Journal of Chemical Health Risks

sanad.iau.ir/journal/jchr



ORIGINAL ARTICLE

Antimicrobial and Hydrophilic Behavior of Soluble Soy Polysaccharide Starch/Cold Water Fish Gelatin Films Incorporated with Nano-Titanium Dioxide

Neda Fallah¹, Narmin Nabeghvatan², Tahereh Sadeghi^{*3}, Anna Etemadi Razlighi^{*4}, Mohammad Mehdi Marvizadeh^{*5}, Abdorreza Mohammadi Nafchi⁶

¹Department of Food Science and Technology, Qom Branch, Islamic Azad University, Qom, Iran

²Department of Food Science and Technology, Faculty of Agriculture, University of Tabriz, Tabriz P.O. Box 51666-16471, Iran

³Department of Food Science and Technology, Science and Research Branch, Islamic Azad University, Tehran, Iran

⁴Departmant of Food Science and Technology, Quchan Branch, Islamic Azad University, Quchan, Iran

⁵Young Researchers and Elite Club, Damghan Branch, Islamic Azad University, Damghan, Semnan, Iran

⁶Food Technology Division, School of Industrial Technology, Universiti Sains Malaysia, Penang, Malaysia

(Received: 18 February 2023 Accepted: 3 May 2023)

	ABSTRACT: Edible films based on starch are applied for food protection to extend shelf-life of food products. The
KEYWORDS	biodegradability of film compounds was significantly improved by the utilization of biopolymer molecules in
	comparison to petroleum packaging, and it could be a replacement for synthetic packaging. In the presentation work,
Bionanocomposite film;	the effects of nano titanium dioxide (TiO2-N) were studied on the moisture content, water solubility, moisture uptake,
Edible film;	and antimicrobial properties of soluble soy polysaccharide (SSPS) starch/cold water fish gelatin films. The nano
Physicochemical	titanium dioxide was incorporated into the SSPS starch/cold water fish gelatin dispersion at various levels (1, 2, 3, and
properties;	5 w/w% based on SSPS starch). The moisture content of the biofilms decreased from 14.12% to 12.28% and water
Solubility	solubility decreased from 93.18% to 71.46% by the incorporation of 5% TiO_2 -N into the biopolymer matrix.
	Increasing TiO_2 -N from 0 to 5% decreased the moisture uptake, a value of 9.83–6.81 g/g dried film. According to an
	increased inhibition zone of 1-5% TiO ₂ -N for Staphylococcus aureus and Escherichia coli from 32.44 to 62.27 mm ²
	and from 22.15 to 45.17 mm ² respectively, SSPS starch/cold water fish gelatin/TiO ₂ -N films showed antimicrobial
	characterizations against Staphylococcus aureus and Escherichia coli. In summary, TiO2-N improves the functional
	properties of SSPS/cold water fish gelatin biofilms and can be applied in the food industry.

INTRODUCTION

For the past decade, the using of packaging based on synthesized material have been provided a major production of residues, which makes the opinion of applying biodegradable materials in film production from biodegradable resources[1, 2]. As renewable and biodegradable materials natural polymers have advantages compared to synthetic polymers. Food quality is maintained during the production and

distribution cycles by biodegradable films that are inhibitory to volatile substances and oils.[3, 4]. There have been found some often occurring problems, however, that prevent their industrial application. First, because of their innately reduced stiffness and strength, in some cases, their mechanical qualities are quite inferior compared to synthetic polymers. Second, many materials, especially in wet conditions, are extremely

^{*}Corresponding author: sadeghibahar2030@gmail.com; anna.etemadi@yahoo.com; engmmarvi@gmail.com (T. Sadeghi; A. Etemadi Razlighi; M.M. Marvizadeh)

DOI: 10.22034/jchr.2023.1980544.1686

sensitive to water, with certain materials dissolving quickly or significantly losing mechanical strength when they absorb water. Third, their current low manufacturing level and high cost prevent them from being used in a broad range of applications.[5].

Recently, soluble soybean polysaccharide (SSPS), a new polysaccharide, was isolated from the cell wall component of soybean cotyledons. A homogalacturonan (α -1,4-galacturonan) and rhamnogalac-turonan (repeating units consisting of α -1,4-galacturonic acid and α -1,2-rhamunose) polysaccharide, branched by β -1,4galactan and α -1,5 or α -1,3- arabinan chains, make up the SSPS-like structure of pectin[6]. The film's properties of SSPS have been measured in previous study [7]and findings indicate that SSPS starch can fabricate biofilms with excellent appearance characterization and good mechanical behavior. However, SSPS starch is sensitive to water vapor and inherently soluble in water [6].

Cephalopods and fish skins, an important by-product of the fisheries industry, can be used to extract fish gelatin. It could also be used as a natural polymer in film preparation because of it's capacity to form film[8]. Fish gelatin films have been the subject of numerous research recently in an effort to distinguish them from synthetic films, which are the main source of environmental contamination and can be applied to preserve food from oxidation and drying [9]. The gelatin biopolymer forms a three-dimensional structure with intermolecular microcrystalline connections, and the low water content of this network may provide brittle biofilms. Therefore, to prepare edible film using sorbitol or glyserol as a plasticizer, its plasticity should be increased, which may also the reduction of vitreous transition[10]. However, different challenges, like sensitivity to moisture and limited antioxidant activity, restrict the usage of fish gelatin films. In terms of water sensitivity, fish gelatin film, partly dissolves and decomposes when used in contact with extremely moisture food [11].

In the recent decade, nanotechnology has helped to solve these problems [12]. Molecules and atoms the physicochemical and biological characterizations of structures at the nanoscale differ significantly from their microscale systems, providing special functional applications [13]. Nanocomposite materials consisting of metal nanoparticles (gold, silver, copper, iron, etc.) have attracted much attention because of their distinct optical, electrical, magnetic, antibacterial, and catalytic properties [14]. Natural biopolymer bionanocompositesbased packaging materials have great potential for enhancing food quality, safety, and stability as an innovative packaging and processing technology [15]. These fabrications of biocomposites containing nanoadditive have more favorable film properties, like excellence in mechanical and hydrophobic behavior. Decreasing the water vapor permeability, improving efficiency in the application of biofilm as an antimicrobial biopackaging. [16]. Acyl lipid constituents, such as linolenic, linoleic, and, oleic acids, have one or more allyl groups within the fatty acid molecule and are readily oxidized to hydroperoxides [17]. Incorporating a nanoadditive into biofilms decreased the penetration to gas molecules and could also partially reduce hydroperoxide compounds[18].

There are some reports about the improvement of biopolymers by the incorporation of nanoparticles, such as nano-titanium dioxide[19]. In the case of nanosized titanium dioxide, it has been extensively investigated in many thermoplastics (PVC, PP, PS, PE, etc.) for its robust and general reactivity, high photostability and, biocompatibility[20]. Nano titanium dioxide is used as a white pigment in paper and plastic because of its UV-resistant properties. TiO₂ can also be found in food itself, for example in sugar glazing in confectionery or instant beverages [21].

In this study, nano titanium dioxide was applied as a nanofiller to fabricate SSPS starch/cold water fish gelatin film. The films based on starch/protein were studied for their antimicrobial, moisture content, and water solubility behavior.

MATERIALS AND METHODS

Materials

SSPS starch was purchased from SIM Co (Penang, Malaysia), and cold water fish gelatin was purchased from Sigma Chemical Co (Louis, USA). Glycerol and sorbitol were obtained from Macklin Biochemical (Shanghai, China). Nano titanium dioxide was purchased from Nano meghyas borhan (Mashhad, Iarn).

Film preparation

Neat SSPS starch/cold water fish gelatin and active film containing Nano-TiO₂ were fabricated by casting method. Nanoadditive at levels of 1, 2, 3, and 5(w/w%, based on SSPS starch) were added to 100 mL of distilled water and aqueous solutions were stirred magnetically for 60 min and solutions were exposed to ultrasound wavelength for 45min. Neat starch/gelatin biofilm suspension was fabricated by dispersing SSPS starch (8 g), cold water fish gelatin(0.8 g) [22, 23], and glycerol/sorbitol (3.2 g) [24] in 100mL of Nano-solution. All the suspensions were heated at 85°C for 1h followed by cooling at 37°C. At last, the suspensions were poured on casting plates, and dried at 25°C (RH = 50%) to form bionaocomposite films.

Characterization of starch/gelatin film

Moisture content

The moisture content of biofilms was measured by evaluating the weight difference of biofilms before and after drying in a desiccator containing CaCl₂ and placed in an oven at 40°C for 24 h. Five replications of each biofilm were applied for calculating the moisture content.

Solubility

Pieces of each biofilm were cut and placed in a glass container with P_2O_5 for 48h and weighed to measure the dry weight of the film. The solubility of the biofilms was evaluated by immersion test in 80 ml of deionized water with periodic agitation for 1 h at 25°C. At last, the remaining pieces of biofilms were filtered and dried at 60°C to achieve constant weight. The solubility (%) of films was calculated as follow:

Solubility(%)= Initial film weight-Final film weight Initial film weight

Antimicrobial characterizations

Antimicrobial experiment on the biofilms was performed by applying the agar diffusion technique based on Maizura, et al. [25]. First 1 mL of inoculation containing 10^{5} – 10^{6} CFU/mL of gram-negative *E. coli* and grampositive *S. aureus* were seeded onto Mueller Hinton agar (MHA) solid media. Film disks were then placed on MHA media. After that, the plates were then incubated for 1 day at 37°C. Following that, the plates had a "zone of inhibition" of the film disc examination. Computing the inhibition zone of gram-negative *E. coli* and grampositive *S. aureus* on MHA media allowed researchers to determine the film's antimicrobial effects. The zone of inhibition of bactrial was determined by calculating the area of the entire zone and subtracting it from the film disk area.

Moisture uptake

Moisture uptake of active film was determined following the technique of Tajik, et al. [26]. Specimen film (2 cm \times 2 cm) was cut and dried with P₂O₅ for 2 days and then weighed (w₁). The dried film was conditioned with potassium sulfate solution (RH = 97%) for 2 days. The active film was removed from control condition and weighed (W₂).

The moisture uptake was stated as following equation:

Moisture uptake = $(w_2 - w_1)/w_1$

Statistical analysis

One-Way ANOVA was utilized to compare data of solubility, moisture content, and antimicrobial behaviors of films at 95% confidence level. Data from antimicrobial and chemical experiments were analyzed by Minitab 17.3.1 software.

RESULTS AND DISCUSSION

Moisture content

The moisture content of the nano titanium dioxide/SSPS starch/cold water fish gelatin films is presented in Table 1. The moisture content decreased as TiO₂-N concentration was enhanced. The interaction between the glycerol and sorbitol, biopolymer molecules, and nanofiller is probably to blame for the decreased availability of the hydroxyl group to interact with water, which results in a less hygroscopic matrix[27]. According to Hajizadeh, et al. [28] interactions between

the titanium and the hydroxyl groups from the glycerol/sorbitol and water, and also between the

titanium and the plasticizer of the ion, occur.

TiO ₂ -N (%)	Moisture content (%)
0	14.12±0.08a
1	13.34±0.05b
2	12.95±0.06c
3	12.56±0.1d
5	12.28+0.06e

Table 1. Moisture content of SSPS starch/cold water fish gelatin/TiO2 film.

Data are mean and SD. Different letters in column show significant difference at p<0.05 among SSPS starch/cold water fish gelatin/TiO₂ films

Solubility

Solubility in water may be an important factor in defining applications for biopolymer composite films [29]. The solubility of the nano titanium dioxide/SSPS starch/cold water fish gelatin films is presented in Table 2. The incorporation of nano titanium dioxide into SSPS starch/cold water fish gelatin matrix significantly decreased the solubility of the biocomposites. These presentation findings are in agreement with other

scholars on biocomposite film containing nanostructure [30, 31]. This result might be explained by interactions between the film-forming component of the biopolymer and TiO₂-N[32]. According to another study, more hydrogen bonds between the components of the matrix and TiO₂ are formed as the nanofiller concentration of film increases[33].

Table 2. Solubility of SSPS starch/cold water fish gelatin/TiO₂ film

TiO ₂ -N (%)	Solubility (%)
0	93.18.±2.11a
1	89.45±1.59a
2	83.85±1.63b
3	77.71±1.87c
5	71.46±1.75d

Data are mean and SD. Different letters in column show significant difference at

p<0.05 among SSPS starch/cold water fish gelatin/TiO2 films

Antimicrobial properties

Table 3 indicates the impacts of TiO_2 -N level on the antimicrobial characterization of different biofilms against gram-negative *E. coli* and gram-positive *S. aureus*. The ncorporation of nano titanium dioxide into SSPS starch/cold water fish gelatin matrix significantly increased the antimicrobial activity of the biocomposites. These findings are consistent with other scholars on nanobiocomposite [34-36]. The physical and/or chemical

contact between nanofiller and the microbial cell wall is one mechanism that relates the antimicrobial property of nanofiller to this phenomenon. The microorganism's cell wall might allow the Ti4+ to permeate and react with different components, which would ultimately influence the viability of the cells [37] and the formation of H_2O_2 due to presence of nanoadditive particles[38].

N. Fallah et al/ Journal of Chemical Health Risks 14(2) (2024) 291-298

TiON (%)	Inhibition zone(mm ²)		
1102-11 (70)	S. aureus	E. coli	
0	1.13±0.05a	1.13±0.05a	
1	32.14±0.46b	22.15±0.51b	
2	37.18±0.73c	27.69±0.79c	
3	42.23±1d	33.24±0.84d	
5	62.27±0.88e	45.17±1e	

Table 3. Antimicroial properties of SSPS starch/cold water fish gelatin/TiO2 film

Data are mean and SD. Different letters in columns show significant difference at p<0.05 among SSPS starch/cold water fish gelatin/TiO₂ films

Moisture uptake

The moisture uptake of pure SSPS/gelatin and SSPS/gelatin / TiO_2 -N nanocomposite films was between 9.83 and 6.81 g/g dried film (Table 4). The findings indicated that neat biofilms had the highest moisture uptake. The incorporation of nanoadditive significantly (p < 0.05) decreased the moisture uptake of the biodegradable films depending on the level of TiO₂-N. The higher moisture uptake present in the neat film based

on SSPS/gelatin may be due to hydroxyl groups in the biopolymer structure.

TiO₂-N may interact with the biopolymer molecules upon adding the nanoadditive into SSPS/gelatin matrix, which may reduce biopolymer-water interaction[39]. Similar findings have been stated by scholars regarding reduced moisture uptake in biofilms incorporated with TiO₂-N[23], and ZnO-nanorod [40].

Table 4. Moisture uptake of SSPS starch/cold water fish gelatin/TiO2 film

TiO ₂ -N (%)	Moisture uptake (g/gdried film)
0	9.83.±0.08a
1	8.54±0.1b
2	8.09±0.11c
3	7.65±0.13d
5	6.81±0.08e

Data are mean and SD. Different letters in column show significant difference at p<0.05 among SSPS starch/cold water fish gelatin/TiO₂ films

CONCLUSIONS

In this study, we incorporated nano-TiO₂ into the SSPS starch/cold water fish gelatin matrix to fabricate bionanocomposites. Results obtained in this study indicate that nano titanium dioxide particles could improve bionanocomposites film properties such as moisture content, moisture uptake, and solubility. Nano-TiO₂ embedded in biofilms based on starch/protein represents good antimicrobial characterization against gram-negative *E. coli* and gram-positive *S. aureus*. The findings indicated that biodegradable film based on SSPS starch/cold water fish gelatin/nano-TiO₂ may have favorable potential in the food packaging industries.

ACKNOWLEDGEMENTS

This research did not receive any specific grant from funding agencies in the public, commercial or notforprofit sectors.

Conflict of interests

The authors declare that there is no conflict of interest.

REFERENCES

1. Abedinia A., Alimohammadi F., Teymori F., Razgardani N., Saeidi Asl M.R., Ariffin F., Mohammadi Nafchi A., Huda N., Roslan J., 2021. Characterization and Cell Viability of Probiotic/Prebiotics Film Based on Duck Feet Gelatin: A Novel Poultry Gelatin as a Suitable Matrix for Probiotics. Foods. 10(8), 1761. 2. Abedinia A., Ariffin F., Huda N., Mohammadi Nafchi A., 2018. Preparation and characterization of a novel biocomposite based on duck feet gelatin as alternative to bovine gelatin. International Journal of Biological Macromolecules. 109, 855-862.

3. Fallah N., Marvizadeh M.M., Jahangiri R., Zeinalzadeh A., Mohammadi Nafchi A., 2022. High-Barrier and Light–protective Bionanocomposite Film Based on Rye Starch/nanorod-ZnO for Food Packaging Applications. Journal of Chemical Health Risks. [In press].

4. Marvizadeh M.M., Tajik A., Moosavian V., Oladzadabbasabadi N., Mohammadi Nafchi A., 2021. Fabrication of Cassava Starch/Mentha piperita Essential Oil Biodegradable Film with Enhanced Antibacterial Properties. Journal of Chemical Health Risks. 11(1), 23-29.

5. Joseph B., Krishnan S., Sagarika V.K., Tharayil A., Kalarikkal N., Thomas S., 2020. Bionanocomposites as industrial materials, current and future perspectives: a review. Emergent Materials. 3(5), 711-725.

6. Akbariazam M., Ahmadi M., Javadian N., Mohammadi Nafchi A., 2016. Fabrication and characterization of soluble soybean polysaccharide and nanorod-rich ZnO bionanocomposite. International Journal of Biological Macromolecules. 89, 369-375.

7. Salarbashi D., Tajik S., Ghasemlou M., Shojaee-Aliabadi S., Shahidi Noghabi M., Khaksar R., 2013. Characterization of soluble soybean polysaccharide film incorporated essential oil intended for food packaging. Carbohydrate Polymers. 98(1), 1127-1136.

 Kchaou H., Benbettaïeb N., Jridi M., Abdelhedi O., Karbowiak T., Brachais C.-H., Léonard M.-L., Debeaufort F., Nasri M., 2018. Enhancement of structural, functional and antioxidant properties of fish gelatin films using Maillard reactions. Food Hydrocolloids. 83(3), 326-339.

9. Etxabide A., Uranga J., Guerrero P., de la Caba K., 2017. Development of active gelatin films by means of valorisation of food processing waste: A review. Food Hydrocolloids. 68(4), 192-198.

10. Ashrafi A., Babapour H., Johari S., Alimohammadi F., Teymori F., Nafchi A.M., Shahrai N.N., Huda N., Abedinia A., 2023. Application of Poultry Gelatin to Enhance the Physicochemical, Mechanical, and

Rheological Properties of Fish Gelatin as Alternative Mammalian Gelatin Films for Food Packaging. Foods. 12(3), 670.

11. Hanani Z.A.N., Yee F.C., Nor-Khaizura M.A.R., 2019. Effect of pomegranate (*Punica granatum* L.) peel powder on the antioxidant and antimicrobial properties of fish gelatin films as active packaging. Food Hydrocolloids. 89, 253-259.

 Rhim J.W., Ng P.K.W., 2007. Natural Biopolymer-Based Nanocomposite Films for Packaging Applications. Critical Reviews in Food Science and Nutrition. 47(4), 411-433.

13. He X., Deng H., Hwang H.M., 2019. The current application of nanotechnology in food and agriculture. Journal of Food and Drug Analysis. 27(1), 1-21.

 Ashaolu T.J., 2021. Emerging applications of nanotechnologies to probiotics and prebiotics. International Journal of Food Science & Technology. 56(8), 3719-3725.

15. Neethirajan S., Jayas D.S., 2011. Nanotechnology for the Food and Bioprocessing Industries. Food and Bioprocess Technology. 4(1), 39-47.

16. Hosseini S. F., Gómez-Guillén M.C., 2018. A stateof-the-art review on the elaboration of fish gelatin as bioactive packaging: Special emphasis on nanotechnology-based approaches. Trends in Food Science & Technology. 79, 125-135.

17. Nobari A., Marvizadeh M.M., Sadeghi T., Rezaeisavadkouhi N., Mohammadi Nafchi A., 2022. Flavonoid and Anthocyanin Pigments Characterization of Pistachio Nut (*Pistacia vera*) as a Function of Cultivar. Journal of Nuts. 13(4), 313-322.

 Marvizadeh M.M., Mohammadi Nafchi A., Jokar M.,
2014. Preparation and Characterization of Novel Bionanocomposite Based on Tapioca Starch / Gelatin / Nanorod-rich ZnO: Towards Finding Antimicrobial Coating for Nuts. Journal of Nuts. 5(2), 39-47.

19. Nassiri R., Mohammadi N. A., 2013. Antimicrobial and barrier properties of bovine gelatin films reinforced by nano TiO_2 . Journal of Chemical Health Risks. 3(3), 21-28

20. Hanemann T., Szabó D.V., 2010. Polymer-Nanoparticle Composites: From Synthesis to Modern Applications. Materials. 3(6), 3468-3517. 21. Noman M.T., Ashraf M.A., Ali A., 2019. Synthesis and applications of nano-TiO₂: a review. Environmental Science and Pollution Research. 26(4), 3262-3291.

22. Marvizadeh M.M., Mohammadi Nafchi A., Jokar M., 2014. mproved Physicochemical Properties of Tapioca Starch / Bovine Gelatin Biodegradable Films with Zinc Oxide Nanorod. Journal of Chemical Health Risks. 4(4), 25-31.

23. Shaili T., Abdorreza M.N., Fariborz N., 2015. Functional, thermal, and antimicrobial properties of soluble soybean polysaccharide biocomposites reinforced by nano TiO_2 . Carbohydrate Polymers. 134, 726-731.

24. Abdorreza M.N., Cheng L.H., Karim A.A., 2011. Effects of plasticizers on thermal properties and heat sealability of sago starch films. Food Hydrocolloids. 25(1), 56-60.

25. Maizura M., Fazilah A., Norziah M.H., Karim A.A., 2007. Antibacterial Activity and Mechanical Properties of Partially Hydrolyzed Sago Starch–Alginate Edible Film Containing Lemongrass Oil. Journal of Food Science. 72(6), C324-C330.

26. Tajik S., Maghsoudlou Y., Khodaiyan F., Jafari S.M., Ghasemlou M., Aalami M., 2013. Soluble soybean polysaccharide: A new carbohydrate to make a biodegradable film for sustainable green packaging. Carbohydrate Polymers. 97(2), 817-824.

27. Marvizadeh M.M., Oladzadabbasabadi N., Mohammadi Nafchi A., Jokar M., 2017. Preparation and characterization of bionanocomposite film based on tapioca starch/bovine gelatin/nanorod zinc oxide. International Journal of Biological Macromolecules. 99, 1-7.

28. Hajizadeh H., Peighambardoust S.J., Peighambardoust S.H., Peressini D., 2020. Physical, mechanical, and antibacterial characteristics of bionanocomposite films loaded with Ag-modified SiO₂ and TiO₂ nanoparticles. Journal of Food Science. 85(4), 1193-1202.

29. Moosavian V., Marvizadeh M.M., Mohammadi Nafchi A., 2017. Biodegradable Films Based on Cassava Starch/Mentha piperita Essence: Fabrication, Characterization and Properties. Journal of Chemical Health Risks. 7(3), 239-245.

30. Arezoo E., Mohammadreza E., Maryam M., Abdorreza M.N., 2020. The synergistic effects of

cinnamon essential oil and nano TiO2 on antimicrobial and functional properties of sago starch films. International Journal of Biological Macromolecules. 157, 743-751.

31. Malathi A., Singh A., 2019. Antimicrobial activity of rice starch based film reinforced with titanium dioxide (TiO₂) nanoparticles. Agric Res J. 56, 111.

32. Marvizadeh M.M., Mohammadi Nafchi A.R., Jokar M., 2016. Obtaining and Characterization of Bionanocomposite Film Based on Tapioca Starch/Bovine Gelatin/Nanorod Zinc Oxide. Conference: Food Structure Design: Antalya, turkey.

33. Fathi N., Almasi H., Pirouzifard M. K., 2019. Sesame protein isolate based bionanocomposite films incorporated with TiO_2 nanoparticles: Study on morphological, physical and photocatalytic properties. Polymer Testing. 77, 105919.

34. Xing Y., Li X., Guo X., Li W., Chen J., Liu Q., Xu Q., Wang Q., Yang H., Shui Y., Bi X., 2020. Effects of Different TiO_2 Nanoparticles Concentrations on the Physical and Antibacterial Activities of Chitosan-Based Coating Film. Nanomaterials. 10(7), 1365.

35. Zhang X., Xiao G., Wang Y., Zhao Y., Su H., Tan T., 2017. Preparation of chitosan-TiO₂ composite film with efficient antimicrobial activities under visible light for food packaging applications. Carbohydrate Polymers. 169, 101-107.

36. Azarashkan Z., Farahani S., Abedinia A., Akbarmivehie M., Motamedzadegan A., Heidarbeigi J., Hayaloğlu A.A., 2022. Co-encapsulation of broccoli sprout extract nanoliposomes into basil seed gum: effects on in vitro antioxidant, antibacterial and anti-Listeria activities in ricotta cheese. International Journal of Food Microbiology. 376, 109761.

37. Abutalib M.M., Rajeh A., 2021. Enhanced structural, electrical, mechanical properties and antibacterial activity of Cs/PEO doped mixed nanoparticles (Ag/TiO₂) for food packaging applications. Polymer Testing. 93, 107013.

38. Marvizadeh M.M., Mohammadi Nafchi A., Jokar M., 2014. Preparation and Characterization of Novel Bionanocomposite Based on Tapioca Starch/Gelatin/Nanorod-rich ZnO: Towards Finding Antimicrobial Coating for Nuts. Journal of Nuts. 5(2), 39-47. 39. Yun Y.H., Youn Y.N., Yoon S.D., Lee J.U., 2012. Preparation and physical properties of starch-based nanocomposite films with the addition of titanium oxide nanoparticles. Journal of Ceramic Processing Research. 13(1), 59-64. 40. Chavoshi N., Marvizadeh M.M., Fallah N., Rezaeisavadkouhi N., Mohammadi Nafchi A., 2023. Application of Novel Nano-biopackaging Based on Cassava Starch/Bovine Gelatin / Titanium oxide nanoparticle/Fennel Essential Oil to Improve Quality of the Raw Fresh Pistachio. Journal of Nuts. 14(1), 19-31.