

Troubleshooting With You



One-Step Reverse Transcription PCR (RT-PCR)

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- **What is RT-PCR?**

Reverse transcription polymerase chain reaction (RT-PCR) is a variant of polymerase chain reaction (PCR). It is a sensitive method for the detection of mRNA expression levels. Traditionally RT-PCR involves two steps: the RT reaction and PCR amplification. RNA is first reverse transcribed into cDNA using a reverse transcriptase; the resulting cDNA is used as templates for subsequent PCR amplification using primers specific for one or more genes. RT-PCR can also be carried out as one-step RT-PCR in which all reaction components are mixed in one tube prior to starting the reactions. Although one-step RT-PCR offers simplicity and convenience and minimizes the possibility for contamination, the resulting cDNA cannot be repeatedly used as in two-step RT-PCR, sometimes abbreviated as RT-PCR.

- **How does it work?**

One-step reverse transcription PCR is carried out using the Access RT-PCR System, (Promega, USA), to detect desired mRNA, using the specific designed primer sets. The total reaction mixture volume (25 μ L) contained 5 μ L (1 μ g) of total RNA, 1.5 mM MgSo₄ (25 mM), 1 μ M of each forward and reverse primer solutions (10 mM), 5 μ L (1X) of AMV/*Tfl* 5X Reaction buffer, 0.2 mM dNTP mix (10 mM), 0.1 U/ μ l of 5 U/ μ l AMV Reverse transcriptase, and 0.1 U/ μ l of 5 U/ μ l *Tfl* DNA polymerase, topped up with DEPC treated water.

- **How does it run?**

The thermal cycling profile is as follows: reverse transcription (45 °C for 45 minutes), inactivation of AMV reverse transcriptase (94 °C for 2 minutes), denaturation of RNA/cDNA primer, (94 °C for 2 minutes), denaturation (94 °C for 30 seconds), annealing (depends on the forward and reverse primers for 1 minute), extension (68 °C for 2 minute), final extension (68 °C for 7 minute), hold (4 °C) with the denaturation, annealing and extension steps repeated for 35 cycles. The PCR products were separated on a 1.0% agarose gel run at 120 V for 25 minutes. The PCR products will be analyzed under UV light after ethidium bromide staining (0.5 μ g/ml).

- **When RT-PCR fails, what is the Solution?**

Low yield or no amplification products may be a result of insufficient number of cycles, incorrectly programmed thermal cycler, improper reaction condition. Solution: reduce the annealing temperature or allow longer extension times.

Multiple or non-specific amplified products may be due to poor primer design, contamination by another target RNA/DNA, existence of multiple target sequence in the target RNA. Solution: Designing new sets of primers might help.

- **Contact us**

For more information or troubleshooting on your Transformation, please do not hesitate to contact us at ijpp@iau-saveh.ac.ir. You can simply mention your problem by attaching your results. We look forward to hearing from you soon.

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Acknowledgements

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Ouyang, D., J. Bartholic and J. Selegean, 2005. 'Assessing sediment loading from agricultural croplands in the great lakes basin'. *Journal of American Science*, 1 (2): 14-21.

Books:

Durbin, R., S. R. Eddy, A. Krogh and G. Mitchison. 1999. *Biological Sequence Analysis: Probabilistic Models of Proteins and Nucleic Acids*. Cambridge: University Press.

A chapter in a book:

Leach, J. 1993. 'Impacts of the zebra mussel (*Dreissena polymorpha*) on water quality and fish spawning reefs of Western Lake Erie'. In *Zebra Mussels: biology, impacts and control*. Nalepa, T. and D. Schloesser (Eds.). Ann Arbor, MI: Lewis Publishers, pp: 381-397.

A Report:

Makarewicz, J. C., T. Lewis and P. Bertram. 1995. *Epilimnetic phytoplankton and zooplankton biomass and species composition in Lake Michigan 1983-1992*. U.S. EPA Great Lakes National Program, Chicago, IL. EPA 905-R-95-009.

Conference proceedings:

Stock, A. 2004. 'Signal transduction in bacteria'. Proceedings of the 2004 Markey Scholars Conference, pp: 80-89.

A thesis:

Strunk, J. L. 1991. *The extraction of mercury from sediment and the geochemical partitioning of mercury in sediments from Lake Superior*. M. Sc. thesis, Michigan State Univ., East Lansing, MI.

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