

## Probiotics and glycemetic control: A simplified interplay model for the pathways behind

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### ABSTRACT

The link between gut microbiota composition, insulin resistance, and diabetes has been recently proposed. As such, the impact of probiotics on improving glycemetic control has been reported recently. Although probiotics have attracted much interest as a complementary approach to improve glucose metabolism, the mechanisms underlying their actions remained to be determined. Hence, here we aim to review the mechanisms by which the probiotics might affect glycemetic control. Probiotics improve glycemetic control through diminishing fermentation of polysaccharides, suppressing inflammation, act as bile acid de-conjugate hydrolase, increase the bioavailability of Gliclazide drugs and changes in incretin secretion. However, the pathway behind the effect of probiotics on glycemetic control is complex with many interplay interactions. The involvement of multiple mechanisms may explain the ambiguities in determining the exact mechanism that is behind this effect.

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### Introduction

Type 2 diabetes is a major metabolic disorder increasing worldwide. The mechanisms related to type 2 diabetes have been at the center of research in the last decade. In this regards, the gut microbiota is known to play a significant role in type 2 diabetes. The type and count of gut microbiota composition differ between type 2 diabetics and healthy individuals (1). The direct contribution among gut microbiota composition, insulin resistance, and diabetes has been recently proposed (2, 3). As such, the use of probiotics to improve hyperglycemia as an innovative therapeutic approach has been reported (4-6). To date, several randomized controlled trials have been conducted to assess the effects of probiotic on glycemetic control (5, 7-9), however, results are controversial. Probiotics consumption demonstrated beneficial effects on improving glycemetic control among pregnant women (5) and type 2 diabetics (9), while, another randomized controlled trial investigating the efficacy of probiotics (8, 10, 11) described no significant improvement. The impact of probiotics in modulating glucose homeostasis in animal and human studies has been well reviewed in systematic review and meta-analysis study before (4, 12). Altogether, it is well documented that probiotics are able to improve glycemetic control at a modest level. Although

probiotics have attracted much interest as a complementary approach to improve glucose metabolism, the mechanisms underlying their actions remained to be determined. Animal model studies and human clinical trials have attempted to elucidate the mechanism behind this effect (5, 9, 13). In this regard, several pathways were identified to attenuate glycemetic control such as inflammation suppression (13, 14), changes in secretion of gut hormones (15) and changes in the harvest of energy (16). Yet, it is important to keep in mind that the glycemetic modulation is affected by several key elements such as inflammation, insulin resistance and adiposity, which are linked and interact together. Hence, here we aim to review the mechanisms by which the probiotics might affect glycemetic control. Moreover, we attempt to explore an interplay model on the possible interactions between these pathways. This will provide a framework to consider approaches that might help to improve the glycemetic control in type 2 diabetic individuals. The main known mechanisms by which the probiotics affect glycemetic control are as follows:

#### 1. Suppressing inflammation

Type 2 diabetes is an inflammatory disease (17). The direct correlation between the low grade of inflammation and

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pathogenesis of type 2 diabetes was demonstrated (18). This hypothesis opens new horizons in the management of type 2 diabetes. Probiotic supplementation has been shown to suppress inflammation through in vivo studies (13, 14, 19). As such, inflammation suppression has been presented as the first step in recognizing a role for probiotics in managing glycemic control. Reduction in level of inflammatory mediators activates signaling pathways resulting in insulin resistance and improve glycemic control (20-22). The effects of probiotics on glycemic control through modulation of inflammation are classified into two modes of action: a) indirectly, through improving the gut integrity, and b) directly by functioning as an antioxidant. These mechanisms are highlighted in the following paragraphs.

## 2. Suppressing inflammation through the improving gut barrier

Several aspects of the modern lifestyle such as high fat diet (23), high carbohydrate diet and sedentary lifestyle have the potential to change the composition of gut microbiota toward being Firmicutes-dominant (24, 25). These phenotypes decrease gut integrity and increase gut permeability, promoting freeing lipopolysaccharides (LPS) and free fatty acids (FFA) in the blood and peripheral tissues (26, 27). LPS and FFA are detected by toll-like receptors (TLR) at the surface of immune cells including T lymphocyte, monocyte, and macrophage (28). TLR comprise a family of cell surface protein receptors that their activation leads to induction of inflammatory responses and this interaction promotes the release of inflammatory markers such as Tumor Necrosis factor  $\alpha$  (TNF- $\alpha$ ) and interleukins from immune cells (28). Inflammatory cytokines, especially TNF- $\alpha$  has been shown to inhibit the insulin signaling pathway and supposed to be the primary trigger for insulin resistance mechanism (25). Moreover, blood FFA is being transported in adipose tissue and leads to either hypertrophy or hyperplasia of adipose cells. The hypertrophic/hyperplastic adipose tissue has been shown to express high levels of biochemical substances. These substances have the ability to recruit monocyte-macrophages toward the hypertrophic fat tissue and attenuate local inflammatory mechanisms (17). Administration of probiotics has been shown to regulate gut permeability by stimulating the secretion of host gut peptides such as glucagon-like peptide 2 (GLP-2). GLP-2 is responsible for the effective protection of gut integrity (29). Selective gut microbiota structure controls and increases endogenous GLP-2 production, which consequently improves gut barrier functions by a GLP-2-dependent mechanism and contributes to the improvement of gut barrier functions (27). Improving gut permeability through probiotic administration has been associated with decreased serum levels of TNF- $\alpha$  and other cytokines such as interleukin-6 (30, 31). Reducing the levels of inflammatory markers decreases insulin resistance and improves insulin sensitivity, which is inversely related to B-cells function.  $\beta$ -cell function improvement promotes insulin secretion and resulting in better

glycemic control (32).

## 3. Suppressing inflammation through antioxidant-like activities

Oxidative stresses play essential roles in the pathogenesis and progression of diabetes (33). The total antioxidant status of type 2 diabetes is lower than healthy individuals (34). Free radicals are produced in an excessive amount in type 2 diabetic individuals. Free radicals cause lipid peroxidation and produce inflammatory cytokines (35). Different strains of probiotics have been reported to act as antioxidant and repress oxidative stress (36-38). The antioxidative mechanisms of probiotics can be contributed to reactive oxygen species scavenging, metal ion chelation, enzyme inhibition, and the reduction activity and inhibition of ascorbate autoxidation (39). In the human model of type 2 diabetes, multistrain probiotic supplementation has shown to increase total antioxidant status, as well as serum levels of different antioxidants such as erythrocyte glutathione, reeducates, erythrocyte superoxide dismutase and glutathione peroxidase (37, 38). Hence, probiotics can scavenge free radicals and suppress inflammation, which leads to improving insulin resistance and glycemic control.

## 4. Changing the gut hormones

It has been well established that the structure of gut microbiota is directly related to the secretion of GLP-1 (40, 41), and GLP-2 (27, 40). GLP1 is a peptide which originates from enteroendocrine cells of the gut and usually circulating in the blood in the form of GLP17-36 amide (42). GLP1 exerts multiple physiological actions include stimulation of insulin secretion, decreasing hunger and controlling energy intake, as well as decreasing gluconeogenesis which lead to control energy intake and glycemic control. These functions (42). GLP2 is a 33-amino acid peptide secreted with GLP1 from enteroendocrine cells in a nutrient-dependent manner. GLP2 rapidly induces hexose transport in jejunal basolateral membrane vesicles leading to the expansion of the mucosal epithelium in the small bowel (43), making it a suitable candidate for maintaining the gut integrity. Considering the link between gut microbiota structure and secretion of gut hormones, manipulation of gut microbiota by probiotics could efficiently alter the secretion pattern of the gut hormones. The pathway in which GLP-2 hormone is involved was earlier discussed in improving gut barrier function and suppressing inflammation. Here, we continue to explore the role of GLP-1 in affecting glycemic control. Short chain fatty acids (SCFAs) driven from gut microbiota affect proliferation, differentiation, and modulation of gene expression in colonic epithelial cells (44). In addition, SCFAs can regulate gene expression by binding to the G-protein-coupled receptors. Signaling through these receptors affects the secretion of the GLP-1 which improves insulin secretion (15, 45). Improving insulin secretion directly will affect glycemic control. In addition, GLP-1 decreases hunger and increasing satiety which leads to

decreasing energy intake and improving glycemic control (40, 46). The GLP-1 hormone also improves glycemic control by inhibiting the gluconeogenesis leading to reduction of monosaccharide flow into the bloodstream which ultimately improves glycemic control (47). In conclusion, probiotic administration modulates gut microbiota composition in favor of GLP-1 and GLP-2 secretion. Secretion of these hormones affects glycemic control via different pathways. Modulation of other types of gut peptides involved in appetite regulation, such as leptin and peptide YY, could be another mechanism by which the gut microbiota might control energy and glucose homeostasis (29). However, there is limited evidence regarding this link in the context of metabolic modulation of glucose control.

### 5. Manipulation of the harvest of energy

Carbohydrates are important sources of energy for human and microbial cells. Human enzymes cannot degrade more complex carbohydrates and plant polysaccharides. These carbohydrates are fermented in the colon by its microbiota to yield energy for microbial growth and end products such as SCFAs (48). Animal model studies revealed that the gut microbiota capacity for energy harvest is higher in obese as compared with lean (49). For example, fermentation of dietary fructans increases when mice have been colonized with probiotics from *Bacteroides* phylum (50). Colonization of germ-free mice with obese microbiota results in a significantly greater increase in total body fat than colonization with a lean microbiota (49).

These interactions promote carbohydrate fermentation more efficiently and increase energy absorption from the gut, resulting in more adiposity. In a human study, the fecal microbiota of obese individuals has an increased capacity to harvest energy (49). The role of the gut microbiota in promoting energy harvest from diet and fat deposition has been demonstrated in mice (16), but most of the evidences in humans have come from indirect studies. Therefore, it can be hypothesized that manipulating the gut microbiota by probiotics changes the harvest of energy in favor of decreasing adiposity (16). Less adipocyte means less inflammation and insulin resistance, which leads to better glycemic control. However, changes in the harvest of energy can affect glycemic control through a more direct pathway. The new gut microbiota structure developed by beneficial microbes has less capacity for fermenting dietary polysaccharides, indigestible by human enzymes. So, less polysaccharide will be added to the pool of gastrointestinal absorbable glucose and will positively affect glycemic control (51). Although many theoretical hypotheses proposed the role of probiotics in the harvest of energy, the changes in the energy elucidation pattern after probiotic consumption in the context of a clinical trial remained to be determined. Thus, whether the subtle theoretical explanation can translate to clinically meaningful

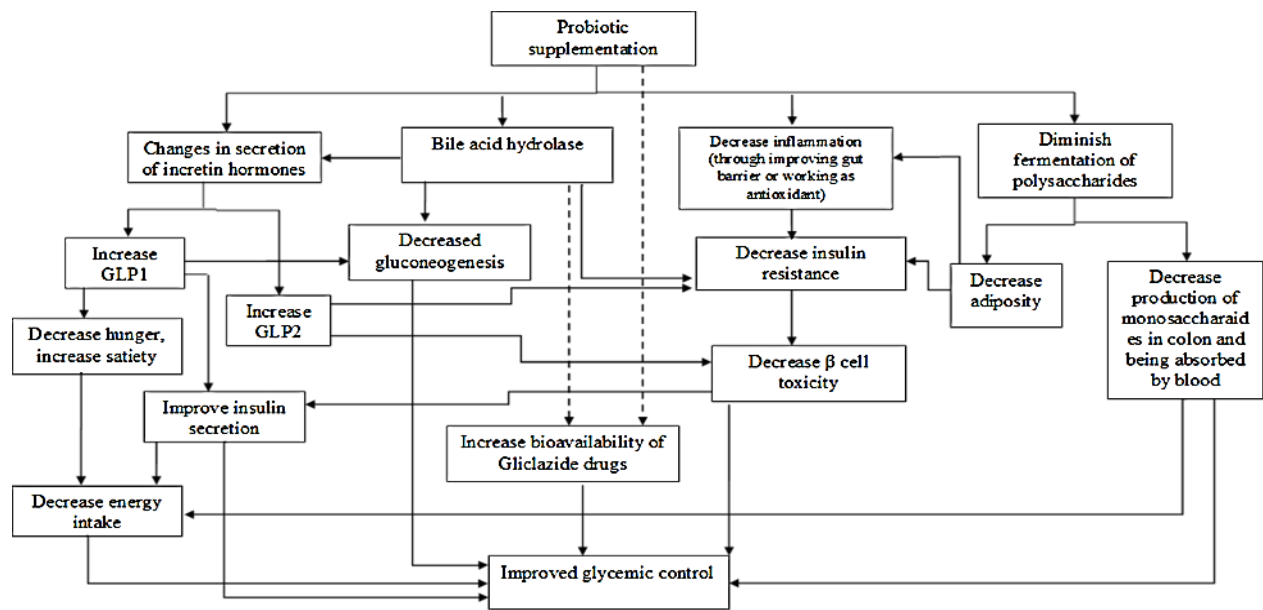
outcomes remains elusive.

### 6. Bile acid deconjugation and activation of Farnesoid X receptor

It is well recognized that some members of the gut microbiota, mostly *Lactobacilli* and *Bifidobacteria*, are known to possess the bile salt hydrolase enzyme and have the ability to de-conjugate bile salts (52, 53). The action of de-conjugated bile acids activates several bile acid signaling pathways, most importantly, nuclear Farnesoid X Receptor (FXR) (54). The FXR is a member of the nuclear receptor family, which is primarily expressed in liver, kidney, and intestine. FXR has a predominant regulatory role in glycemic control. FXR- null mice demonstrated high blood glucose and insulin resistance while activation of it suppressed gluconeogenesis and decreased blood glucose (55). Activation of FXR improves glycemic control through three different ways. (a) Repressing expression of gluconeogenic genes and increasing glycogenesis, which results in serum glucose reduction (55, 56). (b) Decreasing FFA and improving peripheral insulin resistance (55, 57) that ultimately lead to improved glycemic control. (c) Bile acids also are responsible for the secretion of incretins including GLP-1 and GLP-2 (58) which can modulate glycemic control through controlling hunger, stimulating insulin secretion, suppressing gluconeogenesis as well as reducing inflammation (These mechanisms have been discussed in detail earlier in "changing gut hormones" session).

### 7. Improving the bioavailability of Gliclazide drugs

Another pathway for the hypoglycemic activity of probiotics can be attributed to their ability to increase the bioavailability of Gliclazide drug (59). Al-Salami et al., measured Gliclazide serum concentration on healthy and Alloxan induced type 1 diabetic rats supplemented with probiotics (59). They found that probiotic supplementation increased the bioavailability of Gliclazide in diabetic rats -with an unknown mechanism- when compared with the control group. Increased bioavailability increases the time of action for Gliclazide. Although in their study this increase did not lead to glycemic control, the changes in Gliclazide bioavailability potentially may improve glycemic control. However, the manipulation of Gliclazide bioavailability by probiotics should be interpreted with caution and considered hypothesis-generating results rather than a firm indication of the beneficial effects of the probiotic treatment. It is worth to mention that, there is no data to show whether probiotic supplements directly affect the bioavailability of Gliclazide or this effect is modulated by the bile acid hydrolase function of probiotic. In addition, it is unclear whether data obtained from an animal model of Alloxan induced type 1 diabetes can be extrapolated to human type 2 diabetes.



**Fig. 1.** Proposed mechanism of action. GLP1: Glucagon-like peptide 1, GLP2: Glucagon-like peptide 2, Dash lines has been proven in animal model of type 1 diabetes.

## Conclusion

According to the above data, the pathway behind the effect of probiotics on glycemic control is complex with various interactions. These interactions are demonstrated in Fig. 1. As this pathway includes numerous interactions at the cellular and molecular level which most of them are still enigma, we attempted not to get involved at this level. The involvement of multiple mechanisms may explain the ambiguities in determining the exact mechanism that is behind this effect. There is also another mechanism involve such as changes in gene transcription which all remains to be elucidated by the future researches. However, focusing on this model will help researchers to clarify and investigate more precisely the underlying mechanisms of actions.

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## References

- Larsen N, Vogensen FK, van den Berg FWJ, et al. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. *PLOS One*. 2010;5(2):e9085.
- Caricilli AM, Picardi PK, de Abreu LL, et al. Gut microbiota is a key modulator of insulin resistance in TLR 2 knockout mice. *PLOS Biology*. 2011;9(12):e1001212.
- Musso G, Gambino R, Cassader M. Obesity, diabetes, and gut microbiota. The hygiene hypothesis expanded? *Diabetes Care*. 2010;33(10):2277-2284.
- Firouzi S, Barakatun-Nisak MY, Ismail A, Majid HA, Nor Azmi K. Role of probiotics in modulating glucose homeostasis: evidence from animal and human studies. *International Journal of Food Sciences and Nutrition*. 2013;64(6):780-6.
- Laitinen K, Poussa T, Isolauri E. Probiotics and dietary counselling contribute to glucose regulation during and after pregnancy: a randomised controlled trial. *British Journal of Nutrition*. 2009;101(11):1679-1687.
- Andreasen AS, Larsen N, Pedersen-Skovsgaard T, et al. Effects of *Lactobacillus acidophilus* NCFM on insulin sensitivity and the systemic inflammatory response in human subjects. *British Journal of Nutrition*. 2010;104(12):1831-1838.
- Asemi Z, Zare Z, Shakeri H, Sabihi S-S, Esmailzadeh A. Effect of multispecies probiotic supplements on metabolic profiles, hs-CRP, and oxidative stress in patients with type 2 diabetes. *Annals of Nutrition and Metabolism*. 2013;63(1-2):1-9.
- Ivey KL, Hodgson JM, Kerr D a, Lewis JR, Thompson PL, Prince RL. The effects of probiotic bacteria on glycaemic control in overweight men and women: a randomised controlled trial. *European Journal of Clinical Nutrition*. 2014;68(4):447-52.
- Ejtahed HS, Mohtadi Nia J, Homayouni Rad A, Niafar M, Asghari Jafarabadi M, Mofid V. The effects of probiotic and conventional yoghurt on diabetes markers and insulin resistance in type 2 diabetic patients: A randomized controlled clinical trial. *Iranian Journal of Endocrinology & Metabolism*. 2011;13(1):1-8.
- Gobel RJ, Larsen N, Jakobsen M, Molgaard C, Michaelsen KF. Probiotics to obese adolescents; RCT examining the effects on inflammation and metabolic syndrome. *Journal of Pediatric Gastroenterology and Nutrition*. 2012;5(6):673-8.
- Lindsay KL, Kennelly M, Culliton M, et al. Probiotics in obese pregnancy do not reduce maternal fasting glucose: a double-blind, placebo-controlled, randomized trial (probiotics in pregnancy study). *The American Journal of Clinical Nutrition*. 2014;99(6):1432-9.
- Ruan Y, Sun J, He J, Chen F, Chen R, Chen H. Effect of probiotics on glycemic control: A systematic review and meta-analysis of randomized, controlled trials. *PLOS One*. 2015;10(7):e0132121.
- Zarfeshani A, Khaza'ai H, Mohd Ali R, Hambali Z, Wahle KWJ, Mutalib MSA. Effect of *Lactobacillus casei* on the production of pro-inflammatory markers in Streptozotocin-induced diabetic rats. *Probiotics Antimicrob Proteins*. 2011;3(3-4):168-74.
- Naito E, Yoshida Y, Makino K, et al. Beneficial effect of oral administration of *Lactobacillus casei* strain *Shirota* on insulin resistance in diet-induced obesity mice. *Journal of Applied Microbiology*. 2011;110:650-7.
- Yadav H, Lee J, Lloyd J, Walter P, Rane S. Beneficial metabolic effects of a probiotic via Butyrate-induced GLP-1 hormone secretion. *Journal of Biological Chemistry*. 2013;288(35):25088-25097.
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon



- Ji. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*. 2006;444(7122):1027-31.
17. Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. *Nat Rev Immunol*. 2011;11(2):98-107.
  18. Pickup JC. Inflammation and activated innate immunity in the pathogenesis of type 2 diabetes. *Diabetes Care*. 2004;27(3):813-23.
  19. Valladares R, Sankar D, Li N, et al. *Lactobacillus johnsonii* N6. 2 mitigates the development of type 1 diabetes in BB-DP rats. *PLoS One*. 2010;5(5):e10507.
  20. Cani PD, Amar J, Iglesias MA, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007;56(7):1761-72.
  21. Shoelson SE, Lee J, Goldfine AB. Inflammation and insulin resistance. *Journal of Clinical Investigation*. 2006;116(7):1793.
  22. Vandanmagsar B, Youm Y-H, Ravussin A, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nature Medicine*. 2011;17(2):179-88.
  23. Ding S, Chi MM, Scull BP, et al. High-fat diet: bacteria interactions promote intestinal inflammation which precedes and correlates with obesity and insulin resistance in mouse. *PLoS One*. 2010;5(8):e12191.
  24. Parnell JA, Reimer RA. Prebiotic fibres dose-dependently increase satiety hormones and alter Bacteroidetes and Firmicutes in lean and obese JCR:LA-cp rats. *British Journal of Nutrition*. 2012;107(4):601-13.
  25. Escobedo G, López-Ortiz E, Torres-Castro I. Gut microbiota as a key player in triggering obesity, systemic inflammation and insulin resistance. *Revista de Investigación Clínica*. 2015;66(5):450-9.
  26. Cani PD, Bibiloni R, Knauf C, et al. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes*. 2008;57(6):1470-81.
  27. Cani PD, Possemiers S, Van de Wiele T, et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut*. 2009;58(8):1091-103.
  28. Youssef-Elabd EM, McGee KC, Tripathi G, et al. Acute and chronic saturated fatty acid treatment as a key instigator of the TLR-mediated inflammatory response in human adipose tissue, in vitro. *Journal of Nutritional Biochemistry*. 2012;23(1):39-50.
  29. Everard A, Cani PD. Diabetes, obesity and gut microbiota. *Best Practice & Research: Clinical Gastroenterology*. 2013;27(1):73-83.
  30. Imani Fooladi AA, Mahmoodzadeh Hosseini H, Nourani MR, Khani S, Alaviani SM. Probiotic as a novel treatment strategy against liver disease. *Hepatitis Monthly*. 2013;13(2):e7521.
  31. Everard A, Matamoros S, Geurts L, Delzenne NM, Cani PD. *Saccharomyces boulardii* administration changes gut microbiota and reduces hepatic steatosis, low-grade inflammation, and fat mass in obese and type 2 diabetic db/db mice. *mBio*. 2014;5(3):e01011-14.
  32. Bergman RN. Toward physiological understanding of glucose tolerance: Minimal-model approach. *Diabetes*. 1989;38(12):1512-1527.
  33. Henriksen EJ. Bioactive food as dietary interventions for diabetes. *Elsevier*; 2013.
  34. Opara EC, Abdel-Rahman E, Soliman S, et al. Depletion of total antioxidant capacity in type 2 diabetes. *Metabolism*. 1999;48(11):1414-1417.
  35. Maritim AC, Sanders RA, Watkins JB. Diabetes, oxidative stress, and antioxidants: a review. *Journal of Biochemical and Molecular Toxicology*. 2003;17(1):24-38.
  36. Uskova MA, Kravchenko L V. Antioxidant properties of lactic acid bacteria--probiotic and yogurt strains. *Vopr Pitan*. 2009;78(2):18-23.
  37. Ejtahed HS, Mohtadi Nia J, Homayouni Rad A, Niafar M, Asghari Jafarabadi M, Mofid V. Probiotic yogurt improves antioxidant status in type 2 diabetic patients. *Nutrition*. 2012;28(5):539-43.
  38. Asemi Z, Jazayeri S, Najafi M, et al. Effect of daily consumption of probiotic yogurt on oxidative stress in pregnant women: a randomized controlled clinical trial. *Annals of Nutrition and Metabolism*. 2012;60(1):62-8.
  39. Lin M-Y, Yen C-L. Antioxidative ability of lactic acid bacteria. *Journal of Agricultural and Food Chemistry*. 1999;47(4):1460-1466.
  40. Delzenne NM, Neyrinck AM, Cani PD. Modulation of the gut microbiota by nutrients with prebiotic properties: consequences for host health in the context of obesity and metabolic syndrome. *Microb Cell Fact*. 2011;10(Suppl 1):S10.
  41. Zhou J, Martin RJ, Tulley RT, et al. Dietary resistant starch upregulates total GLP-1 and PYY in a sustained day-long manner through fermentation in rodents. *American Journal of Physiology-Endocrinology and Metabolism*. 2008;295(5):E1160-E1166.
  42. Orskov C, Rabenhøj L, Wettergren A, Kofod H, Holst JJ. Tissue and Plasma Concentrations of Amidated and Glycine-Extended Glucagon-Like Peptide I in Humans. *Diabetes*. 1994;43(4):535-539.
  43. Boushey RP, Yusta B, Drucker DJ. Glucagon-like peptide (GLP)-2 reduces chemotherapy-associated mortality and enhances cell survival in cells expressing a transfected GLP-2 receptor. *Cancer Research*. 2001;61(2):687-93.
  44. Davie JR. Inhibition of histone deacetylase activity by butyrate. *Journal of Nutrition*. 2003;133(7 Suppl):2485S-2493S.
  45. Tolhurst G, Heffron H, Lam Y, et al. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. *Diabetes*. 2012;61(2):364-71.
  46. Flint A, Raben A, Astrup A, Holst JJ. Glucagon-like peptide 1 promotes satiety and suppresses energy intake in humans. *Journal of Clinical Investigation*. 1998;101(3):515-20.
  47. Lee Y-S, Shin S, Shigihara T, et al. Glucagon-like peptide-1 gene therapy in obese diabetic mice results in long-term cure of diabetes by improving insulin sensitivity and reducing hepatic gluconeogenesis. *Diabetes*. 2007;56(6):1671-9.
  48. Bergman EN. Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiological Reviews*. 1990;70(2):567-90.
  49. Turnbaugh PJ, Hamady M, Yatsunenkov T, et al. A core gut microbiome in obese and lean twins. *Nature*. 2009;457(7228):480-4.
  50. Samuel BS, Gordon JI. A humanized gnotobiotic mouse model of host-archaeal-bacterial mutualism. *Proceedings of the National Academy of Sciences of the United States of America*. 2006;103(26):10011-6.
  51. Zhang H, DiBaise JK, Zuccolo A, et al. Human gut microbiota in obesity and after gastric bypass. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;106(7):2365.
  52. Degirolamo C, Rainaldi S, Bovenga F, Murzilli S, Moschetta A. Microbiota modification with probiotics induces hepatic bile acid synthesis via downregulation of the Fxr-Fgf15 axis in mice. *Cell Reports*. 2014;7(1):12-8.
  53. Pavlović N, Stankov K, Mikov M. Probiotics-interactions with bile acids and impact on cholesterol metabolism. *Applied Biochemistry and Biotechnology*. 2012;168(7):1880-95.
  54. Mencarelli A, Distrutti E, Renga B, et al. Probiotics modulate intestinal expression of nuclear receptor and provide counter-regulatory signals to inflammation-driven adipose tissue activation. *PLoS One*. 2011;6(7):e22978.
  55. Ma K, Saha PK, Chan L, Moore DD. Farnesoid X receptor is essential for normal glucose homeostasis. *Journal of Clinical Investigation*. 2006;116(4):1102-9.
  56. Thomas C, Pellicciari R, Pruzanski M, Auwerx J, Schoonjans K. Targeting bile-acid signalling for metabolic diseases. *Nature Reviews Drug Discovery*. 2008;7(8):678-93.
  57. Cariou B, van Harmelen K, Duran-Sandoval D, et al. The farnesoid X receptor modulates adiposity and peripheral insulin sensitivity in mice. *Journal of Biological Chemistry*. 2006;281(16):11039-49.
  58. Parker HE, Wallis K, le Roux CW, Wong KY, Reimann F, Gribble FM. Molecular mechanisms underlying bile acid-stimulated glucagon-like peptide-1 secretion. *British Journal of Pharmacology*. 2012;165(2):414-23.
  59. Al-Salami H, Butt G, Fawcett JP, Tucker IG, Golocorbin-Kon S, Mikov M. Probiotic treatment reduces blood glucose levels and increases systemic absorption of gliclazide in diabetic rats. *European Journal of Drug Metabolism and Pharmacokinetics*. 2008;33(2):101-106.