

The study of Allelopathic potential in three species of *Glaucium* Mill. on *Sinapis* arvensis L.

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Abstract

Charlock (*Sinapis arvensis* L.) is a serious *weed* of agriculture, especially in spring sown crops. Genus *Glaucium* Mill. belongs to Papaveraceae and members of this genus have several alkaloids in laticiferes. The present research focuses on effects of aquatic extract of *Glaucium oxylobum*, *G. elegans* and *G. grandiflorum* on many physiological factors of Charlock. Laboratory studies were conducted to determine the seed germination and growth factors in charlock as affected by water soluble constituents of three species of *Glaucium*. Aboveground tissues of these plants were collected during flowering stage and dried. Five concentrations of aqueous extracts were used: full- strength (33.3 g lit⁻¹), three-fourth - strength (25 g lit⁻¹), half- strength (16.7 g lit⁻¹) and quarter- strength (8.3 g lit⁻¹) and blank (0 g lit⁻¹). Charlock seeds were placed in petri dishes containing the *Glaucium* extracts, or distilled water (control). Seed germination percent was calculated after ten days. In general seed germination of Charlock declined progressively with increasing concentration of the *Glaucium* extracts. According to our results, with increasing concentration of aqueous extract, Charlock growth factors such as root and shoot length, root and shoot fresh and dry weight were inhibited.

Keywords: alkaloid; allelopathy, Glaucium; growth factors; seed germination; Sinapis arvensis

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Introduction

Weed is a serious problem that damages most of the crops (Samad et al., 2008). Charlock (Sinapis arvensis L.) is a serious weed of agriculture, especially in spring sown crops. It is frequently found growing in colza fields (Haddadchi and Massoodi Khorasani, 2006) and it

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Received: July, 2011 Accepted: September 2011 is more of a problem in spring-sown than autumn-sown crops (Clapham, 1987).

Allelopathy as a mechanism of plant interference in agro-ecosystems offers an opportunity to manage weeds in a crop sequence, but could also adversely affect crop yields and influence choice of rotation. Chemicals with allelopathic activity are present in many plants and in organs, including leaves, flowers, fruits and buds (Ashrafi et al., 2007, 2008; Inderjit, 1996; Mahall and Callaway, 1991; Putnam and Tang, 1986). Allelopathy is in fact an interference mechanism in which live or dead

plant materials release chemical substances, which inhibit or stimulate the associated plant growth. Several researchers have shown that allelopathy plays an important part in weed and weed interaction (Jabeen and Ahmed, 2009).

Recent studies pay attention allelopathic potential of Glaucium oxylobum Boiss. & Buhse, G. elegans Crantz and G. grandiflorum Boiss. & Huet on seed germination and root, shoot length as well as root, shoot fresh and dry weight of Charlock. Members of Glaucium contain several alkaloids in laticiferes such as G.oxylobum. Four major alkaloids include protopine, bulbocapine, corydine, isocorydine and three minor alkaloids are dihydrochlerythrine, angoline, isocorytuberine (Hadjiakhoondi et al., 1999). G. elegans contain (glaucine, aporphin group isoboioidine, isocorydine and corydine), protopine group (protopin allocryptopin) and and benzophenanthridine group (chelidonin, sanguinarine, chlerythrine, chlirubine) (Yakhontova et al., 1975). G. grandiflorum also alkaloids such as norchelidonine, dihydrochelerythrine, 8acetonyldihydrochelerythrine, protopine, tetrahydrojattorrhizine allocryptopine, (coryplamine) and tetrahydropalmatine (El-afifi et al., 1986).

The purpose of this study was to determine the possible allelopathic effects of *Glaucium* on germination and growth factors *Sinapis arvensis* L.

Materials and Methods

To study the allelopathic effects of Glaucium aqueous extract on seed germination and seedling growth of Sinapis arvensis L., an experiment was carried out in Physiology Laboratory, Department of Biology, Islamic Azad University, Gorgan Branch, Iran. For breaking of dormancy in seeds, we applied several materials, namely, GA (10ppm, 300ppm, 1000ppm, 2000ppm); H_2SO_4 ; HCl and KNO₃ (0.28%). Charlock seeds were first surface-sterilized in 96% ethyl alcohol for 5 min and sterile seeds were imbibed in 10 mm of materials for 24 hr at 0° C, and afterward washed thoroughly to remove the external source of materials. The best result for

breaking of dormancy was observed in GA (1000ppm) (Haddadchi and Massoodi Khorasani, 2006). Fifty seeds were incubated in extracts.

Glaucium oxylobum, G. elegans and G. grandiflorum plants were gathered in spring 2007-2008 from several places in north of Iran, and dried under shade for a few days. All of the specimen were saved and deposited in herbarium of Islamic Azad University, North Tehran Branch (Glaucium oxylobum 4007- IAUNT, G. elegans 4000- IAUNT, and G. grandiflorum 4002- IAUNT). The well dried plants were chopped into about 3 cm pieces with fodder cutter. Chopped plant material was dried in an oven at 90°C for 24 h (Shimifan LO-141). The oven dried shoots was ground in a grinder. 10 g of dried shoots was weighed (Scale: Mettler college -1300) soaked in distilled water (300ml) at 70° C and then placed for 24h in water bath (Behdad-80004) at 60° C. The water extract was obtained by filtering the mixture (herbage and water) through filter paper and then diluted with distilled water to prepare different according to the treatment concentrations (Narwal, 1999). Petri dishes were given a thorough washing with detergent using hot water as precautionary measure against pathogens and pollutants. Glaucium water extracts were diluted with distilled water to prepare solutions of different concentrations (v/v): 25, 50, 75 and 100%. For control treatment only distilled water was used. Fifty seeds of Sinapis arvensis were put in each petri dish after removing of dormancy. Overall 3 dishes were used in the experiment (3 replications) in completely randomized design. Filter paper (Whatman) was used as medium of germination. Five ml of solution was applied to the dishes and the control treatment received 5 ml of distilled water. The outer filter paper was removed just before the initiation of germination. Both treated and control Petri dishes were kept continuously moist by applying distilled water or extracts, whenever needed. The dishes were kept at seed chamber temperature (20° C±2) for seed germination in the Physiology Laboratory (Irankhodsaz, I K H- RH). Germination counts were recorded daily for a period of 10 days. Root and shoot length was recorded using a ruler. Fresh and dry weight of shoot and root were measured using a scale (GF-600), after 10 days.

Statistical analysis

The data were analyzed using Analysis of Variance (ANOVA) test. A one-way ANOVA was performed on all results. Probabilities of less than 0.05 were considered statistically significant. Differences between means were determined using Duncan compromise test. The software SPSS (Ver. 17) was used to conduct all the statistical analysis.

Results

Extracts from shoot of Glaucium, greatly inhibited Charlock seed germination at all concentrations when compared with the water control. Germination and growth factors of Charlock was significantly (P≤0.005) reduced by dried shoot extracts of Glaucium (Tables 1, 2 and 3). The highest inhibition in seed germination (up to 98.67%) was observed in concentration (100%) while the lowest (up to 44.67%- 81.34%) was recorded in concentration (25%). The extracts not only reduced the seed germination but also shoot and root length and fresh and dry weight of shoot and root (Tables 1-3). Our results showed that allelochemicals in Glaucium have significant effect on seed germination percentage and germination index.

Discussion

Shoot aqueous extract of Glaucium had a clear reduction effect on the seedling growth characteristics in Sinapis arvensis. The results of this study suggest that dried shoot of Glaucium

Table 1 Allelopathic effect of Glaucium oxylobum water extract on the germination and seedling growth of Sinapis arvensis

| Parameter | Percentage of Germination | Root Length (mm) | Shoot Length (mm) | FW.Root (mg) | DW.Root (mg) | FW.Shoot (mg) | DW.Shoot (mg) |
|-----------|------------------------------|---------------------|----------------------|-----------------|-----------------|------------------|------------------|
| 0% | 80a | 18.33a | 21.66a | 48.37a | 2.19a | 76.57a | 5.84a |
| 25% | 36.66b | 5.66b | 16.66b | 6.64b | 0.28b | 27.72b | 2.31b |
| 50% | 26c | 1.66c | 3.66c | 1.53c | 0.06bc | 4.43c | 0.32c |
| 75% | 6.66d | 1.33c | 0.66c | 0.28cd | 0c | 0d | 0c |
| 100% | 1.33d | 0.5c | 0c | 0d | 0c | 0d | 0c |

Table 2 Allelopathic effect of Glaucium elegans water extract on the germination and seedling growth of Sinapis arvensis

| Parameter | Percentage of | Root Length | Shoot Length | FW.Root | DW.Root | FW.Shoot | DW.Shoot |
|-----------|---------------|-------------|--------------|---------|---------|----------|----------|
| | Germination | (mm) | (mm) | (mg) | (mg) | (mg) | (mg) |
| 0% | 71.33a | 28.33a | 18.33a | 38.95a | 0.31a | 32.33a | 1.39a |
| 25% | 18.66b | 1.33b | 1.33b | 0.48b | 0.003b | 0.55b | 0.029b |
| 50% | 12.66b | 1.25b | 0.66b | 0.4b | 0b | 0.19b | 0.008b |
| 75% | 11.33b | 0.91b | 0.33b | 0.19b | 0b | 0.08b | 0b |
| 100% | 1.33c | 0.5b | 0b | 0b | 0b | 0b | 0b |

Allelopathic effect of Glaucium grandiflorum water extract on the germination and seedling growth of Sinapis arvensis

| Parameter | Percentage of | Root Length | Shoot Length | FW.Root | DW.Root | FW.Shoot | DW.Shoot |
|-----------|---------------|-------------|--------------|---------|---------|----------|----------|
| | Germination | (mm) | (mm) | (mg) | (mg) | (mg) | (mg) |
| 0% | 80a | 51.66a | 45a | 52.74a | 2.65a | 80.57a | 10.29a |
| 25% | 55.33b | 5.66b | 11.66b | 4.39b | 0.2b | 14.84b | 1.84b |
| 50% | 20.66c | 4bc | 4.66c | 1.09c | 0.05b | 2.75c | 0.24c |
| 75% | 8.66d | 1.66cd | 1c | 0.18c | 0b | 0.19d | 0c |
| 100% | 1.33d | 0.5d | 0c | 0c | 0b | 0d | 0c |
| 10070 | 1.554 | 0.54 | OC . | OC . | OD | ou | 00 |

contain water-soluble allelochemicals which could inhibit seed germination and seedling growth of Charlock. Similar findings were also reported by Chun-Mei et al. (2008) as they showed that rhizome, stem and leaf of ginger could inhibit seed germination and seedling growth of soybean and chive. Also Yarnia et al. (2009) found sorghum extract treatment decreased height, shoot and root dry weight and biomass of Amaranthus retroflexus. Mortality of the seedlings and reduced vigor under laboratory conditions indicated the accumulation of toxic substances (allelopathic potential) of the donor plant is harmful to the growth of seedlings of receptor plants. These findings correlated with the report of Uddin et al. (2007), Khanh et al. (2005), Mubaraki et al. (2009), Siddiqui et al. (2009) and Maharjan et al. (2007). These results suggest that Glaucium has a long-term potential reduce the of growth plants allelopathicity). This response could attributable to a greater contribution of allelochemicals from shoots. The presence of allelochemicals like alkaloids in Glaucium, may be the reason for poor germination and other growth factors of Sinapis arvensis. Several studies found that alkaloidal compounds or extracts from plants are responsible for many allelopathic effects (Elakovich et al., 1996; Petroski et al., 1990; Wink, 1983). From the present study it was concluded that various applications of Glaucium can be successfully used in weed control, especially for *Sinapis arvensis* in farms. It can also be concluded that Glaucium water extract at higher and lower concentrations suppressed the germination, root and shoot growth and root and shoot fresh and dry weight of Charlock and this suppression was possibly due to the presence of allelochemicals in Glaucium plant. Although it was a preliminary laboratory study, it provided encouraging results and a basis for future research.

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