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ORIGINAL ARTICLE

Assessment of the Antioxidant Potential of Methanolic Extracts from *Leontodon lanata* (L.) Fisch, *Convolvulus arvensis* L., and *Ziziphora capitata* L.

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ABSTRACT: Synthetic antioxidants are widely used today to mitigate oxidative stress and counter the harmful effects of free radicals. However, due to the potential side effects associated with synthetic antioxidants, there is growing interest in natural alternatives. The antioxidant properties of medicinal plants offer a compelling rationale for their application in the food, pharmaceutical, and personal care industries. Since plant-based foods, particularly fruits and vegetables, are the richest sources of antioxidants, this study aimed to investigate the antioxidant properties of methanolic extracts from *Leontodon lanata* (L.) Fisch, *Convolvulus arvensis* L., and *Ziziphora capitata* L. *L. lanata* was harvested from the Ilam region, while C. arvensis and Z. capitata were collected from the Andimeshk to Dehloran area. The total antioxidant capacity of these plants was assessed using the ferric reducing antioxidant power (FRAP) assay. The results revealed that *L. lanata* exhibited the highest antioxidant activity with a capacity of 3.87 mmol Fe²⁺ L⁻¹. In comparison, the antioxidant capacities of *C. arvensis* and *Z. capitata* were measured at 1.29 mmol Fe²⁺ L⁻¹ and 1.69 mmol Fe²⁺ L⁻¹, respectively. Among the plants tested, *L. lanata* demonstrated the most potent antioxidant effect. These findings suggest that methanolic extracts of *L. lanata*, *C. arvensis*, and *Z. capitata*, particularly *L. lanata*, could be valuable as natural supplements in the food and pharmaceutical industries, supporting health and therapeutic applications.

INTRODUCTION

Free radicals are generated through natural reactions during cellular respiration in aerobic organisms, especially in animals [1]. Free radicals, such as reactive oxygen species (ROS), are naturally produced in different metabolic pathways in cells and play a key role in signaling process of cells. The body may generate free radicals in response to ultraviolet rays, tobacco smoke, and alcoholic drinks [2]. High amounts of free radicals are dangerous for cells and may cause damage to cell

components like proteins, DNA, phospholipids, and cell membranes, leading to tissue destruction and different diseases, such as inflammation, cancer, etc [3]. Low level of antioxidants in the body leads to early aging, damaged or mutated cells, damaged tissue, and activation of faulty genes in DNA; and it causes an increased pressure on immune system function [4].

Foods containing antioxidants and food antioxidants are an important part of the food industry. In the past,

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antioxidants were primarily used to control oxidation and mitigate damage, but today many are used for their health benefits [3]. In the cosmetic industry, antioxidants are used as ingredients that help protect the skin against oxidative damage and free radicals. These substances aim to improve the appearance and quality of the skin, reduce the signs of aging and maintain the health of the skin. Antioxidants can be obtained from various sources such as vitamins (such as vitamin C and E), minerals, plant extracts and other nutritional values [2-4]. Antioxidants that must be obtained from external sources include vitamins A, C, and E, along with beta-carotene, lycopene, lutein, selenium, manganese, and zeaxanthin. The richest sources of these antioxidants are plant-based foods, particularly fruits and vegetables. Additionally, plant foods contain other antioxidant compounds such as flavonoids, flavones, catechins, polyphenols, and phytoestrogens, which also serve as important phytonutrients. Foods abundant in antioxidants are often referred to as "superfoods" or "functional foods" due to their health-promoting properties [2-4]. The antioxidant activity of plant extracts has been studied for many years; however, most of the studies have been conducted over the past 30 years. During this period, research studies have been conducted on medicinal plants with traditional usage [5]. Antioxidants are important compounds capable of protecting cells against oxidation and are effective in preventing and treating the oxidative damage [6]. The important antioxidants contained in tocopherol include folic acid, ascorbic acid, carotenoid pigments, phenylacrylic acids, and polyphenols, namely flavonoids and anthocyanins that constitute the largest group of natural phenols [7]. Some antioxidants are made by the body itself, and others are provided through feeding and eating foods containing antioxidants, which may also possess anti-inflammatory properties [8]. The most important sources of antioxidants are medicinal plants, fruits, and vegetables. Antioxidants are found in different compounds like carotenoids (e.g., betacarotene), lycopene, and vitamin C. These compounds are some examples of antioxidants that inhibit the oxidation process or cell reaction to oxygen, peroxide, or free radicals [8].

Considering the unwanted side effects of artificial antioxidants, this study aims to investigate the effect of

medicinal plants such as *Leontodon lanata* (L.) Fisch, *Convolvulus arvensis* L and *Ziziphora capitata*, which are useful and anti-inflammatory medicinal plants in traditional medicine and ethnobotanical knowledge of western Iran. They are oxidants and were studied to prove their antioxidant effect.

Desert ivy (Convolvulus arvensis) is a perennial herbaceous plant with branches up to 100 mm long, broad branches and branched stems, herbaceous leaves, ovate petioles, thread-like peduncles, membranous sepals, ovate ovary and white or pink calyx with Dark red streaks. Convolvulus arvensis is an herbaceous perennial plant in the Convolvulaceae family that is native to Europe and Asia. This plant is laxative, antispasmodic, effective in wound treatment, and has antihemorrhoid, antioxidant, anti-chromium and anti-tumor properties [9]. Z. capitata is an annual herb in the family Lamiaceae. This plant has a stem of 5-25 cm, standing, hard and brittle, leaves 4-35 mm long, leaves next to the flowers with various shapes. This plant is an antiflatulence, disinfectant, wound healer, sedative, expectorant, stomach tonic, gastric problems remover, and a body lice killer with external use [10].

Hairy horsehair herb (*Leontodon lanata* (L.)) without hair or spider hair and cabbage green with a stem height of up to 30 cm, standing, deeply branched. The leaves are fresh fleshy and have tabular and purple flowers. *L. lanata* (L.) Fisch is in the Compositae family. This plant is an edible vegetable with antimicrobial and antioxidant effects [11]. The present study aims to evaluate the antioxidant effect of methabolic extract of medicinal plants *L. lanata* (L.) Fisch, *C. arvensis* L, and *Z. capitata*.

MATERIALS AND METHODS

Plant preparation

Medicinal plants *L. lanata* (L.) Fisch, *C. arvensis* L, and *Z. capitata* was collected in June, 2023. This plant is native to Iran and the provinces of Ilam and Khuzestan. The plant *L. lanata* was collected from Ilam and two medicinal plants *C. arvensis* L. and *Z. capitata* L. were collected from Andimeshk to Dehlaran (60 km from Dehlran). Using the morphological keys from the book of plant flora by the botanist of the research center for

biotechnology and medicinal plants and natural resources of Ilam city to identify the medicinal plants. The collected plants were cleaned and shade-dried with full air circulation. The dried plant was powdered using a mixer grinder and used for an antioxidant activity test. Table 1 shows the specifications of the above-mentioned medicinal plants used in this study.

Table 1. Specifications of the studied medicinal plants.

Plant Scientific name	Herbal family	Collection region	Herbarium code	The geographical location
Leontodon lanata (L.) Fisch	Asteraceae	Ilam, Ilam	187	33.638531°N 46.422649°E
Convolvulus arvensis L.	Convolvulaceae	Khuzestan, Andimeshk	265	32°24′52″N 47°49′24″E
Ziziphora capitata L.	Lamiaceae	Khuzestan, Andimeshk	598	32°24′52″N 47°49′24″E

Determination of Antioxidant Activity of Methanolic

Extracts

Preparation of Plant Sample

To prepare the plant sample, 1 gram of the homogenized dry plant powder was shaken for 6 hours with 100 mL of methanol. The resulting solution was transferred to a plastic falcon tube and centrifuged at 6000 rpm for 10 minutes. The supernatant obtained after centrifugation was used as the sample [12].

Preparation of Working Solution

For the working solution, 2.2 mL of reactive substance 2b (R2b) was added to the parent solution of reactive substance 2a (R2a) and vortexed until fully dissolved, forming the reactive solution 2 (R2). This solution was then mixed with 5 times its volume of R1 solution after vortexing. The resulting mixture served as the working solution for the antioxidant assay kit [12].

Preparation of Standard Solution

A standard solution was prepared with concentrations of 0, 0.2, 0.4, 0.6, 0.8, and 1 mL. The linear equation was derived based on the absorbance of Fe²⁺ at varying concentrations [12]. The total antioxidant capacity was

assessed through the ferric reducing antioxidant power (FRAP) method, which measures the ability of the sample to reduce ferric ions, following a single electron transfer mechanism.

Method

In this assay, 5 microliters of the prepared plant extract solution were added to each well of the microplate, followed by the addition of 250 μ L of the working solution to each well. The plate was then incubated for 30 minutes at temperatures ranging between 35 and 50°C. The antioxidant activity was determined by measuring absorbance at 570 nm using an ELISA reader [12].

RESULTS

The results of the antioxidant activity of the plants' extract showed the standard antioxidant activity to be 1 mmol Fe^{2+} L⁻¹ [12]. As shown in Table 2, all medicinal plants studied in this research had antioxidant effects.

Table 2. Total antioxidant capacity of methanolic extract of medicinal plants analyzed in this study

Methanolic extract	Total antioxidant capacity		
Leontodon lanata (L.) Fisch	3.87 mmol Fe ²⁺ L ⁻¹		
Convolvulus arvensis L.	1.29 mmol Fe ²⁺ L ⁻¹		
Ziziphora capitat L.	1.69 mmol Fe ²⁺ L ⁻¹		

According to the obtained results, the methanolic extract of *L. lanata* (L.) Fisch with an antioxidant capacity of

 $3.87 \text{ mmol Fe}^{2+} L^{-1}$ had the strongest antioxidant activity amongst medicinal plants studied in this research.

DISCUSSION

Antioxidants are essential molecules that protect the body from free radicals, which can contribute to the development of diseases such as diabetes and cancer. Studies have revealed that medicinal plants like Ferula assa-foetida (both bitter and sweet varieties) and Bunium persicum exhibited antioxidant capacities of 1.18, 1.09, and 1.75 mmol Fe2+ L-1, respectively [12]. Another investigation showed that methanolic extracts of Ziziphus spina-christi, Salvia rosmarinus, and Satureja khuzistanica displayed antioxidant potentials of 7 mmol Fe²⁺ L⁻¹, 1.93 mmol Fe²⁺ L⁻¹, and 12.99 mmol Fe²⁺ L⁻¹, respectively [13]. Similarly, total antioxidant capacities for Anthemis susiana, Alyssum campestre, and Gundelia tournefortii were recorded as 4.29, 1.01, and 1.25 mmol Fe²⁺ L⁻¹, respectively [14]. Research on the antioxidant activity of Allium ampeloprasum demonstrated a total antioxidant capacity of 3.06 mmol Fe2+ L-1, confirming its significant antioxidant properties [15]. Using the FRAP assay, another study measured total antioxidant capacities of 2.21, 0.78, and 7 mmol Fe2+ L-1 for Mentha longifolia, Pistacia khinjuk, and Eucalyptus globulus, respectively [16]. An investigation into Nasturtium officinale revealed its total antioxidant capacity to be 2.83 mmol Fe2+ L-1 [17]. Further analysis found antioxidant capacities of 4.1, 2.35, and 0.46 mmol Fe²⁺/L for Quercus brantii, Thymbra spicata, and Citrullus colocynthis, respectively [18]. Additionally, Falcaria vulgaris displayed a total antioxidant capacity of 2.86 mmol Fe2+ L-1 [19]. Finally, the methanolic extracts of Cynara scolymus, Echinacea purpurea, and Portulaca oleracea demonstrated antioxidant capacities of 3.45, 1.16, and 1.68 mmol Fe^{2+} L⁻¹, respectively [20]. This study found the medicinal plant extracts with antioxidant properties to have high levels of phenolic compounds [21]. Plant extracts contain different antioxidant compounds including polyphenols. Previous studies have shown the abundant biological activities of these compounds, especially flavonoids. According to these studies, most of the properties of plant species extracts can be assigned to these compounds [22, 23]. Convolvulus arvensis contains main compounds, including flavonoids, steroids, phenols, lipids, coumarins, campesterol, stigma-sterol, and betasitosterol as the most abundant steroid compound [24]. The results of a study showed that Z. capitata contained chemical compounds, including pulegone, limonene, 1,8cineole, p-mentha-3-en-8-ol, neomenthol, bornyl acetate, and piperitone [25]. Sesquiterpene lactone and its glycosides are among the main components of L. lanata [26]. The results of phytochemical studies showed that medicinal plants of the Leontodon genus contained phenolic compounds [27, 28]. As medicinal plants containing phenolic and polyphenolic compounds have antioxidant effects, studied medicinal plants also contained phenolic and polyphenolic compounds, they have antioxidant properties, too. These compounds have hydroxyl groups in their chemical structure, which act as antioxidant agents [29]. Polyphenols have high reducing power. They can react with free radicals of oxygen and nitrogen and reduce oxidative stress, and finally, decrease the damage to the body cells and tissues [30]. These antioxidants mostly are safe and can prevent or treat diseases such as diabetes mellitus [31, 32]. Hence, the daily consumption of plants with high levels of antioxidant activity might be useful in these patients. Antioxidants act as absorbers of free radicals and thus protect the body from damage caused by the presence of free radicals [30-32]. Antioxidants may also boost the immune system and lower the risk of various diseases and infections [33-35]. These antioxidant compounds, through mechanisms such as binding to oxidant molecules or neutralizing free radicals, help diminish the harmful effects of oxidants that contribute to chronic conditions like diabetes, cardiovascular diseases, and kidney disorders, while mitigating their destructive potential [36-43]. Since free radicals are crucial contributors to the development of numerous illnesses, antioxidant agents can play a significant role in both the prevention and treatment of such health issues.

CONCLUSIONS

Today, natural antioxidants are healthier and safer than synthetic ones and are not restricted to terrestrial sources and seaweeds are a rich source of antioxidant compounds. As the medicinal plants studied in this research have natural antioxidants, metabolic extract of

the above-mentioned medicinal plants has antioxidant properties and can be used as food or therapeutic supplements if preclinical and clinical studies are approved.

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Conflict of interests

The authors declare no competing interest.

Consent for publication

Not applicable.

Availability of data and materials

Not applicable.

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REFERENCES

- 1. Emami B., Shakerian A., Sharafati Chaleshtouri R., Rahimi E. 2024. Antioxidant, antimicrobial, and anticancer effects of the Russian olive (*Elaeagnus angustifolia* L.) fruit extracts. Casp J Environ Sci. 1-9. doi:10.22124/cjes.2024.8006.
- 2. Ebrahimi Y., AL-Baghdady H.F., Hameed N.M., Iswanto A.H., Shnain Ali M., Hammoodi H.A., Hashim Kzar H., Aravindhan S., Khodaei S.M., Alikord M., Pirhadi M., 2022. Common fatty acids and polyphenols in olive oil and its benefits to heart and human health. Casp J Environ Sci. 1-7. doi:10.22124/cjes.2022.5976.
- 3. Amendola C., Iannilli I., Restuccia D., Santini I., Vinci G., Akbary P., 2024. Determination of antioxidant and phytochemical properties of premix extract of brown macroalgae *Padina australis, Sargassum licifolium,* and *Stoechospermum marginatum* from Chabahar coast, Southeastern Iran. Aquat Anim Nutr. 10(1), 27-41. doi:10.22124/janb.2024.26283.1229.
- Navarro-Pascual-Ahuir M., Lerma-García M.J., Simó-Alfonso E.F., Herrero-Martínez J.M., 2016.
 Determination of water-soluble vitamins in energy and

- sport drinks by micellar electrokinetic capillary chromatography. Food Control. 63, 110-116.
- 5. Emami B., Shakerian A., Sharafati Chaleshtouri R., Rahimi E., 2024. Antioxidant, antimicrobial, and anticancer effects of the Russian olive (*Elaeagnus angustifolia* L.) fruit extracts. Casp J Environ Sci. 1-9. doi:10.22124/cjes.2024.8006.
- 6. Omotayo A.R., Oseni M.O., Oseni O.A., 2024. Medicinal benefits of aqueous extract of lemongrass (Cymbopogon citratus): effects of some biochemical evaluations, anti-microbial properties and characterization of three metallic nanoparticles. JBiochem Phytomed. 3(2), 62-71. doi: 10.34172/jbp.2024.20.
- 7. Rabiepour A., Babakhani A., Zakipour Rahimabadi E., 2024. Effect of extraction methods on the antioxidant properties of water hyacinth (*Eichhornia crassipes*). Casp J Environ Sci. 1-19. doi:10.22124/cjes.2024.8015.
- 8. Behzadi F., Roosta Y., 2025. The Role of Plant-Based Antioxidants in the Prevention and Mitigation of Hemorrhoid Complications: A Comprehensive Review in Traditional Iranian Medicine. Plant Biotechnol Persa. 7 (1), 119-124
- 9. Kim C.J., Jung Y.H., Oh H.M., 2007. Factors indicating culture status during cultivation of Spirulina (Arthrospira) platensis. J Microbiol. 45(2), 122-127.
- 10. Volkmann H., Imianovsky U., Oliveira J.L., Sant'Anna E.S., 2008. Cultivation of Arthrospira (Spirulina) platensis in desalinator wastewater and salinated synthetic medium: protein content and aminoacid profile. Braz J Microbiol. 39, 98-101.
- 11. Abu-Taweel G.M., Mohsen G.A.M., Antonisamy P., Arokiyaraj S., Kim H.J., Kim S.J., Park K.H., Kim Y.O., 2019. Spirulina consumption effectively reduces anti-inflammatory and pain related infectious diseases. J Infect Public Health. 12(6), 777-782.
- 12. Chia S.R., Chew K.W., Leong H.Y., Manickam S., Show P.L., Nguyen T.H.P., 2020. Sonoprocessing-assisted solvent extraction for the recovery of pigment-protein complex from Spirulina platensis. Chemical Eng J. 398, 125613.
- 13. Gouveia L., Batista A.P., Miranda A., Empis J., Raymundo A., 2007. Chlorella vulgaris biomass used as colouring source in traditional butter cookies. Innov Food Sci Emerg Technolog. 8(3), 433-436.

- 14. Marvizadeh M.M., Akbari N., 2019. Development and Utilization of Rice Bran in Hamburger as a Fat Replacer. J Chem Health Risks. 9(3), 245-251.
- 15. Papadaki S., Kyriakopoulou K., Tzovenis I., Krokida M., 2017. Environmental impact of phycocyanin recovery from Spirulina platensis cyanobacterium. Innov Food Sci Emerg Technol. 44, 217-223.
- 16. Selahvarzi A., Ramezan Y., Sanjabi M.R., Mirsaeedghazi H., Azarikia F., Abedinia A., 2021. Investigation of antimicrobial activity of orange and pomegranate peels extracts and their use as a natural preservative in a functional beverage. J Food Measur Charact. 15(6), 5683-5694.
- 17. Mehta A., Sharma C., Kanala M., Thakur M., Harrison R., Torrico D.D., 2021. Self-Reported Emotions and Facial Expressions on Consumer Acceptability: A Study Using Energy Drinks. Foods. 10(2), 330.
- 18. Taghavi Takyar M.B., Haghighat Khajavi S., Safari R., 2019. Evaluation of antioxidant properties of Chlorella vulgaris and Spirulina platensis and their application in order to extend the shelf life of rainbow trout (*Oncorhynchus mykiss*) fillets during refrigerated storage. LWT. 100, 244-249.
- 19. Tańska M., Konopka I., Ruszkowska M., 2017. Sensory, Physico-Chemical and Water Sorption Properties of Corn Extrudates Enriched with Spirulina. Plant Foods for Human Nutr. 72(3), 250-257.
- 20. Agregán R., Munekata P.E., Franco D., Carballo J., Barba F.J., Lorenzo J.M., 2018. Antioxidant potential of extracts obtained from macro-(Ascophyllum nodosum, Fucus vesiculosus and Bifurcaria bifurcata) and microalgae (*Chlorella vulgaris* and *Spirulina platensis*) assisted by ultrasound. Med. 5(2), 33.
- 21. Shalaby E.A., Shanab S.M., 2013. Comparison of DPPH and ABTS assays for determining antioxidant potential of water and methanol extracts of Spirulina platensis. Indian J Jeo Marine Sci. 42(5), 556-564
- 22. Rodríguez De Marco E., Steffolani M.E., Martínez C.S., León A.E., 2014. Effects of spirulina biomass on the technological and nutritional quality of bread wheat pasta. LWT Food Sci Technol. 58(1), 102-108.
- 23. Hashem Dabaghian E., Rezaei M., Tabarsa M., 2017. Ethanol extraction and solvent-solvent fractionation of algal antioxidant compounds (Enteromorpha intestinalis) Green. Iran J Nat Rec. 69(3), 385-396.

- 24. De Carvalho J.M., Maia G.A., De Figueiredo R.W., De Brito E.S., Rodrigues S., 2007. Development of a blended nonalcoholic beverage composed of coconut water and cashew apple juice containing caffeine. J Food Quality. 30 (5), 664-681.
- 25. Carr A. C., Lykkesfeldt J., 2021. Discrepancies in global vitamin C recommendations: a review of RDA criteria and underlying health perspectives. Crit Rev Food Sci Nutr. 61(5), 742-755.
- 26. Galani J. H.Y., Patel J.S., Patel N.J., Talati J.G., 2017. Storage of Fruits and Vegetables in Refrigerator Increases their Phenolic Acids but Decreases the Total Phenolics, Anthocyanins and Vitamin C with Subsequent Loss of their Antioxidant Capacity. Antioxidants. 6(3), 59.
- 27. Marvizadeh M.M., Mohammadi Nafchi A.R., Jokar M., 2016. Obtaining and Characterization of Bionanocomposite Film Based on Tapioca Starch/Bovine Gelatin/Nanorod Zinc Oxide. Int Cong Food Str Design, Turkey,160.
- 28. Marvizadeh M.M., Tajik A., Moosavian V., Oladzadabbasabadi N., Mohammadi Nafchi A., 2021. Fabrication of Cassava Starch/Mentha piperita Essential Oil Biodegradable Film with Enhanced Antibacterial Properties. J Chem Health Risks. 11(1), 23-29.
- 29. Augustin J., Johnson S.R., Teitzel C., Toma R.B., Shaw R.L., True R.H., Hogan J.M., Deutsch R.M., 1978. Vitamin composition of freshly harvested and stored potatoes. J Food Sci. 43(5), 1566-1570.
- 30. Golmakani M.T., Rezaei K., Mazidi S., Razavi S.H., 2012. Effect of alternative C2 carbon sources on the growth, lipid, and γ -linolenic acid production of spirulina (*Arthrospira platensis*). Food Sci Biotechnol. 21(2), 355-363.
- 31. Figueira F.d.S., Crizel T.d. M., Silva C.R., Salas-Mellado M.d.l.M., 2011. Pão sem glúten enriquecido com a microalga *Spirulina platensis*. Braz J Food Technol. 14, 308-316.
- 32. Sharoba A.M., 2014. Nutritional value of spirulina and its use in the preparation of some complementary baby food formulas. J Food Dairy Sci. 5(8), 517-538.
- 33. Čugura T., Pleština M., Bursać Kovačević D., Vahčić N., Dragović-Uzelac V., Levaj B., 2014. Influence of storage on quality and sensorial properties of sports drink with lemon juice and isomaltulose. Hrvatski časopis za

- prehrambenu tehnologiju. Biotehnologiju i Nutricionizam. 9(3-4), 110-116.
- 34. Dżugan M., Tomczyk M., Sowa P., Grabek-Lejko D., 2018. Antioxidant Activity as Biomarker of Honey Variety. Molecules. 23(8), 2069.
- 35. Marzieh Hosseini S., Shahbazizadeh S., Khosravi-Darani K., Reza Mozafari M., 2013. Spirulina paltensis: Food and function. Curr Nutr Food Sci. 9(3), 189-193.
- 36. Selahvarzi A., Sanjabi M.R., Ramezan Y., Mirsaeedghazi H., Azarikia F., Abedinia A., 2021. Evaluation of physicochemical, functional, and antimicrobial properties of a functional energy drink produced from agricultural wastes of melon seed powder and tea stalk caffeine. J Food Proc Preserv. 45(9), e15726.
- 37. Sözeri Atik D., Gürbüz B., Bölük E., Palabıyık İ., 2021. Development of vegan kefir fortified with *Spirulina platensis*. Food Biosci. 42, 101050.
- 38. Saharan V., Jood S., 2021. Effect of storage on Spirulina platensis powder supplemented breads. J Food Sci Technol. 58(3), 978-984.
- 39. Chaiklahan R., Chirasuwan N., Triratana P., Loha V., Tia S., Bunnag B., 2013. Polysaccharide extraction from Spirulina sp. and its antioxidant capacity. Int J Biolog Macromol. 58, 73-78.
- 40. Souza M.M.D., Prietto L., Ribeiro A.C., Souza T.D.D., Badiale-Furlong E., 2011. Assessment of the antifungal activity of Spirulina platensis phenolic extract against Aspergillus flavus. Ciência e Agrotecnologia. 35(6), 1050-1058.

- 41. Mahmoud S.H., Mahmoud R., Ashoush I., Attia M., 2015. Immunomodulatory and antioxidant activity of pomegranate juice incorporated with spirulina and echinacea extracts sweetened by stevioside. J Agricul Vet Sci. 8(2), 161-174.
- 42. Ishii M., Matsumoto Y., Nishida S., Sekimizu K., 2017. Decreased sugar concentration in vegetable and fruit juices by growth of functional lactic acid bacteria. Drug Discov Theraps. 11(1), 30-34
- 43. Ortega-Calvo J.J., Mazuelos C., Hermosin B., Saiz-Jimenez C., 1993. Chemical composition of Spirulina and eukaryotic algae food products marketed in Spain. J Appl Phycol. 5(4), 425-435.
- 44. Aljobair M.O., Albaridi N.A., Alkuraieef A.N., AlKehayez N.M., 2021. Physicochemical properties, nutritional value, and sensory attributes of a nectar developed using date palm pure and spirulina. Int J Food Properties. 24(1), 845-858.
- 45. Şahin O.I., 2020. Functional and sensorial properties of cookies enriched with Spirulina and Dunaliella biomass. J Food SciTechnol. 57(10), 3639-3646.