ANALYSIS OF WHEAT CROP PARAMETERS UNDER ALLELOPATHIC WITH REDUCED DOSE OF HERBICIDE APPLICATION.

Afza Tabassum^{1,2}*, Ziaur Rahman², Aymen Saeed³, Haleema Bibi³, Baseer Muhammad². Syed Azam Shah⁴

ABSTRACT

Water extracts from allelopathic plants like sorghum, sunflower and brassica having potential to control weeds effectively, especially when it is use in combination with reduced dose of herbicides. A field experiment was carried out to examine the feasibility of using sorghum water extract as a natural weedicide in rabi season crop (wheat) during 2015-2016 at Agronomy Research Farm, The University of Agriculture Peshawar, Pakistan. Sorgaab sprays with three different concentration (1:3, 1:4,1:5) in combination with Weedicide (affinity) in three different levels (1/3,1/2 and 2/3 of the recommended dose), with three application timing (emergence, tillering, 50% at emergence and 50 % at tillering), were tested and compared with hand weeding, unweeded and with weedicide sole. Results of the study showed that the herbicide sole followed by hand weeded plots were more effective among all the treatments for controlling weed dry weight and increasing of the grain yield. Sorghum extract sprayed at 1:5 resulted in decreased dry weight (127.2 g m⁻²), more leaf area tiller⁻¹ (101 cm²), more leaf area index (3.0) and higher grain yield (3727 kg ha⁻¹). Herbicide (affinity) at the rate of 1317g ha⁻¹ clearly decreased dry weight of weeds (133.3 g m⁻²) and increased leaf area tiller⁻¹ (100 cm²), leaf area index (2.9) and higher grain yield (3665 kg ha⁻¹). Sorghum extract concentration with combination of herbicide sprayed at tillering stage gave reduced dry weight (132), increased leaf area tiller⁻¹ (99 cm²), more leaf area index (2.9) and grain yield (3727 kg ha⁻¹). It is concluded that sorghum extract concentration 1:5 with 1317g ha⁻¹ of the herbicide affinity applied at tillering stage is recommended for higher yield and yield components of wheat crop.

Keywords: Wheat, Allelopathy, Affinity, Agriculture, Leaf area tiller⁻¹, Leaf area index, Grain yield.

^{1 -} Department of Agronomy, The University of Agriculture Peshawar-Pakistan.

^{2 -} Department of Agriculture, University of Swabi, Swabi-Pakistan.

^{3 -} Department of Plant Breeding and Genetics, The University of Agriculture, Peshawar-Pakistan.

^{4 -} Soil & Environmental Sciences Division. Nuclear Institute for Food and Agriculture Peshawar. *Corresponding author's email: rayan@aup.edu.pk

I. Introduction

In Pakistan, weeds inflict 20- 30% losses in different crops on the average (Anonymous, 2005). Existing weed control methods are either expensive or hazardous. Heavy use of chemical herbicides in most integrated weed management systems is a most concern since, it causes serious threats to the environment, public health and increase cost of crop production. Small farmers cannot afford the cost of weed management practices for crop production. Thus weeds growing among crop plants adversely affect yield and quality of harvest and increase production costs, resulting in high economic losses. Some species of weed plants might be a serious threat to crop plants diversity, sharing nutrients, moisture, sunlight and space (Ozturket al., 2012). Weeds compete with crops for nutrients, solar radiation, water, carbon dioxide and space. Apart from competition, many other factors like lodging and harvesting problems are created by weeds. In Pakistan, weeds are accountable for 30 percent grain yield losses in wheat which amounts to Rs. 1150 million annually (Marwat et al., 2008). Weed decline yield and quality of crops and leads to higher cost in food production (Pandya et al., 2005). Therefore, weed control is one of the most important aspects of crop production in agricultural systems. In addition to the direct losses caused by the weeds competition with crops for water, mineral nutrients and light, quality of crop may also be compromised by the occurrence of weeds and or their seeds, which habitually causes a great economic loss to the crop. Therefore, alternative strategies against weed must be developed (Rice, 1983) defined allopathy as the effect of one plant on other plants through the release of chemical compounds in the soil environment. Allelopathy is a natural, inexpensive, environmentally safe and an organic approach to control weeds and increase crop yields while conserving the ecosystem. Ahmad et al. (1991) concluded that sorghum is highly allelopathic and sorghum residue could be effectively used to manage some of the important weeds in irrigated wheat crop without affecting crop in semi-arid environment. Mature sorghum plants possess nine water soluble chemicals which are phytotoxic to certain weeds such as Phalaris minor Retz. Chenopodium album L., Rumex dentatus L., and Convolvulus arvensis L. (Cheema, 1988). Water extract of matured sorghum plants was used by Cheema and Khaliq (2000) and reported that water extract spray reduced weed biomass by 35-40% and increased wheat yield by 10-21%. Sorghum roots exudates reduced growth of various weed species at very low concentration (Roth et al., 2000). It has also been documented that production of allelochemicals in plants is influenced by environmental factors and greater quantities of allelochemicals have been found in plants grown under drought and mineral stress (Roth et al., 2000; Suthepet al., 2001). In recent times, studies on unitization of allelopathic chemicals (allelochemicals) as natural substances from plants for weed control in crop production have been widely noticed (Iqbal et al., 2010; Elahi et al., 2011; Afridi et al., 2013; Ahmed et al., 2014). In general, allelochemicals from plants are considered to be safe and beneficial.

Most allelochemicals are classified as secondary metabolites of the plant (Kruse *et al.*, 2000). It is well documented that the production of secondary metabolites is characterized by the plant's genetic and environmental conditions during its growth (Quader *et al.*, 2001). However, these stimulatory and inhibitory effects depend on the concentration of the compounds (Bhowmik and Inderjiit, 2003). Sorghum residues release sorgoleone, cyanogenic glycosides-dhurrin, and a number of breakdown products of phenolics that bring about weed suppression (Guenzi and McCalla, 1966; Nicollier *et al.*, 1983; Putnam, 1988; Weston *et al.*, 1989 Sorghum allelopathy for weed management in wheat 259 Weston, 1996). Cheema and Khaliq (2000) performed a series of experiments in semiarid region of Punjab, Pakistan, to explore the use of allelopathic properties

of sorghum for weed control in irrigated wheat. Allelochemicals of *P.hysterophorus*can be exploited as a source of natural herbicides to control other invasive species (Mulatu *et al.*, 2009). Earlier research (Khaliq *et al.*, 2002; Jabran *et al.*, 2008; Mahmood *et al.*, 2009; Razzaq *et al.*, 2010, 2012) findings have shown that the use of reduced doses of herbicides with allelopathic plant water extracts has increased the desirable characters of crops.

Keeping in view the losses due to weeds, resistance creating to herbicide application and the recognized importance of allelochemicals in weed management, a present field experiment were conducted under the agro climatic condition of Peshawar.

Material and methods

The experimental work was evaluated at Agronomy Research Farm, The University of Agriculture Peshawar, Pakistan. The experiment was laid out in a randomized complete block design (RCBD) having three replications in plot of size 3 m x 1.8 m (L x W). Each plot consist of six rows having 30 cm row to row distance. Wheat variety (Atta Habib 2010) at the seed rate of 120 Kg ha⁻¹ was sown on 21 November 2015 with the help of seed drill, and harvested on 2 June 2016 by hand sickle. Nitrogen and Phosphorous fertilizers were applied at the rate of 120-90 kg ha⁻¹, respectively, and source of N and P fertilizer were applied in the form of urea (46% N) and diammonium phosphate (46% P₂O₅ and 18% N). All phosphorous fertilizer and half of nitrogen were applied at the time of sowing, remaining nitrogen was applied at first irrigation. Irrigations were applied whenever required.

The following factors and their levels were studied:

Concentration of sorghum plant having three levels as factor A, C1=1:3, C2=1:4 and C3= 1:5 (i.e 1kg sorghum plant herbage+3, 4 and 5 liters of water). Herbicide ratio as factor B with three levels: R1=1/3, R2=1/2 and R=2/3 of the recommended dose of affinity (post emergence herbicide). Application times as factor C: AT1= Full at emergence (E), AT2=Full at tillering stage (T) and AT3 = Half at E + half at T. Different concentration (1:3), (1:4) and (1:5) i.e. (1 kg sorghum and 3, 4 and 5 liters of water) of sorghum extract were sprayed at three stages full at emergence, full tillering and half at emergence plus half at tillering. Experiments were also including control (without weed), hand weeding (HW) and herbicides application. Herbicide "Affinity" was applied at three different ratios i.e. one third (1/3) of the recommended dose (670 g ha⁻¹), half (1/2) of the recommended dose (988 g ha⁻¹) and two third (2/3) of the recommended dose (1317 g.ha⁻¹). All other agronomic practices were applied uniformly.

Preparation of sorghum water extracts concentrations

Mature sorghum was collected from Agronomy Research Farm, The University of Agriculture Peshawar, Pakistan. Then chopped into small pieces with the help of fodder cutting machine and soaked in tubs for 48 hours by maintaining (1:3,1:4,1:5) i.e. (1 kg plant herbage and 3, 4 and 5 liters of water) for preparation of different concentrations. Then sieved the mixture through muslin cloth to remove sorghum crop herbage and to obtain extract of sorghum. Data were recorded for dry weeds weight (g m⁻²) (40 and 60 days after sowing), Leaf area tiller⁻¹ (cm²), Leaf area index (LAI) and Grain yield (Kg ha⁻¹) according to the recommended methods.

Statistical analysis

The data recorded were analyzed statistically using analysis of variance techniques appropriate for randomized complete block design. Means will be compared using LSD test at 0.05 level of probability, when the F-values was significant (Jan *et al.*, 2009).

RESULTS AND DISCUSSION Weed dry weight (g m⁻²)

Weeds dry weight (g m⁻²) was significantly influenced by sorghum concentration (SC), affinity ratio (AR), application time (AT), control vs rest, hand weeding vs herbicide, herbicide sole vs ratio, and R x T interaction, whereas herbicide sole, C x R, C x T, C x R x T interaction were found non-significant (Table 1). More weeds dry weight recorded 40 DAS compared to 60 DAS. Maximum weeds dry weight (149.1g m⁻²) was obtained from sorghum concentration 1:3 (1kg sorghum and 3 liter water). Thereafter, weeds dry weight decreased with decreased of sorghum concentration. Minimum weeds dry weight (127.2g m⁻²) were recorded from 1:5 (1kg sorghum and 5 liters water). Lower affinity dose 670 g resulted in higher weeds dry weight (143.5g m⁻²). Higher affinity ratio (1317 g) resulted in lower weeds dry weight (133.3g m⁻²). Sorghum herbage applied at tillering produced lower weeds dry weight (132g m⁻²), while the higher weeds dry weight $(143,142 \text{g m}^{-2})$ were obtained from sorghum herbage applied at emergence or 50 at emergence + 50 at tillering stage. R x T interaction indicated that minimum weeds dry weight were recorded from emergence time x 670 g affinity, while maximum weeds weight were observed at tillering and 1317g herbicide applied. Control vs rest contrast indicated that reduced weeds dry weight (134.28g m⁻²) were obtained from rest (treated plot), while control plot resulted in higher weeds dry weight (164.80g m⁻²). Hand weeded plots produced minimum weeds dry weight (111.77g m⁻ ²) compared with herbicide treated plots (135.01g m⁻²). In herbicide vs ratio contrast the lowest weeds dry weight was obtained from ratios (138.92g m⁻²), while herbicide treated plots gave maximum weeds dry weight (144.75g m⁻²). These results are supported by Khan et al. (2015), Arif et al. (2015), Awan et al. (2012), Shahid et al. (2007), Hussain et al. (2014) who recorded significant decreased in weed dry weight with water extract obtained from different allelopathic crops.

Leaf area tiller⁻¹ (cm²)

Data analyzed statistically showed that leaf area tiller⁻¹ (cm²) was significantly influenced by sorghum concentration (SC), affinity ratio (R), application time (AT), control vs rest, hand weeding vs herbicide, herbicide sole vs ratio, and C x R whereas, C x T, R x T, C x R x T interaction were found non-significant (Table 2). Minimum leaf area tiller⁻¹ (93 cm²) was obtained from sorghum concentration 1:3 (1kg sorghum and 3 liter water). Maximum leaf area tiller⁻¹ (101 cm²) were recorded from 1:5 (1kg sorghum and 5 liters water) which was statistically similar with sorghum concentration 1:4 (1kg sorghum and 4 liters water).). Lower affinity dose (670 g) resulted in lower leaf area tiller⁻¹ (95 cm²). Leaf area tiller⁻¹ increased with each increments of herbicide and maximum leaf area tiller⁻¹ (100 cm²) was recorded from affinity applied at 1317g which was statistically at par with herbicide applied at the rate of 988g. Sorghum herbage applied either at tillering or at emergence stage produced higher leaf area tiller⁻¹ (99 cm²) than sorghum applied 50 at emergence + 50 at tillering stage (96 cm²). R x T interaction revealed that minimum leaf area tiller⁻¹ (93.4 cm²) was recorded from sorghum concentration (1:3) x 670g affinity, while maximum leaf area tiller⁻¹ (101.5cm²) was noted at 1:5 sorghum concentration and 1317 g herbicide applied. The control vs rest comparison indicated that higher leaf area tiller⁻¹ (97.76cm²) were obtained from treated plots while control plots (no treatments) give less leaf area tiller⁻¹ (89.35 cm²). Hand weeded plots produced higher leaf area tiller⁻¹ (112.9 cm²) compared with herbicide treated plots (97.3 cm²). Herbicide vs ratio contrast that maximum leaf area tiller⁻¹(97.87 cm²) was obtained from ratio while herbicide treated plots gave less leaf area tiller⁻¹ (91.68 cm²). Our results are in line with Anwar et al., (2003) who reported highest leaf area with herbicides application followed

by hand weeding. Combination of grassy and broad leaf herbicides was superior to their separate application for weed control in wheat as reported by Cheema and Akhtar (2005). Similar results were reported by Cheema *et al.*, (2000), Anwar *et al.*, (2003) who observed that concentrated sorghum water extract applied at various timing significantly increased leaf area.

Leaf area index (LAI)

Leaf area index was significantly influenced by sorghum concentration, affinity ratio, application timing, control vs. rest, hand weeding vs. herbicide, herbicide sole vs. ratio, C x R interaction, whereas, herbicide sole, C x T, R x T, C x R x T interaction were found non-significant (Table 3). Minimum leaf area index (2.5) was obtained from sorghum concentration 1:3 (1kg sorghum and 3 liter water), leaf area increased with decrease of sorghum concentration. Maximum leaf area index (3.0) was recorded from 1:5 (1kg sorghum and 5 liter water). Lower affinity dose (670 g) resulted in lower leaf area index (2.6). Higher affinity ratio (1317 g) resulted in maximum leaf area index (2.9) which was statistically at par with herbicide ratio (988g). Sorghum herbage applied at tillering stage produced higher leaf area index (2.9) which was statistically similar when applied at emergence (2.8) than sorghum applied 50 at emergence + 50 at tillering stage (2.6). C x R interaction revealed that minimum leaf area index was recorded from sorghum concentration (1:3) x 670 g affinity, while maximum leaf area index (3.0) was noted at 1:5 sorghum concentration and 1317 g herbicide applied. Control vs rest contrast indicated that maximum leaf area index (2.7) were obtained from rest (treated plot), while control plot (no treatment) resulted in minimum leaf area index (2.1). Hand weeded plots produced higher leaf area index (3.5) compared with herbicide treated plots (2.7). In herbicide vs ratio contrast the maximum leaf area index (2.7) was obtained from ratios, while herbicide treated plots gives lowest leaf area index (2.5). Similar results were reported by Mubeen et al. Jamil et al. (2009) who reported higher LAI with herbicides application followed by hand hoeing.

Grain yield (kg ha⁻¹)

Statistical analysis of the data showed that grain yield (kg ha⁻¹) was significantly influenced by sorghum concentration (SC), affinity ratio (AR), application time (AT), control vs rest, hand weeding vs herbicide and R x T interaction (Table 4) whereas, herbicide sole vs ratio, herbicide sole, C x R, C x T, C x R x T interaction were found non-significant. Minimum grain yield (3384 kg ha⁻¹) was obtained from sorghum concentration 1:3 (1kg sorghum and 3 liter water) which was statistically at par with sorghum concentration 1:4 (1kg sorghum and 4 liter water). Maximum grain yield (3727 kg ha⁻¹) were recorded from 1:5 (1kg sorghum and 5 liters water). Lower affinity dose (670 g) resulted in lower grain yield (3379 kgha⁻¹). Higher affinity ratio (1317 g) resulted in higher grain yield (3665 kg ha⁻¹). Sorghum herbage applied at tillering stage produced higher grain yield (3616 kg ha⁻¹), while the lowest grain yield (3420 kgha⁻¹) were obtained from sorghum herbage applied at emergence. R x T interaction indicated that minimum grain yield (3420.1kg ha⁻ ¹) was recorded from emergence time x 670g affinity, while maximum grain yield (3616.3kg ha⁻¹) was noted at tillering and 1317g herbicide applied. The herbicide vs. hand weeding contrast revealed that hand weeding resulted in higher grain yield (4016.0 kg ha⁻¹) compared to herbicide treated plots (3520.4 kg ha⁻¹). The control vs. rest contrast indicated that treated plots resulted in higher grain yield (3536.35 kg ha⁻¹) over control plots (2917.33 kg ha⁻¹). These findings are in line with Ahmad et al. (1993), Singh and Singh (1996) and Subhan et al. (2003) who concluded that herbicide application and hand weeding increased grain yield of wheat compared to weedy check. Similarly the results are also in conformity with the findings of Awan et al., (1990), Hassan et al. (2003) and Tunio et al. (2004) who reiterated the efficacy of herbicide applications having

been influential in raising the grain yield of wheat. It has been noted that grassy weeds like *A. fatua* and broadleaf weeds like *R. dentatus* are harmful for all the winter crops including wheat. Due to strong competitive ability of these weeds with wheat crops, the management of these weeds need to be addressed to avoid the grain yield losses (Tauseef *et al.*, 2013).

Treatments	Samplin (days af	Mean		
Concentration of sorghum (kg) L ⁻¹ water	40 days	60 days		
1:3		193.7	104.6	149.1a
1:4		188.4	92.4	140.4 b
1:5		173.2	81.3	127.2 c
Affinity (1976 g ha ⁻¹) [†]				
670	189.1	97.9	143.5 a	
988	187.3	92.6	140.0 a	
1317	178.8	87.7	133.3 b	
Application time				
Emergence (E)	189	97	143 a	
Tillering (T)	175	89	132 b	
1/2at E +1/2 at T		192	92	142 a
Planned Mean Comparison				
Control	213.25	116.36	164.80 a	
Rest of treatment	178.44	90.12	134.28 b	
Hand weeding		155.96	67.59	111.77 b
Herbicide	179.16	90.85	135.01 a	
Herbicide sole		185.58	103.92	144.75 a
Ratio		185.08	92.76	138.92 b
Affinity recommended dose	(Y	•	•
LSD _{0.05} for concentration	=	4.69		
LSD _{0.05} for application time	=	4.69		
LSD _{0.05} for ratio	=	4.69		

Table 1. Weed dry weight (g m⁻²) in wheat as affected by allelopathic and chemical with different application time.

Means within the same category followed by different letters are significantly different at P \leq 0.05 using LSD test.

Affinity	Concentration	Application Time					
$(1976 \text{ g ha}^{-1})^{\dagger} \qquad \text{of sorghum} \\ (\text{kg}) \text{ L}^{-1} \text{ water}$		Emerger (E)	nce	Tillering (T) 1/2 at		E +1/2 at T	Mean
670	1:3	95		98		90	94
	1:4	95		93		98	95
	1:5	98		96		94	96
988	1:3	95		98		89	94
	1:4	106		103		97	102
	1:5	100		98		101	100
1317	1:3	95		88		91	91
	1:4	100		101		97	99
	1:5	110		113		104	109
670		96		96		94	95 b
988		100		100		96	98 a
1317		102		101		97	100 a
	1:3	95		95		90	93 b
	1:4	100		99		97	99 a
	1:5	103		102		100	101 a
Mean		99 a		99 a		96 b	
Control							89.35 b
Rest of							97 76 a
Hand weeding							112 9a
Herbicide							97.3 h
Herbicide sole							91.68 h
Ratio							97 87 a
[†] Affinity recommend	ded dose)/.0/ u
LSD _{0.05} for concer	ntration	= 2.86					
LSD _{0.05} for applica	ation time	= 2.86					
LSD _{0.05} for ratio		= 2.86					

Table 2. Leaf area tiller⁻¹(cm²) in wheat as affected by allelopathic and chemical with different application timing.

Means within the same category followed by different letters are significantly different at $P \le 0.05$ using LSD test.

	Concentration				
Affinity (1976 g ha ⁻¹) [†]	of sorghum (kg) L ⁻¹ water	Emergence (E)	Tillering (T)	1/2at E +1/2 at T	Mean
670	1:3	2.3	2.7	2.3	2.4
	1:4	2.5	2.6	2.7	2.6
	1:5	2.8	2.8	2.5	2.7
988	1:3	2.8	2.8	2.2	2.6
	1:4	3.5	3.0	2.5	3.0
	1:5	2.9	2.8	2.8	2.8
1317	1:3	2.6	2.5	2.7	2.6
	1:4	2.9	2.9	2.6	2.8
	1:5	3.5	3.5	3.0	3.3
670		2.5	2.7	2.5	2.6 b
988		3.1	2.9	2.5	2.8 a
1317		3.0	3.0	2.8	2.9 a
	1:3	2.6	2.7	2.4	2.5 c
_	1:4	2.9	2.8	2.6	2.8 b
	1:5	3.0	3.1	2.8	3.0 a
Mean		2.8 a	2.9 a	2.6 b	
Control					2.10 a
Rest of treatments					2.76 b
Hand weeding					3.5 a
Herbicide					2.7 b
herbicide sole					2.52 b
Ratio					2.76 a
[†] Affinity recomme	nded dose				
LSD _{0.05} for concentration		= 0.14			
LSD _{0.05} for appli	cation time	= 0.14			
LSD _{0.05} for ratio		= 0.14			

Table 3. Leaf area index (LAI) in wheat as affected by allelopathic and chemical with different application timing.

Means within the same category followed by different letters are significantly different at P \leq 0.05 using LSD test.

LSD_{0.05} for ratio

Affinity	Concentration				
(1976 g ha ⁻¹) [†]	of sorghum (kg) L ⁻¹ water	Emerger (E)	nce Tillering (T)	1/2at E +1/2 at T	Mean
670	1:3	3214	3235	3466	3305
	1:4	3442	3115	3280	3279
	1:5	3516	3662	3486	3555
988	1:3	3188	3380	3452	3340
	1:4	3207	3472	3580	3420
	1:5	3650	3998	3610	3753
1317	1:3	3504	3660	3360	3508
	1:4	3333	3905	3606	3615
	1:5	3727	4120	3773	3873
670		3391	3337	3411	3379 с
988		3348	3617	3547	3504 b
1317		3521	3895	3580	3665 a
	1:3	3302	3425	3426	3384 b
	1:4	3328	3497	3489	3438 b
	1:5	3631	3927	3623	3727 a
Mean		3420 b	3616 a	3513 ab	
Control					2917.33b
Rest of					
treatments					3536.35 a
Hand weeding					4016.0a
Herbicide					3520.4 b
herbicide sole					3556.65
Ratio					3516.33
[†] Affinity recomme	nded dose				
$LSD_{0.05}$ for conc	entration	= 114.91			
LSD _{0.05} for appli	cation time	= 114.91			

Table 4. Grain yield (kg ha⁻¹) in wheat as affected by allelopathic and chemical with different application timing.

Means within the same category followed by different letters are significantly different at $P \leq 0.05$ using LSD test.

114.91

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Refferences

- Afridi, R.A., M.A. Khan, Z. Hussain, S. Saleem, Sh. Khan, Khan. Afridi and M. Ali, 2013. Allelopathic effects of Rice straw extract on different crops and weeds. ARPN J. of Agric. and Biol. Sci., 8(5): 411-418.
- Ahmad, K., Z. Shah, I. Khan, M. Khan and M.Q. Khan. 1993. Effect of post emergence herbicides application and hand weeding on wheat and weed pressure. Pak. J. Weed Sci. Res. 6 (1-2): 40-45.
- Ahmad, S., Z.A. Cheema and A. Mehmood. 1991. Response of some rabi weeds in wheat to allelopathic effects of irrigated sorghum in a sorghum wheat cropping systems. Pak. J. Weed Sci. Res., 4:81-88.
- Ahmad, W., M. Akbar, U. Farooq, A. Alia and F. Khan, 2014. Allelopathic effects of aqueous extracts of Avena fatua on seed germination and seedling growth of Triticum aestivum (variety GW-273). IOSR J. Of Environ. Sci., Toxic. And Food Techn., 8(2): 38-42.
- Allelopathy.A Science for the future, Cadiz, Spain.Anonymous. 2005. Weed Science Society of Pakistan.Wesite:wssp.org.pk.
- Anwar, S., W.A. Shah, M. Shafi, J. Bakht and M.A. Khan. 2003. Efficiency of sorgaab (sorghum water extract) and herbicide for weed control in wheat (*Triticum aestivum L.*) crop. Pak. J. Weed Sci. Res., 9(3&4):161-170.
- Arif, M., Z.A. Cheema, A. Khaliq and A. Hassan. 2015. Organic weed management in wheat through allelopathy. Int. J. Agric.Biol.,17: 127-134.
- Awan, F.K., M. Rasheed, M. Ashraf and M.Y. Khurshid. 2012. Efficacy of brassica sorghum and sunflower aqueous extracts to control wheat weeds under rainfed conditions of Pothwar, Pak. J. Anim. Pl. Sci.,22: 715-721.
- Awan, I.U., M. Iqbal and H.K. Ahmad. 1990. Screening of different herbicides for the control of weeds in wheat crop. Gomal Univ. J. Res. 10 (2): 77-83.
- Bhowmik PC, Inderjit 2003. Challenges and opportunities in implementing allelopathy for natural weed management. Crop Prot 22: 661-671.
- Cheem, M.S. and Akhtar. 2005. Efficacy of different post emergence herbicides and their application methods in controlling weeds in wheat. Pak. J. Weed Sci. Res.11(1-2):2330.
- Cheema, ZA.1988. Weed control in wheat through sorghum allelochemicals. Ph.D. Thesis, Deptt of Agron, Univ of Agric Faisalabad, Pakistan.
- Cheema, Z.A., and A. Khaliq. 2000. Use of sorghum allelopathic properties to control weeds in irrigated wheat in a semi-arid region of Punjab. Agric. Ecosystems Environ., 79:105-112.
- Cheema, Z.A., Sadiq, H.M.I. and Khaliq, A.2000 . Efficacy of sorgaab (sorghum water extract) as a natural weed inhibitor in wheat. Int. J. Agric. Biol. 2,144–146.
- Elahi, M., Z.A. Cheema, Sh.M.A.Basra, M. Akram and Q. Ali, 2011. Use of allelopatic water extract of field crops for weed control in wheat. Int. Res. J. of Plant Sci., 2(9): 262-270.

- Guenzi, W.D. and McCalla, T.M. 1966. Phenolic acids in oats, wheat, sorghum and corn residues and their phytotoxicity. Agron. J. 58, 303–304.
- Hassan, G., B. Faiz, K. B. Marwat and M. Khan. 2003. Effects of planting method and tank mixed herbicides on controlling grassy and broadleaf weeds and their effect on wheat cv Fakhr-e-Sarhad. Pak. J. Weed Sci. Res. 9: 1-11.
- Hussain, S., F. Hassan, M. Rasheed, S. Ali and M. Ahmed. 2014. Effects of allelopathic crop water extracts and their combinations on weeds and yield of rainfed wheat. J. Food. Agric. Environ., 12: 161-167.
- Iqbal, J., F. Karim and S. Hussain, 2010. Response of wheat crop (*Triticum aestivum* L.) and its weeds to allelopathic crop water extracts in combination with reduced *Tritium aestivum* L. herbicide rates. Pak. J. Agri. Sci., 47(3): 309-316.
- Jabran, K., Z.A. Cheema, M. Farooq, S.M.A. Basra, M. Hussain and H. Rehman. 2008. Tank mixing of allelopathic crop water extracts with pendimethalin helps in the management of weeds in canola (*Brassica napus*) field. Int. J. Agric. Biol., 10: 293-296.
- Jamil M, Cheema ZA, Mushtaq MN, Farooq M &Cheema MA. 2009. Alternative control of wild oat and canary grass in wheat fields by allelopathic plant water extracts. Agron Sustain Dev29: 475- 482.
- Jan, M.T., P. Shah, P.A. Hollington, M.J. Khan and Q. Sohail. 2009. Agric. Res. Des. Anal. A Monograph. NWFP Agric. Univ. Pesh. Pak.
- Khaliq, A., Z. Aslam and Z.A. Cheema. 2002. Efficacy of different weeds management strategies in mungbean (*Vigna radiata L.*). Int. J. Agric. Biol., 4: 237-239.
- Khan, E. A., A. Z. Khakhwani, M. Munir and G, Ullah. 2015. Effect of allelopathic chemicals extracted from various plant leaves on weeds control and wheat crop productivity. Pak. J. Bot. 47 (2): 735-740.
- Kruse M, Strandberg M, Strandberg B. 2000. Ecological effects of allelopathic Plants a Review. Department of Terrestrial Ecology, Silkeborg. pp 66.
- Mahmood, K., M.B. Khan, M. Hussain and M.A. Gorchani. 2009. Weed Management in Wheat Field (*Triticum aestivum L*) using allelopathic crop water extracts. Int. J. Agric. Biol., 11-6: 751-755.
- Marwat, K.B., S. Muhammad, H. Zahid, B. Gul, H. Rasheed. 2008. Study of various weed management for weed control in wheat under irrigated conditions. Pak. J. Weed sci. Res.14 (1-2):1-8.
- Mulatu, W., B. Gezahegn and T. Solomon. 2009. Allelopathic effects of an invasive alien weed *Parthenium hysterophorus L.* compost on lettuce germination and growth. African J. Agric. Res., 4(11): 1325-1330.
- Nicollier, J.F., Pope, D.F. and Thompson, A.C. 1983. Biological activity of dhurrin and other compounds from johnsongrass (*Sorghum halepense*). J. Agric. Food Chem. 31, 744–748.

- Ozturk, M., U. Kebapci, Gucel, E. Cetin and E. Altundag. 2012. Biodiversity and land degradation in the lower Euphrates sub region of Turkey. J. Environ. Biol. 33: 311-323.
- Pandya, N., G.S. Chouhan and V. Nepalia. 2005. Effect of varieties, crop geometries and weed management on nutrient uptake by soybean (*Glycine max*) and associated weeds. Indian J. Agron. 50 (3): 218-220.
- Putnam, A.R. (1988) Allelochemicals from plants as herbicides. Weed Technol. 2, 510–518.
- Quader M, Daggard G, Barrow R, Walker S, Sutherland MW. 2001. Allelopathy, DIMBOA production and genetic variability in accessions of *Triticumspletoides*. J ChemEcol 27: 742–760.
- Razzaq, A., Z.A. Cheema, K. Jabran, M. Farooq, A. Khaliq, G. Haider and S.M.A. Basra. 2010. Weed management in wheat through combination of allelopathic water extract with reduced doses of herbicides. Pak. J. Weed Sci. Res., 16 (3): 247-256.
- Razzaq, A., Z.A. Cheema, K. Jabran, M. Hussain, M. Farooq and M. Zafar. 2012. Reduced herbicide doses used together with allelopathic sorghum and sunflower water extracts for weed control in wheat. J. Plant Prot. Res., 52: 281-285.
- Rice, E. L. 1983. Allelopathy. 2nd edn., Aacademic press New York. Pp. 368.
- Roth, C.M. J.P. Shroyer and G.M. Paulsenice. 2000. Allelopathy of sorghum on wheat under several tillage systems. Agron. J., 92:855-860.
- Shahid, M., B. Ahmad, R. A. Khattak and M. Arif. 2007. Integration of herbicides with aqueous allelopathic extracts for weeds control in wheat. African Crop Sc. Conf. Proc. 8: 209-212.
- Singh, G. and O.P. Singh. 1996. Response of late sown wheat seeding methods and weed control measures in flood prone areas. Indian J. Agron. 41 (2): 237-242.
- Subhan, F., M. Khan and G.H.Jamro. 2003. Weed management through planting date, seeding rate and weed control methods in wheat.Pak. J. Weed Sci. Res. 9 (1-2): 49-57.
- Suthep, T., K. Kobayashi and K. Usui. 2001. Allelopathic activity of Mexican sunflower *Tithoniadiversifolia*(Hems) A. Gray in soil under natural field conditions and different moisture conditions. Weed Biol. Manag., 1: 115 -120.
- Tauseef, A., S. Khalid, Y. Arafat, S. Sadia, and S. Riaz. 2013. Allelopathic suppression of Avena fatuaand Rumexdentatusin associated crops. Pak. J. Weed Sci. Res., 19 (1): 31-43.
- Tunio, S.D., S.N. Kaka, A.D. Jarwar and M.R. Wagan. 2004. Effect of integrated weed management practices on wheat yield. Pak. J. Agric. Engg. Vet. Sci.20(1): 5-10.
- Weston, L.A. (1996) Distinguishing resource competition and chemical interference overcoming the methodological impasse. Agron. J. 88, 866–875.
- Weston, L.A., Harmon, R., and Mueller, S. 1989. Allelopathic potential of sorghum sudan grass hybrid (Sudex). J. Chem. Ecol. 15, 1855–1865.