, 2012). The avermectins (for instance abamectin) are agonists of these receptors (Casida and Durkin, 2013), and are particularly deadly to all arthropods (for instance scroungers, bumble bees, and 8legged animal), and they are the most unsafe pest sprays to an aquatic organism.

### Possible strategy to reduce toxicity of pesticides

### Phytohormones-mediated mitigation of pesticides toxicity

Modern agricultural practices have been known for the excessive utilization of synthetic chemicals known for their capacity to decrease physiological efficiency of the plant and negative impact on human health, and degrade the natural environment. Therefore, a crucial, premeditated line of the plan is required to limit the practices of these agrochemicals, and cost effective and innovative methodology should be implemented for sustainable agricultural production. Furthermore, current farming practices need to explore ecologically competent practices with few or no risk of environmental threat and sustain yield and productivity of crops that provide stability to agricultural business (McBratney et al., 2005; Watcharaanantapong et al., 2014). Thus, in this context, we will emphasize the best possible strategy either adopted by the plant itself or scientifically opted approaches to reduce the toxicity caused by pesticide application on growth and development of the plant.

Plant overall growth and development metabolism well-coordinated is by phytohormones for normal growth and development of crop (Wani et al., 2016; Ahmad et al., 2017). Full-on literature has also been documented to prove phytohormones as an efficient method to reduce the various abiotic stresses such as drought, salinity, heavy metal toxicity, radiation, and also many biotic stress

(Khan et al., 2015; Ahmad et al., 2018). Recently, the involvement of phytohormones has gained more attention due to their potent efficiency against various pesticides applied to agricultural crops (Fatma et al., 2018). In this context, salicylic acid (SA), important plant signaling molecules and a well-known growth regulator, has been reported to regulate local defense and systemic resistance response to various pathogen attack (Klessig and Malamy, 1994; Ahmad et al., 2019). Previous studies showed that 0.1 mM SA applied to Brassica napus seeds reduced the napropamide levels (herbicides) and also improved the growth parameters by decreasing the abundance of  $O^{2-}$ . H<sub>2</sub>O<sub>2</sub> as well as activities of superoxide dismutase, catalase, and ascorbate peroxidase, and increased activities of guaiacol peroxidase (POD) and glutathione-S-transferase (GST) in napropamideexposed plants (Cui et al., 2009). Pre-soaking treatment of salicylic acid (1 mM) before the application of Mancozeb pesticides reduced the phytotoxicity effect by regulating the level of detoxifying enzymes and photosynthetic efficiency in Vigna radiata (Fatma et al., 2018). exogenous Moreover, application of brassinosteroids increased the activity of various components of antioxidant defense system comprising of SOD, CAT, GPOX, GR, DHAR, and MDHAR, and increased the tolerance against pesticides application on rice by accumulating the proline and other stress combater under chlorpyrifos (CPF) and imidacloprid pesticide exposure (Sharma et al., 2015; Kumar et al., 2016; Sharma et al., 2017). The treatment significantly enhanced the expression of P450 (P450 monooxygenase), GST, and MRP (multidrug resistance-associated protein) genes responsible for pesticide detoxification in plants (Sharma et al., 2018).

Nowadays, nitric oxide (NO) is extensively used as a suitable candidate for stress signaling under abiotic stress (Yadu et al., 2017). Keeping in mind the role of NO carried out study by application of 100  $\mu$ M SNP as a treatment against mancozeb and chlorpyroios and found enhanced antioxidant activities, increased growth parameters such vigor index and reduced phytotoxicity effect on model plant *Allium cepa* (.

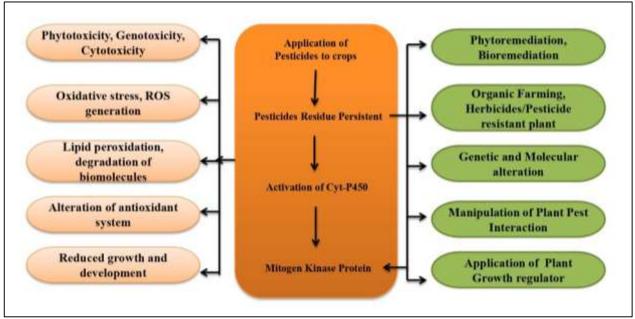


Fig.I. Possible mechanism involved in application of phytohormones and bioremediation process in reducing pesticide toxicity; the (pink part) shows the pesticide-induced cytotoxicity and phytotoxicity responsible for excessive generation of ROS and producing oxidative stress that leads to degradation of biomolecules and reduces plant growth. The green part shows the possible strategies to overcome the toxicity caused by pesticides

In another study, it was reported that mitogenactivated protein kinase (MAPK) and nitric oxide (NO) play an essential role in BR-mediated pesticide detoxification (Yin et al., 2016). Hence, the synergistic effect of the application of phytohormones could be an efficient method to achieve the best amelioration effect. The possible underlying mechanism has been outlined in Fig. I. Recently, JA seed treatment resulted in the significant recovery of chlorophyll content and seedling growth and reduced content of superoxide anion, hydrogen peroxide, and malondialdehyde were enhanced with IMI application. JA seed treatment up-regulated the NADH-ubiquinone expressions of RUBISCO, oxidoreductase (NADH), carboxylesterase (CXE), and P450 under imidacloprid (IMI) toxicity in Brassica juncea plants (Sharma et al., 2018). The detailed mechanism of mitigation of toxicity by salicylic acid, nitric oxide, and jasmonic acid requires extensive scientific research. However, salicylic acid is associated with activation of mitogen kinase protein when higher production of ROS takes place which in turn accumulates and is linked with activation of the full component of Cyt-P450. The works of literature regarding the prominent role of other plant growth regulators is very scanty and provide plant researcher and

broad level of research that should be carried out in very precise manner.

#### Use of pesticides/herbicides-tolerant plants

The toxicological impacts of herbicides are major issues for human wellbeing and environmental assurance organizations (Sunohara et al., 2010). The production of commercially approved herbicides with specificity to specific weeds and effective in low dosage is essential in controlling unwanted growth without affecting another non-target organism. Apparently, lessening herbicide decisions may result in significant monetary and ecological outcomes to farming. The assessment of herbicide-safe weeds includes mind-boggling and exorbitant techniques. Due to cross-resilience, consistent determinations have to be done to minimize the herbicide blockade (Mahmood et al., 2014). The regular course for creating herbicide-resistant crops (HRCs) was first completed by conventional reproducing techniques. Later on, genetic manipulation proved an efficient method to develop HRCs, yet the innovation has been under investigation for its advantages and other ethical issues. Genetically modified HRCs crop are

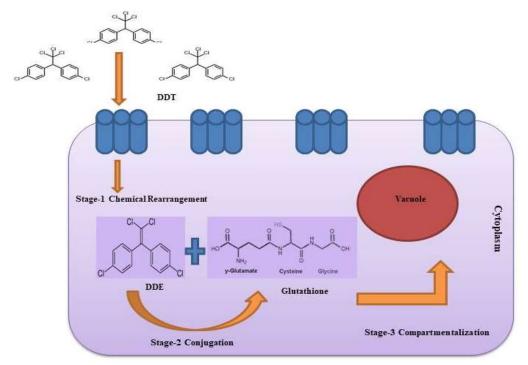


Fig. II. Steps involved in plant driven phytoremediation in plant cells (adopted from Sandermann, 1994); detailed mechanism has been discussed in text.

generally outcomes of cross breeding such as Brassica rapa, Glyphosate-tolerant soybean crops, and most common HR rice varieties (Carpenter and Gianessi, 2001; Madsen and Streibig, 2013). The primary machinery involved in resistance against pesticides in plants is a muddled procedure including various parts, for example, phytochromes, plant cell reinforcement hardware, glycoproteins, and communication of different metabolic framework (Mizutani and Ohta, 2010; Mahmood et al., 2014). The activity found in Cyt P450 in plants has a significant capacity in detoxification, which is very like an animal model system that intervenes with the actuation of a few organophosphates, most widely utilized pesticides. Most of the organophosphates are detoxified through Cyt P450 monooxygenases to accomplish herbicide obstruction; diminished actuation may also be involved with the procedure and it is proved to be a common resistance pathway. Besides extensive environmental studies, it is still unclear what route is opted by plants to treat different contaminants from soil and water. But in some studies it was found that pesticides specifically targeting biologically active compounds lost their functionality due to overexpression or overstimulation of the specific proteins thus plants may use molecular strategy to protect them with specific pre and posttranslation modification (Singer et al., 2003; Mahmood et al., 2014).

#### Plant-associated remediation

Phytoremediation has been considered as the most efficient, innovative, and economical technique used to degrade pesticide residual by using a plant rhizosphere system (Hussain et al., 2009). However, the whole process is very complicated and multistep process comprises of various bio-physiochemical reactions in a systematic manner such as hydrolysis, reduction, and chemical conjugation; physical rearrangement takes place both inside and outside of plant (Hoagland et al., 2000). Numerous plants have been reported to tolerate the higher accumulation of pesticides significantly. The reaction catalyzed during plant-mediated phytoremediation in the cell is represented in Fig. II. These plants are regularly considered for phytoremediation in light of their high number of cultivars because of their significance horticulture and in crop improvement, just as their excellent aggregation capability of a broad scope of natural

contaminants (White, 2010). Organic farming is characterized as the arrangement of creations that are fit for maintaining a strategic distance from or significantly obstruct the utilization of pesticides and artificial composts; domesticated animals' feed adds substances and development regulators. To a vital degree, adaptable natural cultivating framework depends upon crop residues, green manure, pest weeds, crop rotations, off-farm organic wastes, legumes, aspects of biological pest control insects, animal manures. Strategies for organic harvesting are utilized broadly in developing countries, significantly in view of financial elements and fewer assets of synthetic substances. However, they are rising as generally endorsed ideas in the created nations because of long stretches of destructive industry synthetics.

### Use of Bio-pesticide over conventional pesticides

Bio-pesticides are naturally occurring substances extracted from plant isolates, fatty acids, or pheromones that control pests by nontoxic mechanisms. The bio-pesticides are broadly categorized under microbial pesticides, plantincorporated-protectants (PIPs), and biochemical pesticides. The main advantage of bio-pesticides is that they specifically affect the target and related species and do not affect other non-target organisms, are required in low quantity, and are environmental friendly as they are decomposed very speedily by themselves and thus sidestepping the contamination problems caused by a conventional type of pesticides.

The most widely known microbial pesticides have been obtained from bacterium *Bacillus thuringiensis*, which produces a protein that is harmful to specific insect pests and protects crop insect attacks (Greenplate et al., 2003). The commercial mycoinsecticide 'Boverin' is based on *B. bassiana* in combination with a lower dose of trichlorophon to suppress the second-generation epidemics of *Cydiapomonella* (Ferron, 1971). It was also reported that *B. bassiana* and sublethal concentrations of insecticides increased mortality rate when applied to Colorado potato beetle (*Leptino tarsadecemlineata*), recognizing complex synergism between two agents. Recently,

genetically modified (GM) crops have been exploited to express genes encoding insecticidal toxins to reduce the damage caused by arthropod pests. Azadirachtin, a tetranortritarpinoid, is a main active component well-known to interrupt the metamorphosis of insects (Sharma and Malik, 2012). Besides this, meliantetyraolenone and odoratone have also exhibited insecticidal activities against Anopheles stephensi (Siddigui et al., 2003). Peptidomimetics are a non-peptide network and have a strong affinity towards the target site in insectophore and interfere with functional moieties due to similar structures and are considered for the potential in the development of novel insecticides which helps in penetration in insect's guts. The development of a peptidomimetic insecticide is likely to be challenging since non-critical residues determined in insect toxicity bioassays may be vital for averting vertebrate toxicity, via steric hindrance. In addition, these non-critical residues maybe essential for providing insect target subtype selectivity (Nicholson et al., 2007).

### Future prospective

A different class of pesticides has been made by overall agrochemical associations, which generally controls overall sustenance creation, applying new engineered substances with pesticide properties and complete biotechnological advance, thus veering from standard rural strategies. Besides, current rural practices depend on the wide utilization of chemical pesticides that have been related to adverse effects on human wellbeing, untamed life, and typical habitat. These chemical pesticides are a major threat to the environment. Their resistance in soil and plant and slow degradation naturally makes them enter into food chain and therefore, they are a major issue of health risks for humans and other organisms. Pesticides are mainly used for proper growth and good yield of a crop by protecting them from pests, insects, and additional unwanted growth but due to inadequate knowledge of the environment and unskilled hands that fail to maintain the prescribed level of the dose especially in developing countries the outcomes of these higher doses and application of non-specific pesticides are

responsible for the negative effects on plant growth at various morphological, physiological, cellular, and molecular levels. The major consequences caused by these pesticides at the morphological, genetic, and cellular level and associated antioxidant defense system have been discussed in the earlier section of the study. The main aim of this study was to critically review the most appropriate information available regarding pesticides and plant interaction in the natural environment. Besides, various strategies have also been examined such as regulating the defense system, maintaining proper signaling pathway linked with phytohormones, using herbicidetolerant plants, and the application of pesticides and organic farming. However, there is very little information regarding molecular approaches that have been available. Thus a detailed and extensive effort is still required to dissect molecular and genetic approaches to develop pesticide tolerant plants.

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#### References

- Ahemad, M. and M.S. Khan. 2012 'Ecological Assessment of Biotoxicity of Pesticides Towards Plant Growth promoting Activities Of Pea (*Pisum Sativum*)-Specific Rhizobium Sp. Strainmrp1'. *Emirates Journal of Food and Agriculture*, 334-343.
- Ahmad, F., A. Singh and A. Kamal. 2017. 'Ameliorative effect of salicylic acid in salinity stressed *Pisum sativum* by improving growth parameters, activating photosynthesis and enhancing antioxidant defense system'. *Biosci Biotech Res Comm*, 10, 481-489.
- Ahmad, F., A. Singh and A. Kamal. 2019. 'Salicylic Acid–Mediated Defense Mechanisms to Abiotic Stress Tolerance'. Plant Signaling Molecules, Wood head Publishing. pp. 355-369

- Ahmad, F., A. Singh and A. Kamal. 2018. 'Crosstalk of brassinosteroids with other phytohormones under various abiotic stresses'. J Appl Biology & Biotech, 6(1), 56-62.
- Aksoy, O., A. Deveci, S. Kizilirmak and G.B. Akdeniz. 2013. 'Phytotoxic effect of quizalofop-P-ethyl on soybean (*Glycine max* L)'. J Biol Environ Sci 7:49–55.
- Aktar, M. W., D. Sengupta and A. Chowdhury. 2009. 'Impact of pesticides use in agriculture: their benefits and hazards.' *Interdiscip Toxicol.* 2, 1-12.
- Anitha, S.R. and G. Savitha. 2013. 'Impact of mancozeb stress on seedling growth, seed germination, chlorophyll and phenolic contents of rice cultivars'. IJSR, 4(7), 292-296.
- Anjum, S.A., X.Y. Xie, L.C. Wang, M.F. Saleem, C. Man and W. Lei. 2011. 'Morphological, physiological and biochemical responses of plants to drought stress'. African Journal of Agricultural Research, 6(9), 2026-2032.
- Bianchi, J., T.C.C. Fernandes and M.A. Marin-Morales. 2016. 'Induction of mitotic and chromosomal abnormalities on *Allium cepa* cells by pesticides imidacloprid and sulfentrazone and the mixture of them'. *Chemosphere*, 144, 475-483.
- Cali, I.O. and F. Candan. 2009. 'The effect of fungicide application on pollen structure in tomato (*Lycopersicon esculentum Mill.*)'. *Plant J Appl Biol Sci* 3:37–40.
- **Carpenter, J.E.** and **L.P. Gianessi**. 2001. 'Agricultural biotechnology: Updated benefit estimates'. Washington, DC: National Center for Food and Agricultural Policy. (pp. 1-46)
- **Casida, J.E.** and **K.A. Durkin**. 2013. 'Neuroactive insecticides: targets, selectivity, resistance, and secondary effects'. *Annual review of entomology*, 58, 99-117.
- Chahid, K., A. Laglaoui, S. Zantar and A. Ennabili. 2013. 'Effect of three insecticides on tomato (*Solanum lycopersicum*) seedling germination and early plants growth'. *Biological Diversity* and Conservation, 6(1), 57-61.
- **Chang, S.S., F.M. Ashton** and **D.E. Bayer**. 1985. 'Butachlor influence on selected metabolic processes of plant cells and tissues'. *Journal of Plant Growth Regulation*, 4(1-4), 1-9.

- Chris, A., G. Luxmisha, M. Jamson and G. Abraham. 2011. 'Growth, photosynthetic pigments and antioxidant responses of *Azolla filiculoides* to monocrotophos toxicity'. *Journal of chemical and pharmaceutical research*, 3(3), 381-388.
- Cortés-Eslava, J., S. Gómez-Arroyo, M.C. Risueño and P. S. Testillano. 2018. 'The effects of Organophosphorus insecticides and heavy metals on DNA damage and programmed cell death in two plant models'. *Environmental pollution*, 240, 77-86.
- Dalzell, S. A. and B. F. Mullen. 2004. 'Application of pesticides suppress foliar proanthocyanidin content in *Leucaena* species'. *Animal Feed Science and Technology*, 113: 191-198..
- DeLorenzo, M. E., G.I. Scott and P.E. Ross. 2001. 'Toxicity of pesticides to aquatic microorganisms: a review'. *Environ Toxicol Chem* 20:84–98
- Dias, M. C. 2012. ' Phytotoxicity: An overview of the physiological responses of plants exposed to fungicides'. *Journal of botany*, Volume 2012, Article ID 135479, https://doi.org/10.1155/2012/135479.
- Faize, M., L. Burgos, L. Faize, A. Piqueras, E. Nicolas, G. Barba- Espin, M.J. Clemente-Moreno, R. Alcobendas, T. Artlip and J.A. Hernandez. 2011. 'Involvement of cytosolic ascorbate peroxidase and Cu/ Zn-superoxide dismutase for improved tolerance against drought stress'. J Exp Bot, 62:2599–2613.
- Fatma, F., A. Kamal and A. Srivastava. 2018. 'Exogenous Application of Salicylic Acid Mitigates the Toxic Effect of Pesticides in Vigna radiata (L.) Wilczek'. Journal of Plant Growth Regulation, 37(4), 1185-1194.
- Ferron, P. 1971. 'Modification of the development of *Beauveria tenella* mycosis in *Melolontha melolontha* larvae, by means of reduced doses of organophosphorus insecticides'. *Entomologia Experimentalis et Applicata*, 14(4), 457-466.
- Greenplate, J.T., J.W. Mullins, S.R. Penn, A. Dahm, B.J. Reich, J.A. Osborn and Z.W.Shappley. 2003. 'Partial characterization of cotton plants expressing two toxin proteins from *Bacillus thuringiensis*: relative toxin contribution, toxin interaction, and

resistance management'. *Journal of Applied Entomology*, 127(6), 340-347.

- Hoagland R.E., Z.M. Zablotowicz and J.C. Hall. 2001. 'Pesticide metabolism in plants and microorganisms: An overview'. Weed Science, 51(4):472-495.
- Horbowicz, M., C. Sempruch, R. Kosson, D. Koczkodaj and D. Walas. 2013. 'Effect of fluazifop-p-butyl treatment on pigments and polyamines level within tissues of non-target maize plants'. *Pesticide biochemistry and physiology*, 107(1), 78-85.
- Huang, Y.Y., L. Wang, L.F. Huang, Y.L.Yu, Y.H. Zhou and J.K Yu. 2006. 'Pesticides-induced depression of photosynthesis was alleviated by 24-epibrassinolide pretreatment in *Cucumis sativus* L.' *Pestic Biochem Physiol* 86:42–48
- Hussain, S., T. Siddique, M. Arshad and M. Saleem. 2009. 'Bioremediation and phytoremediation of pesticides: recent advances'. *Critical Reviews in Environmental Science and Technology*, 39(10), 843-907.
- Hutter, H.P. and H. Moshammer. 2018. 'Pesticides Are an Occupational and Public Health Issue?' *Int J Environ Res Public Health*. 15(8): 1650.
- Jayaraj, R., P. Megha and P. Sreedev. 2016. 'Organochlorine pesticides, their toxic effects on living organisms and their fate in the environment'. *Interdisciplinary toxicology*, 9(34), 90-100.
- **Kaur K.** and **R. Kaur**. 2018. 'Occupational pesticide exposure, impaired DNA repair, and diseases'. *Indian journal of occupational and environmental medicine*, 22(2):74.
- Khan, J.A., S. Khan and S.F. Usmani. 2013. 'Effect of Endosulfan on seed germination, growth and nutrients uptake of fenugreek plant'. *Journal of Industrial Research & Technology*, 2(2), 88-91.
- Khan M. and C.A. Damalas. 2015. 'Farmers' knowledge about common pests and pesticide safety in conventional cotton production in Pakistan'. *Crop Protection*, 77, 45-51.
- Khan, M.I.R., M. Fatma, T.S.Per, N.A. Anjum and N.A.Khan. 2015. 'Salicylic acid-induced abiotic stress tolerance and underlying mechanisms in plants'. *Frontiers in Plant*

*Science*, 6, 462. https://doi.org/10.3389/fpls.2015.00462

- Kilic, S., Y. Coskun and R.E. Duran. 2015. 'The effects of the insecticide pyriproxyfen on germination, development and growth responses of maize seedlings'. *Fresenius Environ Bull*, 24, 278-284.
- Kilic, S., R.E. Duran and Y. Coskun. 2015. 'Morphological and physiological responses of maize (*Zea mays* L.) seeds grown under increasing concentrations of chlorantraniliprole insecticide'. *Pol J Environ Stud*, 24:1069–1075.
- Klessig, D., F. Daniel and J. Malamy. 1994. 'The salicylic acid signal in plants'. *Plant molecular biology*,1439 -1458.
- Kumar, A. and J.P. Verma. 2018. 'Does plant— Microbe interaction confer stress tolerance in plants: A review?' *Microbiological research*, 207, 41-52.
- Kumar, S. and J.G. Sharma. 2017. 'Effect of Malathion on Seed Germination and Photosynthetic Pigments in Wheat (*Triticum* aestivum L.)'. Asian Journal of Applied Science and Technology 1(7), 158-167.
- Laskay, G. and A. Lakos. 2011. 'Counteracting the effects of parquet on photosynthesis by chlorogenic acid.' Acta Biologica Szegediensis, 55(1), 101-103.
- Llorens, N., L. Arola, C. Bladé and A. Mas. 2000. 'Effects of copper exposure upon nitrogen metabolism in tissue cultured *Vitis vinifera*.' *Plant Science*, 160(1), 159-163.
- Madsen, K.H. and J.C. Streibig. 2013. 'Benefits and risks of the use of herbicide-resistant crops.' Available at www.fao.org. Accessed on, 2013.
- Mahmood, Q., M. Bilal and S. Jan. 2014. 'Herbicides, pesticides, and plant tolerance: an overview. In Emerging technologies and management of crop stress tolerance'. *Academic Press*. pp. 423-448
- Matysik, J., B.A. Bhalu and P. Mohanty. 2002. 'Molecular mechanisms of quenching of reactive oxygen species by proline under stress in plants'. *Current Science*, 525-532.
- McBratney, A., B. Whelan, T. Ancev and J. Bouma. 2005.' Future directions of precision agriculture'. *Precision agriculture*, 6(1), 7-23.

- Michałowicz, J., M. Posmyk and W. Duda. 2009. 'Chlorophenols induce lipid peroxidation and change antioxidant parameters in the leaves of wheat (*Triticum aestivum* L.)'. *Journal of plant physiology*, 166(6), 559-568.
- Mieldazys, A., R. Mieldazys, G. Vilkevicius and A. Stulginskis. 2015. 'Agriculture-Use of pesticides/plant protection products'. EU-OSHA.
- Mizutani. M. and D. Ohta. 2010. 'Diversification of P450 genes during land plant evolution.' *Annual review of plant biology*, 61, 291-315.
- Mostafalou, S. and M. Abdollahi. 2013. 'Pesticides and human chronic diseases: evidences, mechanisms, and perspectives'. *Toxicology and applied pharmacology*. 268(2):157-77.
- Nicholson, G. M. 2007. 'Fighting the global pest problem: preface to the special Toxicon issue on insecticidal toxins and their potential for insect pest control'. *Toxicon*, *49*(4), 413-422.
- Nicolopoulou-Stamati, P., S. Maipas, C. Kotampasi, P. Stamatis and L. Hens. 2016. 'Chemical pesticides and human health: the urgent need for a new concept in agriculture'. *Frontiers in public health*, 4, 148.
- Parron, T., M. Requena, A.F. Hernandez and R. Alarcon. 2011. 'Association between environmental exposure to pesticides and neurodegenerative diseases'. *Toxicol. Appl. Pharmacol*, 256(3), 379-385.
- Parween, T., S. Jan and T. Fatma. 2011. 'Alteration in nitrogen metabolism and plant growth during different developmental stages of green gram (*Vigna radiata L.*) in response to chlorpyrifos'. Acta physiologiae plantarum, 33(6), 2321-2328.
- Parween, T., S. Jan, S. Mahmooduzzafar, T. Fatma and Z.H. Siddiqui. 2016. 'Selective effect of pesticides on plant—A review'. *Critical reviews in food science and nutrition*, 56(1), 160-179.
- Rio, A.D., J. Bamberg, R. Centeno-Diaz, A. Salas,
  W. Roca and D. Tay. 2012. 'Effects of the pesticide furadan on traits associated with reproduction in wild potato species'. Am J Plant Sci 3: 1608–1612.
- Royuela, V., J. Suriñach and M. Reyes. 2003. 'Measuring quality of life in small areas over

different periods of time'. *Social Indicators Research*, 64(1), 51-74.

- Saladin, G., C. Magné and C. Clément. 2003. 'Effects of fludioxonil and pyrimethanil, two fungicides used against Botrytis cinerea, on carbohydrate physiology in *Vitis vinifera* L.' *Pest Management Science*, 83–109
- Sanborn, M., K.J. Kerr, L.H. Sanin, D.C. Cole, K.L. Bassil and C. Vakil. 2007. 'Non-cancer health effects of pesticides. Systematic review and implications for family doctors'. *Can Fam Physician*. 53:1712–20.
- Sanchez-Bayo, F.P. 2012. 'Insecticides mode of action in relation to their toxicity to nontarget organisms'. *Journal of Environmental and Analytical Toxicology*, 4, S4-002
- Scarponi. L., C. Vischetti and N.M. Hassan. 2002. 'Effects of propachlor on the formation of carbohydrates and proteins in *Vicia faba* and the response of its defence mechanisms'. *Agrochimica XLVI* (3-4): 165-175.
- Shakir, S.K., S. Irfan, B. Akhtar, S. Rehman, M.K. Daud, N. Taimur and A. Azizullah. 2018. 'Pesticide-induced oxidative stress and antioxidant responses in tomato (Solanum lycopersicum) seedlings'. Ecotoxicology, 27(7), 919-935.
- Shakir, S.K., M. Kanwal, W. Murad, Z. Rehman, S. ur Rehman, M.K. Daud and A. Azizullah. 2016. 'Effect of some commonly used pesticides on seed germination, biomass production and photosynthetic pigments in tomato (*Lycopersicon esculentum*)'. *Ecotoxicology*, 25(2), 329-341.
- Sharma, D., M.P. Singh, D. Vimal, S. Kumar, R.R. Jha and D.K. Chowdhuri. 2018. 'Benzene induced resistance in exposed *Drosophila melanogaster:* Outcome of improved detoxification and gene modulation'. *Chemosphere*, 201, 144-158.
- Sharma, I., R. Bhardwaj and P.K. Pati. 2015. 'Exogenous application of 28homobrassinolide modulates the dynamics of salt and pesticides induced stress responses in an elite rice variety Pusa Basmati-1.' *Journal of plant growth regulation*, 34(3), 509-518.
- Sharma, S. and P. Malik. 2012. 'Biopesticides: Types and applications'. *International Journal*

of Advances in Pharmacy, Biology and Chemistry, 1(4), 508-515.

- Siddharth, M., S.K. Datta, S. Bansal, M. Mustafa, B.D. Banerjee, O.P. Kalra and A.K. Tripathi. 2012. 'Study on organochlorine pesticide levels in chronic kidney disease patients: Association with estimated glomerular filtration rate and oxidative stress'. J Biochem. Mol. Toxicol. 26(6), 241-247
- Siddiqui, B.S., F. Afshan and T. Gulzar. 2003. 'Tetracyclic triterpenoids from the leaves of *Azadirachta indica* and their insecticidal activities'. *Chem Pharm Bull*.(Tokyo).51:415-417.
- Singh, P. and S.M. Prasad. 2018. 'Antioxidant enzyme responses to the oxidative stress due to chlorpyrifos, dimethoate and dieldrin stress in palak (*Spinacia oleracea L.*) and their toxicity alleviation by soil amendments in tropical croplands'. *Science of the Total Environment*, 630, 839-848.
- Song, N.H., X. Le. Yin, G.F. Chen and H. Yang. 2007. 'Biological responses of wheat (*Triticum aestivum*) plants to the herbicide chlorotoluron in soils'. *Chemosphere*, 68(9), 1779-1787.
- Tort, N., I. Oztork and A. Guvensen. 2005. 'Effects of some fungicides on pollen morphology and anatomy of tomato (*Lycopersicon esculentum mill.*)'. *Pak J Bot*. 37:23–30
- Untiedt, R. and M.M. Blanke. 2004. 'Effects of fungicide and insecticide mixtures on apple tree canopy photosynthesis, dark respiration and carbon economy'. *Crop Protection*, 23(10), 1001-1006.
- Van Maele-Fabry, G., P. Hoet, F. Vilain and D. Lison. 2012. 'Occupational exposure to pesticides and Parkinson's disease: A systematic review and meta-analysis of cohort studies'. *Environ. Int.* 46, 30-43
- Wang, M.E. and Q.X. Zhou. 2006. 'Effect of herbicide chlorimuron-ethyl on physiological mechanisms in wheat (*Triticum aestivum*)'. *Ecotoxicol. Environ. Safety*, 64, 190–197.
- Wani, S.H., V. Kumar, V. Shriram and S.K. Sah. 2016. 'Phytohormones and their metabolic engineering for abiotic stress tolerance in crop plants'. *The Crop Journal*, 4(3), 162-176.
- Watcharaanantapong, P., R.K. Roberts, D.M. Lambert, J.A. Larson, M. Velandia, B.C.

**English** and **C. Wang**. 2014. 'Timing of precision agriculture technology adoption in US cotton production'. *Precision agriculture*, 15(4), 427-446.

- Weisany, W., Y. Sohrabi, G. Heidari, A. Siosemardeh and K. Ghassemi-Golezani. 2012. 'Changes in antioxidant enzymes activity and plant performance by salinity stress and zinc application in soybean ('Glycine max'L.)'. Plant Omics, 5(2), 60- 67.
- Wen, Y. M., L.Y.Meng, S.T. Liu, C.M. Zhu, Y.J. Liu and A.J. Qu. 2017. 'Ultrastructure effects of omethoate and cypermethrin insecticides on maize seedlings'. *Chemistry and Ecology*, 33(9), 883-892.
- Xia, X.J., Y.Y. Huang and L. Wang. 2006. 'Pesticides-induced depression of photosynthesis was alleviated by 24epibrassinolide pretreatment in *Cucumis* sativus L.' *Pesticide Biochemistry and Physiology*, 86(1), 42-48.
- Xu, W., H. Yang, Y. Liu, Y. Yang, P.Wang, S.H. Kim and L.X. Liu. 2011. 'Oncometabolite 2hydroxyglutarate is a competitive inhibitor of

α-ketoglutarate-dependent dioxygenases'. *Cancer cell*, 19(1), 17-30.

- Yadu, S., T.L. Dewangan, V. Chandrakar and S. Keshavkant. 2017. 'Imperative roles of salicylic acid and nitric oxide in improving salinity tolerance in *Pisum sativum* L.' *Physiology and molecular biology of plants*, 23(1), 43-58.
- Yin Z. and H. Wang. 2017. 'Role of atmospheric circulations in haze pollution in December 2016'. Atmospheric Chemistry and Physics, 17(18), 11673-11681.
- You, W. and A.V. Barker. 1997. 'Herbicidal actions of root-applied glufosinate ammonium on tomato plants'. *Journal of Amercian Society for Horticultural Science*, 127(2): 200-204.
- Zhang, L.Z., N. Wei, Q.X. Wu and M.L. Ping. 2007. 'Anti-oxidant response of *Cucumis sativus* L. to fungicide carbendazim'. *Pesticide biochemistry and physiology*, 89(1), 54-59.