

Green synthesis and study of antioxidant activity of benzochromene derivatives using nano KF/Clinoptilolite

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Abstract: An efficient procedure for the synthesis of benzofuran derivatives employing 2-amino-4-hydroxy acethophenone, isopropenylacetylene, aldehyde, alkyl bromides, activated acetylenic compounds and triphenylphosphine in the presence of KF/CP (NPs) as a heterogeneous base nanocatalyst in water at room temperature is investigated. Also, the antioxidant activity of some synthesized compounds was studied. The workup of reaction is simple and the products can be separated easily from the reaction mixture. KF/CP NPs show a good improvement in the yield of the product. The catalyst displayed significant reusable activity.

Keywords: Water, Aldehyde, 1-(6-hydroxy-2-isopropenyl-1-benzofuran-yl)-1-ethanone, Three-component reaction.

Introduction

Employing of green method is to find out procedure for saving resources and decrease prices. Use of ecologically solvents instead of toxic solvents and employing of moderate conditions and cheap reagents are the most attractive methods to expand a simple and green synthesis of organic compounds [1-3]. Water as an available and cheap solvent in large amounts can increase the rate of organic reactions even for compounds that is water-insoluble. Also isolation of product in water is performed by simple filtration. Catalysts have a chief function in green chemistry. It can provide the best yield of the reaction in the low temperatures. Important properties of magnetic nanoparticles (MNPs) for example large surface area to volume ratios, biocompatibility, non-toxicity and easy conversion made them gorgeous for numerous biomedical applications.

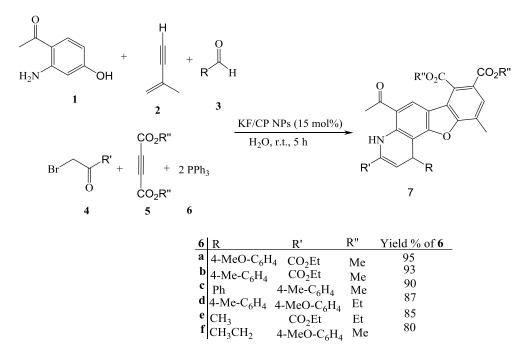
Simple recovery of MNPs by using of an external magnet because of their superparamagnetic property makes them the best catalyst for green and maintainable chemistry. Chromenes are important nucleus in organic and medicinal chemistry due to their power and broad spectrum of biological activities involving antimicrobial [4], antioxidant [5], antimalarial [6], Antibacterial [7] and anticancer [8]. Among different chromenes, benzochromenes are important and considerable compounds due to their biological properties in different subjects [9]. The synthesis of benzochromenes has been investigated in the presence of different catalysts involving lipase[10], Zn(L-proline)₂ [11], DBU [12], Triethylbenzylammonium Chloride (TEBA) [13], Et₃N [14] and 1-butyl-3-methyl imidazolium hydroxide ([bmim]OH) [15]. However, some of the reported methods have disadvantages involving high reaction times, employing of toxic and non-reusable catalyst and use of specific conditions. Consequently, the study of an efficient and available catalyst with high catalytic

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activity and short reaction time for the preparation of benzochromenes is still preferred. Certainly, the synthesis of benzochromenes via multicomponent reactions (MCR) has much notice due to good synthetic yield and easy separation of product. Lately, there has been an enhanced interest for new applications of potassium fluoride impregnated on zeolites and clays, as a new natural and inexpensive solid base system [26-34]. Among them Clinoptilolite, a natural zeolite with a high internal surface area, is much more effective because of its high exchange capability for cations particularly for K⁺, therefore, more free fluoride anions are capable of functioning as an effective base. On the other hand, the preparation of potassium fluoride impregnated Clinoptilolite (KF/CP) is very simple without the need for any pre-activation [35-36]. In continuation of our attempts to expand new synthetic procedure for chief organic compounds [16-20] herein, we investigated a "green" procedure for the synthesis of some benzofurane derivatives *via* an efficient three component reaction of 2-amino-4-hydroxy acethophenone **1**, isopropenylacetylene **2**, aldehyde **3**, alkyl bromides **4**, activated acetylenic compounds **5** and triphenylphosphine **6** in the presence of KF/CP (NPs) with good yields (Scheme 1) [24]. Moreover, the antioxidant activities of some derivatives were investigated by DPPH radical scavenging and ferric ion reducing power test.

Result and Discussion

The synthesis of some benzofurane derivatives was performed *via* an efficient multi component reaction of 2-amino-4-hydroxy acethophenone 1, isopropenylacetylene 2, aldehyde 3, alkyl bromides 4, activated acetylenic compounds 5 and triphenylphosphine 6 in the presence of KF/CP (NPs) with good yields (Scheme 1).



Scheme 1. Multi component reaction for the synthesis of benzofurane derivatives of 7 in water.

Catalytic activity of KF/CP NPs in synthesis of benzochromene derivatives

In the starting stage of this work, condensation reaction of 2-amino-4-hydroxy acethophenone 1, isopropenylacetylene 2, aldehyde 3, alkyl bromides 4, activated acetylenic compounds 5 and triphenylphosphine 6 in water at room temperature was employed as a sample reaction to achieve the optimum conditions (Table 1).

Entry	Catalyst	Temp.	catalyst	Time	Yield
		(°C)	(mol%))	(h)	% ^a
1	none	-	-	15	-
2	none	80	-	15	10
3	none	90	-	15	10
4	KF/CP NPs	80	10	5	85
5	KF/CP NPs	80	15	5	95
6	KF/CP NPs	90	15	5	95
7	KF/CP NPs	80	20	5	90
8	Et ₃ N	80	15	8	65
9	ZnO-NR	80	15	12	70
10	ZnO-NR	90	15	12	70
11	CuO-NPs	80	15	8	85
12	TiO ₂ -NPs	80	15	10	80

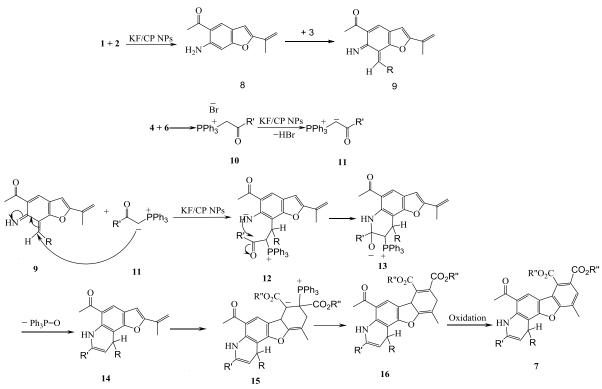
Table 1. Effect of catalyst, its loading and temperature on the condensation reaction of compound 7a

These reactions weren't performed without any catalyst even after 15h (entry 1, Table 1). By increasing the reaction temperature to 80 °C, a trace amount of 7a was generated after 15 h (entry 2, Table 1). With the purpose of get better this procedure, 10 mol% KF/CP NPs was added to the reaction mixture. After 5 h, 85% yield of 7a was produced (entry 4, Table 1). Then, the reaction was carried out in the presence of 15 mo% of KF/CP NPs as catalyst. As expected, in these conditions, the yield of product 7a was achieved in 95% yield after 5 h (entry5, Table 1). As a result, to discover the optimal catalyst loading, different amounts (10-20 mol%) of KF/CP NPs were employed. The results displayed that 15 mol% of catalyst are enough for produce an excellent yield of 7a (entry 5, Table 1). In order to more evaluate the catalytic activity, another catalyst such as ZnOnanorods, CuO-NPs, TiO₂-NPs and Et₃N were used in this reaction. Consequently, these results showed the main function of KF/CP NPs as catalyst in this reaction. In this research the effects of some solvents was also investigated on the production of 7a in the presence of 15 mol% of KF/CP NPs. The results tabulated in Table 2 display that H₂O is the best solvent for these reaction.

Table 2. Effects of solvent and temperature on generation of **7a** compound in presence of 15 mol% of KF/CP NPs.

Entry	Solvent	Temperature (°C)	Time (h)	Yield % ^a
1	EtOH	80	15	None
2	EtOH	90	15	None
3	CH_2Cl_2	-	8	75
4	CH_2Cl_2	50	8	75
5	H_2O	70	5	85
6	H ₂ O	80	5	95
7	H_2O	90	5	95
8	Solvent- free	80	8	90
10	DMf	80	15	45
11	toluene	80	12	75
12	CHCl ₃	50	10	75

According to the outcomes of optimization reported in the Tables 1 and 2, KF/CP NPs (15 mol%) as catalyst, water as solvent, and 80 °C were estimated to be the optimum reaction conditions. The reusability of the catalyst was confirmed in the model reaction (the synthesis of compound 7a). The results showed that the catalyst can be reused five times without loss of activity. After each run, the catalyst was extracted by external magnet and washed with water. It was then dried at ambient temperature for 24 h and employed for the next catalytic cycle. The structures of compounds 5 were confirmed by IR, ¹H NMR, ¹³C NMR, and mass spectral data. For example, the ¹H NMR spectrum of 7a revealed two singlets at $\delta = 2.15$ and 2.52 ppm for methyl protons, four singlets at 4.58, 5.37, 6.14 and 7.75 ppm for methin proton along with signals for aromatic moiety. In the ¹³C NMR spectrum, the signals corresponding to the carbonyl group of 7a were observed at δ 160.2 and 197.6 ppm. The IR spectrum of 7a was displayed characteristic C=O bands. Although there is no information about the mechanistic details, the reaction can be described by the mechanism proposed in Scheme 2.



Scheme 2. Proposed mechanism for the formation of 7

First, 2-amino-4-hydroxy acethophenone 1 and isopropenylacetylene 2 react together and produced intermediate 8 and react with aldehyde 3 in the presence of KF/CP NPs that is generated intermediate 9. Intermediate 9 is attacked to compound 11 in the presence of KF/CP NPs and produced Diels-Alder production 12. The chief benefits of our method are high atom economy, green reaction conditions, use a small amount nanocatalyst, higher yield, shorter reaction times, and easy work-up, which are in good agreement with some principles of green chemistry.

Investigation of antioxidant activity using DPPH

Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging test is broadly employed to estimate the ability of compounds to capture free radicals and their antioxidant activity in foods and biological systems [25-26]. The DPPH analyze donating activity of the hydrogen atom (or one electron) and gives an evaluation of antioxidant activity because of free radical scavenging. The antioxidant activity of **7a-7d** was investigated by testing their ability to the DPPH radical. DPPH radical shows the absorption in area 517 nm but its absorption decreases when is reduced by an antioxidant or a radical species. In this study, the antioxidant activity of **7a-7d** was compared to BHT and TBHQ at different concentrations from 200 mmol/L to 1000 mmol/L (Figure 1).

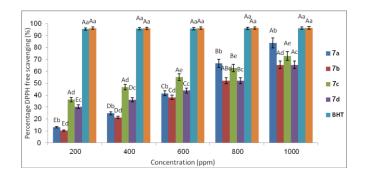


Figure 1. Radical scavenging activity (RSA) of **4a-4d** At all concentrations, the new synthesized compounds had significant differences compared to BHT and TBHQ (Figure 1). Overall, the all compounds were shown excellent free radical scavenging performance compared to BHT and TBHQ at 1000 ppm concentration (Figure 1).

Ferric ions (Fe³⁺) reducing potential (FRAP)

The ability of the synthesized compounds to reduce Ferric ions (Fe³⁺) was studied by measuring the amount of exchange of Fe³⁺/ferricyanide complex to the Fe²⁺/ ferrous shape at 700 nm. The ability of compound to reducing may act as a important indicator of its potential antioxidant activity. Compound **7a** and **7b** was displayed moderate reducing activity compared to standards (BHT and TBHQ) but **7c** and **7d** had weaker Fe⁺³ reducing potential than to **7a**, **7b**, BHT and TBHQ. It appears that the **7a** and **7b** had the 1-(4*H*-chromene-8-yl) ethanone core with stronger iron chelating potential. The results are shown in Figure **2**.

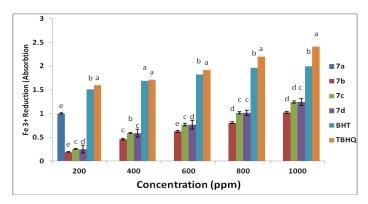


Figure 2. Ferric ions (Fe^{3+}) reducing antioxidant power (FRAP) of compounds **7a-7d.**

Conclusion

In summary, we investigate an useful, green, and environmentally procedure including 1-(6-hydroxy-2isopropenyl-1-benzofuran-yl)-1-ethanone, aldehydes, alkyl bromides and triphenylphosphin in the presence of KF/CP NPs at room temperature in water which provides a new path to the synthesis of benzochromens. The present method has many advantages such as high atom economy and yield, mild and clean reaction condition, low catalyst loading, and short reaction time. Also, the antioxidant activities of 7a-7d compounds were evaluated by DPPH radical scavenging and ferric reducing power analyzes. The compounds 7a-7d exhibit good DPPH radical scavenging activity, but showed moderate FRAP compared to synthetic antioxidants BHT and TBHQ.

Acknowledgments

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Experimental

All chemicals employed in this work were prepared from Fluka (Buchs, Switzerland), Merck, Lolitech, and Aldrich Chemical Companies and employed without further purification. KF/CP NPs were produced according to a reported method. The morphology of Fe₃O₄-MNPs was confirmed by scanning electron microscopy (SEM) employing a Holland Philips XL30 microscope. Crystalline structure of KF/CP NPs was discovered by Xray diffraction (XRD) analysis at room temperature employing a Holland Philips Xpert X-ray powder diffractometer, with CuKa radiation (1 =0.15406 nm), with 20 ranging from 20 to 80° . The elemental analyses for the determination of C, H, and N were performed employing a Heraeus CHNO-Rapid analyzer. The mass spectra were recorded on a FINNIGAN-MAT 8430 spectrometer operating at an ionization potential of 70 eV. IR spectra were measured on a Shimadzu IR-460 spectrometer. The ¹H and ¹³C NMR spectra were measured employing a Bruker DRX-500 advance spectrometer at 500.1 MHz and 125.8 MHz, respectively. The ¹H and ¹³C spectra were achieved for CDCl₃ solutions emploing TMS as the internal standard or 85 mass % H₃PO₄ as external standard; chemical shifts (d) are given as parts per million (ppm).

General procedure for preparation of compounds 7a-f

2-amino-4-hydroxy acethophenone 1(2 mmol). isopropenylacetylene 2 (2 mmol) and KF/CP NPs (15 mol%) in water (3 mL) was at room temperature for an appropriate time showed in Table 3. Aldehyde 3 (2 mmol) was added to previous mixture. The alkyl bromides 4 (2 mmol) react with triphenylphosphine 6 (2 mmol) in another pot and added to first pot. Finally activated acetylenic compounds 5 (2 mmol) and triphenylphosphine 6 (25 mmol) was added and after completion the reaction, the KF/CP NPs were separated by filteration. The organic and aqueous layers were separated by filtration and washed with Et_2O to afforded pure title compound 7.

Compound 7a: Yellow powder, mp 173-175°C, Yield: 1.08 g (95%). IR (KBr) (v_{max} /cm⁻¹): 1742, 1735, 1683, 1585, 1462, 1274 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 1.32 (3 H, t, ³*J* = 7.4 Hz, CH₃), 2.15 (3 H, s, Me), 2.52 (3 H, s, Me), 3.75 (3 H, s, MeO), 3.85 (3 H, s, MeO), 4.03 (3 H, s, MeO), 4.26 (2 H, q, ³*J* = 7.3 Hz, CH₂O), 4.47 (1 H, d, ²*J* = 4.5 Hz, CH), 5.87 (1 H, d, ²*J* = 4.5 Hz, CH), 7.12 (2 H, d, ³*J* = 7.6 Hz, 2 CH), 7.63 (2 H, d, ³*J* = 7.6 Hz, 2 CH), 7.75 (1 H, s, CH), 8.13 (1 H, s, CH) ppm. ¹³C NMR (125.7 MHz, CDCl₃): 14.2 (Me), 22.6 (Me), 30.2 (Me), 41.2 (CH), 51.2 (MeO), 52.3 (MeO), 55.7 (MeO), 61.4 (CH₂O), 109.3 (2 CH), 113.2 (C), 114.3 (C), 115.6 (CH), 123.4 (C), 124.2 (C), 124.8 (CH), 127.2 (C), 127.3 (2 CH), 127.6 (CH), 128.2 (C), 130.5 (C), 135.4 (C), 145.2 (C), 153.7 (C), 159.2 (C), 159.8 (C), 160.3 (C), 161.2 (C=O), 165.3 (C=O), 166.4 (C=O), 197.6 (C=O) ppm. EI-MS: 572 (M⁺, 15), 529 (86), 43 (100). Anal. Calcd for $C_{32}H_{28}O_{10}$ (572.56): C 67.13, H 4.93; Found: C 67.34, H 5.18.

Compound 7b: Yellow powder, mp 162-164°C, Yield: 1.03 g (93%). IR (KBr) (v_{max}/cm^{-1}): 1740, 1735, 1726, 1683, 1575, 1472, 1295 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 1.34 (3 H, t, ${}^{3}J = 7.4$ Hz, CH₃), 2.16 (3 H, s, Me), 2.23 (3 H, s, Me), 2.53 (3 H, s, Me), 3.87 (3 H, s, MeO), 4.05 (3 H, s, MeO), 4.25 (2 H, q, ${}^{3}J = 7.4$ Hz, CH₂O), 4.65 (1 H, d, ${}^{2}J$ = 4.5 Hz, CH), 6.12 (1 H, d, ${}^{2}J = 4.5$ Hz, CH), 7.32 (2 H, d, ${}^{3}J = 7.6$ Hz, 2 CH), 7.58 (2 H, d, ${}^{3}J$ = 7.6 Hz, 2 CH), 7.78 (1 H, s, CH), 8.25 (1 H, s, CH) ppm. ¹³C NMR (125.7 MHz, CDCl₃): 13.5 (Me), 22.0 (Me), 22.7 (Me), 30.4 (Me), 40.8 (CH), 51.4 (MeO), 52.6 (MeO), 61.6 (CH₂O), 112.3 (C), 114.2 (C), 115.8 (C), 124.2 (2 CH), 124.8 (C), 125.2 (CH), 126.3 (2 CH), 127.2 (C), 127.8 (CH), 128.2 (C), 128.6 (C), 130.2 (C), 134.3 (C), 136.2 (C), 145.5 (C), 153.2 (C), 158.4 (C), 159.6 (C), 160.7 (C=O), 166.3 (C=O), 167.2 (C=O), 197.4 (C=O) ppm. EI-MS: 556 (M⁺, 15), 513 (64), 43 (100). Anal. Calcd for C₃₂H₂₈O₉ (556.56): C 69.06, H 5.07; Found: C 69.22, H 5.24.

Compound 7c: Yellow powder, mp 175-177°C, Yield: 1.008 g (90%). IR (KBr) (v_{max}/cm^{-1}): 1742, 1738, 1725, 1692, 1563, 1485, 1274 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 2.15 (3 H, s, Me), 2.35 (3 H, s, Me), 2.54 (3 H, s, Me), 3.83 (3 H, s, MeO), 3.96 (3 H, s, MeO), 4.74 (1 H, d, ${}^{2}J = 4.5$ Hz, CH), 6.12 (1 H, d, ${}^{2}J$ = 4.5 Hz, CH), 6.75 (1 H, t, ${}^{3}J$ = 7.5 Hz, CH), 7.12 (2 H, t, ${}^{3}J = 7.5$ Hz, 2 CH), 7.35 (2 H, d, ${}^{3}J = 7.8$ Hz, 2 CH), 7.46 (2 H, d, ${}^{3}J$ = 7.5 Hz, 2 CH), 7.73 (2 H, d, ${}^{3}J$ = 7.8 Hz, 2 CH), 7.82 (1 H, s, CH), 8.32 (1 H, s, CH) ppm. ¹³C NMR (125.7 MHz, CDCl₃): 16.7 (Me), 21.6 (Me), 30.5 (Me), 42.3 (CH), 51.5 (MeO), 52.7 (MeO), 102.4 (CH), 112.3 (C), 114.4 (C), 115.3 (CH), 124.2 (2 CH), 124.6 (CH), 125.2 (2 CH), 125.6 (C), 126.5 (2 CH), 127.2 (CH), 127.6 (C), 128.4 (2 CH), 128.8 (C), 130.2 (C), 131.3 (C), 131.8 (C), 134.2 (C), 136.5 (C), 153.3 (C), 153.8 (C), 159.3 (C), 160.3 (C), 166.3 (C=O), 167.2 (C=O), 197.5 (C=O) ppm. EI-MS: 560 $(M^+, 15), 517 (64), 43 (100)$. Anal. Calcd for $C_{35}H_{28}O_7$ (560.59): C 74.99, H 5.03; Found: C 75.12, H 5.18.

Compound 7d: Yellow powder, mp 187-189°C, Yield: 1.07 g (87%). IR (KBr) (v_{max}/cm⁻¹): 1738, 1735, 1725, 1692, 1578, 1487, 1295 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 1.12 (3 H, t, ${}^{3}J = 7.4$ Hz, CH₃), 1.28 (3 H, t, ${}^{3}J$ = 7.4 Hz, CH₃), 2.12 (3 H, s, Me), 2.17 (3 H, s, Me), 2.56 (3 H, s, Me), 3.87 (3 H, s, MeO), 4.26 (2 H, q, ${}^{3}J$ = 7.4 Hz, CH₂O), 4.32 (2 H, q, ${}^{3}J$ = 7.4 Hz, CH₂O), 4.82 (1 H, d, ${}^{2}J = 4.7$ Hz, CH), 5.24 (1 H, d, ${}^{2}J = 4.7$ Hz, CH), 7.32 (2 H, d, ${}^{3}J$ = 7.6 Hz, 2 CH), 7.38 (2 H, d, ${}^{3}J = 7.6$ Hz, 2 CH), 7.62 (2 H, d, ${}^{3}J = 7.6$ Hz, 2 CH), 7.75 (2 H, d, ${}^{3}J$ = 7.6 Hz, 2 CH), 7.85 (1 H, s, CH), 8.35 (1 H, s, CH) ppm. ¹³C NMR (125.7 MHz, CDCl₃): 13.8 (Me), 14.2 (Me), 16.8 (Me), 21.8 (Me), 30.6 (Me), 42.4 (CH), 55.6 (MeO), 61.3 (CH₂O), 62.4 (CH₂O), 102.3 (CH), 112.3 (C), 114.5 (2 CH), 114.9 (C), 115.3 (CH), 124.2 (2 CH), 124.7 (CH), 125.2 (C), 125.8 (2 CH), 126.3 (C), 127.2 (C), 128.2 (2 CH), 128.7 (C), 129.3 (C), 129.8 (C), 134.2 (C), 135.8 (C), 153.6 (C), 154.2 (C), 158.3 (C), 159.2 (C), 159.8 (C), 160.2 (C=O), 164.3 (C=O), 198.2 (C=O) ppm. EI-MS: 618 (M⁺, 20), 575 (62), 43 (100). Anal. Calcd for C₃₈H₃₄O₈ (618.67): C 73.77, H 5.54; Found: C 73.85, H 5.65.

Compound 7e: Yellow powder, mp 135-137°C, Yield: 0.86g (85%). IR (KBr) (v_{max} /cm⁻¹): 1743, 1738, 1735, 1725, 1642, 1587, 1468, 1275 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 1.28 (3 H, t, ${}^{3}J = 7.4$ Hz, CH₃), 1.35 (3 H, t, ${}^{3}J = 7.4$ Hz, CH₃), 1.42 (3 H, t, ${}^{3}J = 7.4$ Hz, CH₃), 1.56 (3 H, d, ${}^{3}J$ = 7.4 Hz, CH₃), 2.43 (3 H, s, Me), 2.56 $(3 \text{ H}, \text{ s}, \text{ Me}), 3.85-3.92 (1 \text{ H}, \text{ m}, \text{CH}), 4.18 (2 \text{ H}, \text{ q}, {}^{3}J =$ 7.4 Hz, CH₂O), 4.25 (2 H, q, ${}^{3}J = 7.4$ Hz, CH₂O), 4.32 $(2 \text{ H}, \text{q}, {}^{3}J = 7.4 \text{ Hz}, \text{CH}_{2}\text{O}), 5.73 (1 \text{ H}, \text{d}, {}^{2}J = 4.0 \text{ Hz},$ CH), 7.54 (1 H, s, CH), 8.27 (1 H, s, CH) ppm. ¹³C NMR (125.7 MHz, CDCl₃): 13.5 (Me), 13.8 (Me), 14.2 (Me), 16.5 (Me), 21.2 (Me), 29.5 (Me), 31.2 (CH), 61.6 (CH₂O), 62.4 (CH₂O), 62.8 (CH₂O), 111.4 (C), 114.8 (C), 115.2 (CH), 123.7 (C), 124.2 (CH), 127.2 (C), 128.6 (C), 129.2 (CH), 129.8 (C), 134.6 (C), 143.7 (C), 153.6 (C), 159.6 (C), 160.4 (C), 161.2 (C=O), 164.7 (C=O), 170.5 (C=O), 198.3 (C=O) ppm. EI-MS: 508 (M⁺, 10), 465 (68), 43 (100). Anal. Calcd for C₂₈H₂₈O₉ (508.52): C 66.13, H 5.55; Found: C 66.26, H 5.

Compound 7f: Yellow powder, mp 139-141°C, Yield: 0.84g (80%). IR (KBr) (v_{max} /cm⁻¹): 1745, 1742, 1738, 1695, 1595, 1487, 1292 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): 1.12 (3 H, t, ³*J* = 7.2 Hz, CH₃), 1.48-1.57 (1 H, m, CH), 1.65-1.78 (1 H, m, CH), 2.45 (3 H, s, Me), 2.52 (3 H, s, Me), 3.79-3.87 (1 H, m, CH), 3.87 (3 H, s, MeO), 3.96 (3 H, s, MeO), 4.06 (3 H, s, MeO), 5.78 (1 H, d, ²*J* = 4.2 Hz, CH), 7.28 (2 H, d, ³*J* = 7.6 Hz, 2 CH), 7.65 (2 H, d, ${}^{3}J$ = 7.6 Hz, 2 CH), 7.87 (1 H, s, CH), 8.23 (1 H, s, CH) ppm. 13 C NMR (125.7 MHz, CDCl₃): 12.8 (CH₃), 16.7 (Me), 29.2 (CH₂), 29.8 (Me), 30.8 (CH), 51.3 (MeO), 52.4 (MeO), 55.8 (MeO), 103.5 (CH), 111.5 (C), 113.8 (2 CH), 114.5 (C), 115.3 (C), 123.4 (C), 124.2 (CH), 125.3 (C), 126.4 (C), 127.2 (2 CH), 128.2 (C), 130.4 (C), 134.2 (C), 151.2 (C), 156.3 (C), 158.3 (C), 159.7 (C), 160.2 (C), 168.2 (C=O), 169.5 (C=O), 197.2 (C=O) ppm. EI-MS: 528 (M⁺, 15), 525(86), 43 (100). Anal. Calcd for C₃₁H₂₈O₈ (528.55): C 7044, H 5.34; Found: C 77.46, H 6.38.

1,1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging test

Radical scavenging activity of 7a-7d was measured by DPPH 2-diphenyl-1-picrylhydrazyl) radical (2,scavenging test according to the reported method by Shimada et al.[38]. Different concentrations of 7a-7d (200-1000 ppm) were added to an equal volume of methanolic solution of DPPH (1 mmol/L). The mixtures were well shaken and then placed in a dark room. After 30 min at room temperature, the absorbance was recorded at 517 nm. In the control sample, 7a-7d were replaced with 3 mL methanol. hydroxytoluene Butylated (BHT) and 2tertbutylhydroquinone (TBHQ) were used as standard controls. The percentage inhibition of the DPPH radical was calculated according to the formula of Yen and Duh [39].

Reducing power test

The ability of compounds **7a-7d** to reduce iron (III) was evaluated by the method of Yildirim et al. [40] Samples (1 mL) were mixed with 2.5 mL of phosphate buffer (0.2 mol/L, pH 6.6) and 2.5 mL of potassium ferricyanide (K3Fe(CN)6; 10g/L) and showed for 30 min at 50 8C. Then, 2.5 mL of trichloroacetic acid (10% w/v) were added to the solution and centrifuged for 10 min. Finally, 2.5 mL of supernatant was combined with 2.5 mL of distilled water and 0.5 mL FeCl₃ (1 g/L). The absorbance of samples was measured at 700 nm. Higher absorbance means higher reducing power.

Each measurement was carried out in triplicate. The data were analyzed by running one way analysis of variance (ANOVA) using SPSS software version 18.0. A one way ANOVA was employed to evaluate difference in the mean value of samples and control. All mean separations were performed by Duncan

multiple range test using the significance level of 95% (P < 0.05).

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