

Sodium nitroprusside and salicylic acid decrease antioxidant enzymes activity in soybean

Leila Aalam, Mohammad Sedghi* and Omid Sofalian

Department of Agronomy and Plant Breeding, Faculty of Agriculture and Natural Resources, University of Mohaghegh Ardabili, Iran

Abstract

The present study investigated the effect of salicylic acid (SA) and sodium nitroprusside (SNP) on germination and activity of antioxidant enzymes in soybean under oxidative stress using a factorial experiment based on completely randomized design with three replications. Treatments included SA (0 and 1 mM), SNP (0, 30, and 60μ M), and H₂O₂ (0, 50, and 100 μ M). Results showed that the highest germination percentage (97%) was related to 1 mM SA without SNP and H₂O₂ which was 6% greater than the control and the highest germination rate (0.72) was related to 1 mM SA and 60 μ M SNP without H₂O₂ (6.63% greater than the control). The highest activity for superoxide dismutase (89.17 units mg⁻¹ protein) was achieved by application of 100 μ M H₂O₂ without SNP. In conclusion, oxidative stress increased all antioxidant enzymes while SNP application decreased the enzyme activity and stress severity.

Keywords: soybean; germination; hydrogen peroxide; oxidative stress; catalase

Aalam, L., M. Sedghi and O. Sofalian. 2019. 'Sodium nitroprusside and salicylic acid decrease antioxidant enzymes activity in soybean'. *Iranian Journal of Plant Physiology*, 10(1), 3073-3077.

Introduction

Oxidative injuries pose limitation for growth and production in plants. Reactive oxygen species (ROS) may have harmful effects like lipid peroxidation, changes in cell membrane integrity, and consequently disintegration of its structure (Allen, 1995).

Salicylic acid (SA) is a plant hormone which regulates the antioxidant mechanisms of plants and defense against oxidative stress (Singh Gill and Tuteja, 2010). SA has inhibitory effects on catalase activity and subsequently on establishing the defense mechanisms in plants (Durant and Dong, 2004).

Sodium nitroprusside (SNP) is a nitric oxide (NO) releasing compound (Zheng et al., 2009) that participates in some physiological processes in plants like germination, ROS metabolism, and signal transduction (Nill et al., 2002). Under biotic and abiotic stresses, the concentration of various ROS, e.g. superoxide, hydroxyl radicle, and H_2O_2 increases in plants (Streeter et al., 2001). ROS damage cell membrane and chloroplasts and macromolecules such as proteins, nucleic acids, and lipids (Mittler, 2002). Therefore, this experiment was conducted to study the effect of SA and SNP on germination and its related traits in soybeans under oxidative stress.

^{*}Corresponding author *E-mail address*: m_sedghi@uma.ac.ir Received: December, 2018 Accepted: September, 2019

SA (ma N 4)	SNP	H_2O_2	GP	GR	GU	
(mM)	(µM)	(μM)	(%)	(%)		
		0	91.67 ± 1.53	$0.44\ \pm 0.01$	$\textbf{3.25}\pm\textbf{0.1}$	
	0	50	85 ± 1	$\textbf{0.33} \pm \textbf{0.21}$	$\textbf{4.1} \pm \textbf{0.07}$	
	Ū	100	71 ± 1	$\textbf{0.27} \pm \textbf{0.011}$	5 ± 0.1	
	30	0	89.33 ± 1.53	$\textbf{0.45} \pm \textbf{0.01}$	$\textbf{2.46} \pm \textbf{0.07}$	
	50	50	84.33 ± 2.1	$\textbf{0.36} \pm \textbf{0.02}$	$\textbf{3.53} \pm \textbf{0.11}$	
0		100	$\textbf{79.67} \pm \textbf{1.53}$	$\textbf{0.31} \pm \textbf{0.01}$	$\textbf{4.18} \pm \textbf{0.03}$	
	60	0	$\textbf{91.33} \pm \textbf{1.53}$	$\textbf{0.57} \pm \textbf{0.02}$	$\textbf{1.66} \pm \textbf{0.05}$	
		50	$\textbf{86.33} \pm \textbf{1.53}$	$\textbf{0.47} \pm \textbf{0.02}$	$\textbf{2.29} \pm \textbf{0.07}$	
		100	$\textbf{74.67} \pm \textbf{1.53}$	$\textbf{0.41} \pm \textbf{0.01}$	$\textbf{3.45} \pm \textbf{0.08}$	
		0	$\textbf{97.67} \pm \textbf{1.53}$	$\textbf{0.64} \pm \textbf{0.01}$	$\textbf{2.22} \pm \textbf{0.08}$	
		50	93.67 ± 1.53	$\textbf{0.52} \pm \textbf{0.005}$	$\textbf{2.53} \pm \textbf{0.13}$	
	0	100	$\textbf{90.33} \pm \textbf{1.53}$	$\textbf{0.45} \pm \textbf{0.004}$	$\textbf{2.6} \pm \textbf{0.06}$	
		0	$\textbf{96.67} \pm \textbf{1.53}$	$\textbf{0.67} \pm \textbf{0.01}$	$\textbf{3.14} \pm \textbf{0.1}$	
1	30	50	93 ± 2	$\textbf{0.63} \pm \textbf{0.10}$	$\textbf{2.85} \pm \textbf{0.13}$	
		100	90 ±1	$\textbf{0.53} \pm \textbf{0.01}$	$\textbf{2.23} \pm \textbf{0.08}$	
	60	0	95.33 ± 1.53	$\textbf{0.72} \pm \textbf{0.02}$	$\textbf{1.84} \pm \textbf{0.11}$	
		50	$\textbf{92.33} \pm \textbf{0.58}$	$\textbf{0.68} \pm \textbf{0.01}$	$\textbf{2.2}\pm\textbf{0.06}$	
		100	88.33 ± 1.53	$\textbf{0.58} \pm \textbf{0.01}$	$\textbf{1.71} \pm \textbf{0.08}$	

Table 1 Comparison of physiological traits of soybeans under the influence of SA, SNP, and oxidative stress caused by H_2O_2

SA (salicylic acid), SNP (sodium nitroprusside), H₂O₂ (hydrogen peroxide), GP (germination percentage), GR (germination rate), GU (germination uniformity)

Materials and Methods

A factorial experiment was conducted to investigate the effect of SA and SNP on soybean (cv. Katul) seed germination under oxidative stress based on completely randomized design with three replications in 2017. Treatments consisted of SA (0, 1 mM), SNP (0, 30, and 60 μ M), and H₂O₂ (0, 50, and 100 μ M). Seeds were surface sterilized with sodium hypochlorite 10% and rinsed with distilled water. Then, 25 seeds were planted between paper in Petri dishes and treatment solutions were added. Petri dishes were placed in a germinator (25 ± 1 °C) for 8 days and germinated seeds were counted daily.

Germination percentage (GP), germination rate (GR), mean germination time (MGT), and germination heterogeneity (GU) were calculated using GERMIN program. Seedling growth test was conducted and on day 5 of the study, five seedlings were selected from each treatment and placed in liquid nitrogen immediately for measuring the activity of antioxidant enzymes.

Catalase (EC: 1.11.1.6) activity assayed according to Chance and Maehly procedure (1955). Also, peroxidase (EC: 1.11.1.7) activity was

measured according to the method of MacAdam et al. (1992). Furthermore, superoxide dismutase (EC: 1.15.1.1) activity was assayed according to Sen Gupta et al (1993) method. Data were subjected to analysis of variance using SAS 9.1 software. Means were compared by least significant differences (LSD) test at 5% probability level.

Results

The highest GP (97.67%) was observed by using 1mM SA without SNP and H₂O₂ treatment and GP (84/33%) was observed by 30 μ M SNP and 50 μ M H₂O₂ (Table 1). The highest GR (0.72) was obtained by 1mM SA and 60 m SNP without application of H₂O₂ which was significantly different from 1 mM SA, 60 μ M SNP and 50 μ M H₂O₂ that was 63.6% greater than control and GR (0.63%) was obtained by 1mM SA, 30 µM SNP and 50 μ M H₂O₂ (Table 1). The lowest MGT (1.48 day) was achieved by application of 1 mM SA without H₂O₂ (Table 2) which was 41.9% reduction compared with control and MGT (1.8 day) observed in the treatment with $60 \,\mu\text{M}$ SNP and 50 μ M H₂O₂. The lightest GU (1.66) was observed in the application of 60 μ M SNP while GU (2.85) was Table 2

Comparison of means for the interaction of SA and H_2O_2 on MGT (mean of germination time) in soybean

SA (mM)	H ₂ O ₂ (μM)	MGT (day)
	0	0.25 ± 2.1
0	50	0.43 ± 2.65
	100	0.54 ± 3.11
	0	0.07 ± 1.48
1	50	0.20 ± 1.65
	100	0.23 ± 1.93

SA (Salicylic acid), H₂O₂ (hydrogen peroxide)

Table 3

Table 4

Comparison of means for the interaction of SNP and ${\rm H_2O_2}$ on soybean antioxidant enzymes

SNP	H_2O_2	SOD		POX (unit mg ⁻
(µM)	(µM)	(unit	mg ⁻¹	¹ protein)
		protein)		
	0	6±38.5		7.42±79.33
0	50	6.99±67		8.31±88.5
	100	7.86±89.17		8.48±102.5
	0	6.19±34.5		6.54±74
30	50	3.22±48		9.04±82.17
	100	4.62±58.17		10.85±91.17
	0	5.19±28.83		9.18±61
60	50	7.76±35.83		10.94±72
	100	10.13±44.67		11.17±82.5

SOD (superoxide dismutase), POX (peroxidase)

variance of the effect of treatments on soybean seed germination traits and antioxidant enzymes were shown in Tables 5 and 6.

Discussion

 H_2O_2 is the free radical especially in cell membranes which causes oxidative stress while SA decreases the severity of oxidative stress and increases the proline content (Shi et al., 2009). Protecting plants against ROS was related to SA and its contribution in the synthesis of antioxidant enzymes and activity of the enzymes (Wu and Du, 2008). SA can control the activity of antioxidant enzymes through the temporary accumulation of ABA (Hayat and Ahmad, 2007). SA reduces the activity of catalase at the early stages of stress and increases concentration of H_2O_2 which acts as a secondary messenger for the activation of resistance genes (Hernandez et al. 2001).

SNP is an NO donator which participates in transduction pathway and contributes in some processes like germination, root growth, and stomatal closure (Zheng et al. 2009). Positive effects of SNP on seed germination was reported by Fan et al. (2012) in *Cacumis satirus*. NO

SA (mM)	SNP (µM)	SOD	CAT	POX
		(unit mg ⁻¹ protein)	(unit mg ⁻¹ protein)	(unit mg ⁻¹ protein)
	0	21.3±59	4.37±13.89	9.84±83
0	30	11.36±43.11	4.07±9.89	5.83±74.55
	60	5.17±29.67	4.74±10	8.63±62.78
	0	22.97±70.78	5.12±18.78	10.78±97.22
1	30	9.7±50.67	3.24±17.33	9.38±90.33
	60	9.11±43.22	2.71±14.89	10.24±81.22

SOD (superoxide dismutase), POX (peroxidase), CAT (catalase)

Comparison of means for the interaction of SA and SNP on soybean antioxidant enzymes

obtained by 1 mM SA, 30 μM SNP, and 50 μM H_2O_2 (Table 1).

The highest SOD (89.17 units mg⁻¹ protein) and POX activities (102.5 units mg⁻¹ protein) were observed by using 100 μ M H₂O₂ without SNP and SOD (48 units mg⁻¹ protein) and POX activities (82.17 units mg⁻¹ protein) was observed in the treatment involving 50 μ M H₂O₂ and 30 μ M SNP (Table 3). The highest activities of SOD, CAT, and POX (70.78, 18.78, and 97.22-unit mg⁻¹ protein, respectively) were observed in the treatment with 1 mM SA without SNP (Table 4). Analysis of

releasing compounds have the capability to fight ROS and reduce oxidative damages (Asadi Karam et al., 2016). Effect of NO is dose-dependent and its physiological effects varies at different concentrations and in environmental conditions (Shi et al. 2009).

GR is one of the most important indices among agronomic characters and low GR leads to a lack of homogeneity and density under field conditions. Homogeneity and germination rate of wheat was reported to decrease under drought stress (Fateh et al. 2012). In conclusion, under

Sov	Df	GP	GR	GU	MGT
SA	1	1176**	0.55**	12.33**	11.55**
SNP	2	3.02 ^{ns}	0.77**	5.98**	1.82**
H ₂ O ₂	2	585.41**	0.11**	2.73**	2.47**
SA × SNP	2	14.39**	0.005**	1.73**	0.4**
SA × H ₂ O ₂	2	92.67**	0.002**	4.46**	0.39**
SNP × H ₂ O ₂	4	15.85**	0.001**	0.18**	0.11**
SA × SNP × H ₂ O ₂	4	12.22**	0.0004*	0.22**	0.01 ^{ns}
Error	36	2.2	0.0002	0.008	0.006
CV%		1.68	2.6	3.15	3.75

Table 5 Analysis of variance of the effect of treatments on soybean seed germination traits

ns, *, and ** are non-significant, and significant at 0.05 and 0.01% probability levels, respectively. SA (salicylic acid), SNP (sodium nitroprusside), H₂O₂ (hydrogen peroxide), GP (germination percentage), GR (germination rate), GU (germination uniformity), MGT (mean germination time.

Table 6	Та	bl	le	6
---------	----	----	----	---

Analysis of variance of the effect of treatments on soybean seed antioxidant enzymes

SOV	Df	SOD	CAT	POX
SA	1	**1622.52	**444.91	**3520.30
SNP	2	**3726.52	**71.68	**1487.63
H_2O_2	2	**4075.24	**353.46	**1886.24
SA × SNP	2	**42.74	**9.80	20.52*
$SA \times H_2O_2$	2	13.02 ^{ns}	2.35 ^{ns}	**26.35
$SNP \times H_2O_2$	4	**509.46	3.57 ^{ns}	17.18*
$SA \times SNP \times H_2O_2$	4	16.57 ^{ns}	4.91 ^{ns}	3.91 ^{ns}
Error	36	6.42	2.02	5.70
CV%		5.13	10.05	2.93

ns, *, and **, are non-significant, and significant at 0.05 and 0.01% probability levels, respectively.

SOD (superoxide dismutase), CAT (catalase), POX (peroxidase)

oxidative stress 60 μ M of SNP is recommended to use in soybeans in order to have a better germination and seedling emergence.

References

- Allen, R.D., 1995. 'Dissection of oxidative stress tolerance using transgenic plants'. *Plant Physiology*, **57**: 1049-1054.
- Asadi Karam, E., B. Keramat and H. Mozaffari, 2016. 'Reducing arsenic toxicity stress in soybean (*Glycine max* L.) by using of sodium nitroprusside'. *Journal of Crop Ecophysiology*, 10(1): 225- 242.
- Chance, B. and A.C. Maehly, 1955. Assay of catalases and peroxidases: In: "Methods in Enzymology". Academic Press.
- Durrant, W.E., and X. Dong, 2004. 'Systemic acquired resistance'. *Annual Review in Phytopathology*, 42: 185- 209.

- Fan, H., C. Du and S. Guo, 2012. 'Effect of nitric oxide on proline metabolism in cucumber seedlings under salinity stress'. Journal American Society of Horticultural Science, 137: 127-133.
- Fateh, E., M. Jiriaii, S. Shahbazi and R. Jashni. 2012. 'Effect of salicylic acid and seed weight on germination of wheat (cv. Roshan) under different levels of osmotic stress'. European Journal of Experimental Biology, 5: 1680-1684.
- Hayat, S., and P.S. Ahmad, 2007. 'Salicylic Acid: A Plant Hormone'. Springer, 97-99.
- Hernandez, J.A., M.A. Ferrer, A. Jimenez, A.R. Barcelo and F. Sevilla. 2001. 'Antioxidant systems and O₂⁻/ H₂O₂ production the apoplast of pea leaves. Its relation with saltinduced necrotic lesions in minor veins'. *Plant Physiology*, 127: 827-831.
- MacAdam, J.W., R. Nelson and E. Sharp. 1992. 'Peroxidase activity in the leaf elongation

zone of tall fescue'. *Plant Physiology*, 99: 872-878.

- Mittler, R. 2002. 'Oxidative stress, antioxidant and stress tolerance'. *Annual Review* of *Plant Science*, 7: 405-415.
- Neill, S.J., R. Desikan, A. Clarke, R.D., Hurst and J.T. Hancock. 2002. 'Hydrogen peroxide and nitric oxide as signaling molecules in plants'. *Journal of Experimental Botany*, 53: 1237-1242.
- Neill, S.J., R. Desikan, A. Clarke, R.D., Hurst and J.T. Hancock. 2002. 'Hydrogen peroxide and nitric oxide as signaling molecules in plants'. *Journal of Experimental Botany*, 53: 1237-1242.
- Sedghi, M., M. R. Shakiba, H. Alyari, A. Javanshir and M. Valizadeh. 2005. 'Effect of rhizobia, nitrogen and weeds on soybean (*Glycine max* L.) grain protein and oil'. *Turkish Journal of Field Crops*, 10(2):64-72
- Sen Gupta, A., R. P. Webb, A. S. Holaday and R. D. Allen. 1993. 'Overexpression of superoxide dismutase protects plants from oxidative stress'. *Plant Physiology*, 103: 1067–1073.
- Shi, G.R., Q.S. Cai, Q.Q. Liu and L. Wu. 2009. 'Salicylic acid-mediated alleviation of cadmium toxicity in hemp plants in relation

to cadmium uptake, photosynthesis, and antioxidant enzymes'. *Acta Physiologia Plantarum.* 31: 969–977.

- Singh Gill, S. and N. Tuteja. 2010. 'Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants'. *Plant Physiology and Biochemistry*, 48: 909-930.
- Streeter, T.C., S.M. Rengel, Neate and R.D. Graham. 2001. 'Zinc fertilisation increases tolerance to *Rhizoctonia solani* (AG 8) in *Medicago truncatula'*. *Plant and Soil*, 228: 233-242.
- Tian, X., and Y. Li. 2006. 'Nitric oxide treatment alleviates drought stress in wheat seedlings'. *Plant Biology*, 50: 775-778.
- Wu G.L. and G.Z. Du. 2008. 'Germination is related to seed mass in grasses (Poaceae) of the eastern Qinghai-Tibetan Plateau, China'. *Nordic Journal of Botany*, 25: 361–365.
- Zheng, C., D. Jiang, F. Liu, T. Dai, W. Liu, Q.i. Jing and W. Cao. 2009. 'Exogenous nitric oxide improves seed germination in wheat against mitochondrial oxidative damage induced by high salinity'. *Environmental and Experimental Botany*, 67: 222-227.