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Antimicrobial activity of Ethanolic extracts of (*Tragopogon collinus* L. and *Allium eriophyllum* L.) on some selected environmental and pathogenic bacteria

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Abstract

Due to the increase in the incidence of infectious diseases and the prevalence of antibiotic resistance in bacteria in different environments, there is an urgent need to discover new antimicrobial compounds. The present study aimed to determine the antibacterial activity of the ethanolic extracts of the edible plants Tragopogon Collinus L. and Allium eriophyllum L. against some environmental and pathogenic bacteria such as Staphylococcus aureus, Pseudomonas aeruginosa, Klebsiella pneumoniae, Escherichia coli, and Enterococcus faecalis. The extraction of medicinal plants was conducted using ethanol and the percolation method. The well diffusion method was used to determine the antimicrobial effects of the extracts, and the minimum inhibitory concentration (MIC) of microbial growth. The ethanol extract of T. collinus showed an inhibitory effect on all bacteria in 250 µg/ml concentration, and the greatest effect was observed on S. aureus and E. faecalis, with 20 and 18 mm diameter of zone, respectively. In the 125 μ g/ml concentration, with a 21 mm diameter of zone, it had the highest antimicrobial effect on K. pneumoniae, compared with other bacteria. Regarding the ethanolic extract of A. eriophyllum L., the highest antibacterial effect at 250 µg/ml concentration was 20 and 22 mm zone diameters. The effect of this plant extract on P. aeruginosa was lower than its effect on other bacteria. The MIC of the extract of *Tragopogon collinus* L.was found at a 5.75 μ /ml concentration on *S. aureus* and *E. faecalis*. The MIC of the ethanolic extract of A. eriophyllum L. was found at 11.5 µ/ml concentration on S. aureus, P. aeruginosa, and K. pneumoniae. The results of this study indicate that T. collinus and A. eriophyllum have antibacterial properties.

Key words: Antibacterial activity, Ethanolic extract, Tragopogon collinus L., Allium eriophyllum L

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1. Introduction

Antimicrobial resistance is a global public health crisis (Pierce et al., 2021). Plants contain many biologically active natural materials. Although these bioactive compounds are not nutritional, they have important medicinal properties (Ivanišová et al., 2021). Plants are important natural materials used in complementary medicine (Saleh Mohammed et al., 2022). Indeed, plants have been used as medicines for more than 5000 years, as a source of antibiotics, antineoplastic, analgesics, cardio protective. (Newman and Cragg., 2016). About 70–90% of the population in developing countries continues to use ancient medicines based on plant extracts (Anand et al., 2019).

Tragopogon collinus is an herbaceous plant from the family of chicory (Maleki et al., 2021) The compounds identified in these plants are flavonoids, terpenes, saponins, benzyl and hydroisocoumarin, phenolic compounds, and sterols, in which, many of them have been identified during chemotaxonomic studies of these plants. Some plants in the TPE genus, by their strong antioxidant effect, prevent inflammatory damage to tissues and in some concentrations prevent DNA damage (Farzaei et al., 2014). This plant has chemical compounds such as carbon hydrate and inulin, which are used in traditional medicine as an appetize.

Allium eriophyllum is from the garlic family. It is actually a type of mountain garlic (Zhao et al., 2022). It has chemical compounds composed of sulfur and polysulfuronyl polyvinyl sulfate and a type of unstable aldehyde, and its essential oil contains sulfur compounds. The therapeutic properties of Allium eriophyllum essential oil are the same as those of garlic essential oil since its compounds are similar to those of garlic. Because of the presence of large amounts of cysteine sulfoxides and sulfoxide amino acids in it, this plant also has anti-diabetic and antioxidant properties (Mosavi et al., 2019). Garlic was shown to have anti-diabetic, antihypertensive, antihyperlipidemic, antiplatelet, anticoagulant, antiatherosclerotic, and anti-oxidative stress properties (Mozafari et al., 2017).

Many studies by different researchers have

reported that plants have many important biological activities such as antioxidant, antimicrobial. anti-aging, anticancer, antitumor, DNA protective, antiallergic (Nisa et al., 2019; Zazharskyi et al., 2019; Nguyen et al., 2020; Dias et al., 2021; Akgül et al., 2022). Tragopogon collinus and Allium eriophyllum grow naturally in the western regions of Iran and are edible. The nomads and people of the western region of Iran, especially Kermanshah province, have used these medicinal plants without knowing enough about their antimicrobial properties. Since these plants have been used in society for centuries, this study aimed to investigate the antibacterial effects of their ethanolic extract on some indicator and environmental bacteria.

2. Materials and Methods 2.1. Microbial Strains

In this study, five standard strains, including *E.coli* ATCC 25922, *Staphylococcus aureus* ATCC 25923, *Pseudomonas aeruginosa* ATCC 27853, *Klebsiella pneumoniae* ATCC 10031, and *Enterococcus faecalis* ATCC 9854 were purchased from the Persian Type Culture Collection (PTCC, IROST, Tehran/Iran). Lyophilized vials were cultured under sterile conditions according to the manufacturer's instructions. In addition, five environmental isolates of the same strains kept in the microbiology laboratory of the Lahijan Branch, Islamic Azad University were used in this study.

2.2. Collection of Plants and Preparation of Ethanolic Extract

Plants were collected from their natural habitat (Dalahoo mountains Kermanshah province, Iran), dried in the air, and then kept in shade for a week. Identification of collected plants was performed in the herbarium of botany in the Gorgan Branch, Islamic Azad University, Gorgan, Iran.

For preparing the extracts, the leaves of the dried plants were powdered by an electric mill, then 200 grams of powder was carefully weighed and used to extract by the percolation method. Then, ethanol was eliminated using a rotary evaporator and vacuum distillation method. This extract was considered a pure extract, of *Tragopogon collinus* and *Allium eriophyllum* plants. Then, the concentrations of 250, 125, 62.5, 31.25,





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15.62, 7.81, 3.9, 1.95, 0.97, and 0.48 μ g/ml using 100% Dimethyl Sulfoxide (DMSO) were prepared by serial dilution.

2.3. Examining the Antibacterial Effect

After preparing the extracts, the agar well diffusion method was used to determine antibacterial activities, and serial dilution was used to determine the MIC of the extracts.

2.3.1. Well Diffusion Method

In the agar well diffusion method, a microbial suspension the turbidity equivalent to McFarland 0.5 was prepared from the 24-hour bacterial cultivation in Mueller Hinton broth and then cultured on the Mueller Hinton agar plates. Using a cork borer, wells with a diameter of 7 mm were made. 100 microliters of different concentrations of the extracts were poured into the wells and then incubated for 24 hours at 37 °C. By measuring the diameter of the bacteria to the extracts was determined. In this study, gentamicin and 100% DMSO were used as positive control and negative controls, respectively

2.3.2. Minimum inhibitory concentration determination

A serial dilution was prepared from the plant extracts used in the 100% DMSO solution. For this purpose, 5 ml of primary extract of plants (500 μ g/ml concentration) was added to the tubes containing 100% DMSO until it was completely dissolved, and a uniform solution was prepared. Plants were dissolved in 15 ml of 100% DMSO solution. For this reason, the initial concentration of the extracts of Tragopogon collinus and Allium eriophyllum was determined to be from 0.48 μ g/ml to 250 μ g/ml.After autoclaving, the Erlenmeyer flasks containing Mueller Hinton broth were cooled to 50°C, and 5mL of different concentrations of the prepared extracts were added to each Erlenmeyer flask. Then shaken gently so that the extract was completely distributed in the medium. In this way, the final concentrations of extracts in Mueller Hinton Agar medium were estimated to be 23, 11.5, 5.75, 2.87, 1.43, 0.71, 0.35, 0.17, 0.08, and 0.04 µg/ml. Then, suspensions from each bacterial equal to 0.5 McFarland standards were prepared. Twenty microliters of each concentration were inoculated on the surface of each plate. Plates were incubated for 24 hours at 37°C, and bacterial growth was checked for each concentration. The lowest concentration of the extract inhibited the growth considered MIC. All experiments were performed in triplicates.

3. Results

The ethanolic extract of the *Tragopogon collinus* showed an inhibitory effect on all bacteria at a concentration of 250 µg/ml. This effect on *S.aureus* and *E. faecalis* was the greatest among the bacteria, with inhibition zone diameters of 20 and 15 mm, respectively. The least amount of concentration of the extract that had a significant inhibitory effect (\geq 12mm) was 3.9 µg/ml and 0.97 µg/ml against *S. aureus* and *E. faecalis* and were 1.95, 62.5, and 31.25 µg/ml on *P. aeruginosa, E. coli*, and *K. pneumoniae*, respectively (Table 1).

 Table 1. Diameter of Inhibition zone of ethanolic extract of Tragopogon collinus against bacteria in comparison with gentamicin using agar well diffusion method

| | | | | con | centratio | ns (µg/ | ml) | | | | | |
|------------------------|-----|-----|------|-------|-----------|---------|-----|------|------|------|------------------|----------|
| Diameter of inhibition | 250 | 125 | 62.5 | 31.25 | 15.62 | 7.81 | 3.9 | 1.95 | 0.97 | 0.48 | Positive control | Negative |
| zone | | | | | | | | | | | 1.25 µg/ml | control |
| Bacteria | | | | | | | | | | | | |
| staphylococcus aureus | 20 | 18 | 19 | 19 | 16 | 13 | 13 | 10 | 10 | 9 | 27 | - |
| pseudomonas aeruginosa | 15 | 14 | 13 | 12 | 11 | 10 | - | - | - | - | 18 | - |
| Escherichia coli | 14 | 15 | 13 | 10 | 10 | 10 | 8 | - | - | - | 28 | - |
| Klebsiella pneumonaie | 15 | 21 | 20 | 17 | 11 | - | - | - | - | - | 28 | - |
| Enterococcus faecalis | 18 | 18 | 18 | 18 | 15 | 13 | 13 | 12 | 12 | 10 | 28 | - |

Note. (-) indicates neutral; Negative control: DMSO 100%; Positive control: Gentamicin





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K. pneumoniae and *S. aureus* showed the greatest inhibition zone when treated with ethanolic extracts of *Allium eriophyllum*; however, the diameter of this zone was the same as the zone caused by gentamicin. The zone diameter ≥ 12 mm was not observed in any case at a concentration lower than 7.8 µg/ml (Table 2).

 Table 2. Inhibition zone diameter of the Ethanolic extract of Allium eriophyllum against bacteria in comparison with gentamicin using agar gel diffusion method.

| | | | | Concen | tration (µ | ıg/ml) | | | | | | |
|------------------------|-----|-----|------|--------|------------|--------|-----|------|------|------|------------------|----------|
| Zone diameter of the | 250 | 125 | 62.5 | 31.25 | 15.62 | 7.81 | 3.9 | 1.95 | 0.97 | 0.48 | Positive control | Negative |
| bacterium | | | | | | | | | | | 1.25 µg/ml | control |
| Staphylococcus aureus | 20 | 19 | 17 | 15 | 13 | 12 | 10 | - | - | - | 27 | - |
| pseudomonas aeruginosa | 16 | 15 | 15 | 13 | 12 | 11 | 10 | 8 | - | - | 18 | - |
| Escherichia coli | 17 | 16 | 15 | 12 | 10 | 8 | 7 | - | - | - | 28 | - |
| Klebsiella pneumoniae | 22 | 20 | 18 | 14 | 11 | 9 | 8 | - | - | - | 28 | - |
| Enterococcus faecalis | 18 | 17 | 15 | 14 | 13 | 12 | 10 | - | - | - | 28 | - |

Note. (-) indicates neutral; Negative control: DMSO 100%; Positive control: Gentamicin

The determined MIC concentrations of the ethanolic extracts of Tragopogon collinus and Allium eriophyllum against E. faecalis and S. aureus, P. aeruginosa, E. coli and K. pneumoniae are shown in Table 3.

 Table 3. The MIC concentrations of ethanolic extracts of Tragopogon collinus and Allium eriophyllum against bacterial strains

| | Plant Extract MIC µg/ml | | | | | | |
|------------------|-------------------------|--------------------|--|--|--|--|--|
| Type of bacteria | Tragopogon collinus | Allium eriophyllum | | | | | |
| S. aureus | 11.5 | 5.75 | | | | | |
| P. aeruginosa | 11.5 | 11.5 | | | | | |
| E. coli | 23 | 11.5 | | | | | |
| K. pneumoniae | 11.5 | 11.5 | | | | | |
| E. faecalis | 23 | 5.75 | | | | | |

4. Discussion

The therapeutic agents derived from plants are justified by the emergence of diseases and the growth of scientific knowledge about herbal medicines as important alternatives or complementary treatments for diseases (Sasidharan et al., 2011). The discovery of new antibacterial agents capable of dealing with antibiotic-resistant bacteria is a priority in the field of scientific research (Martinez et al., 2021 a). In this field, certain natural compounds of plant origin have emerged as powerful potential therapeutic tools (Martinez et al., 2021 b). Many studies have shown that medicinal plants contain coumarins, flavonoids, phenolics, alkaloids, terpenoids, tannins, essential oils, lectins, polypeptides, and polyacetylenes. These bioactive compounds are used as a starting point for antibiotics synthesis to treat infectious diseases (Parvin et al., 2014). Plants produce an invaluable source of secondary metabolites in response to environmental factors such as attack by herbivores, abiotic stress, or interspecific interactions (Yang et al., 2018). Since ancient times, humans have used these secondary metabolites of plants in various fields, including medicine and gastronomy. The vast chemical diversity of plant secondary metabolites and their long history of traditional use make plants very attractive natural reservoirs for research into the discovery of new antimicrobial compounds. Rapid technological advancement and the application of new, increasingly efficient methodologies have allowed the identification and characterization of numerous antibacterial agents in recent years (Katz and Baltz, 2016).

In Iran, medicinal plants have not been exploited and used in the same way as they are in developed countries, so most people do not know





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about the valuable plants scattered throughout Iran or they are not familiar with their healing properties. Although edible plants may have therapeutic effects, they may not be beneficial to the body when consumed in such a way. The lack of knowledge of their existence in Iran or the lack of recognition of these plants is still the biggest obstacle to their use (Zargari, 1996).

In this study, two plant extracts were used because the properties of these plants remain unknown, and they are widely used for medicinal and edible purposes, especially in the west of Iran. Because there are currently no scientific studies that examine the antimicrobial properties of these plants' extracts, this research is of great significance. Examining the antimicrobial effects of the ethanolic extract of these plants on bacterial strains is important.

The results of the present study showed that the ethanol extracts of plants at different concentrations had antibacterial effects on all bacteria tested.

According to the results of this research, ethanol extracts from these plants worked differently against different types of bacteria depending on the concentration used. The extract of the Tragopogon collinus had the most significant antibacterial effect on S. aureus in 250 µg/ml concentration with a 20 mm zone diameter. Moreover, in the 125 μ g/ml concentration, it had the most significant antibacterial effect on K. pneumoniae with a 21 mm inhibitory growth zone diameter. The ethanol extract of the Allium eriophyllum had the most significant antibacterial effect on K. pneumoniae at 250 µg/ml concentration with a 22 mm inhibitory growth zone diameter. The results of the antibacterial effect of the Tragopogon collinus and Allium eriophyllum on S. aureus were also significant, whereby the most significant effects of the two plants were obtained in 250 µg/ml concentration with 20 mm inhibitory growth zone diameter. In determining the MIC of these plants on S. aureus, Tragopogon collinus extract at a concentration of 5.75 µg/ml showed an inhibitory effect on S. aureus, while the MIC of Allium eriophyllum extracts on this bacterium at a concentration of 11.5 µg/ml was obtained. Hence, an extract of Tragopogon collinus with a lower concentration was more effective against this bacterium. Therefore, it can be used as a candidate drug for treating *S. aureus*, one of the most important infectious agents.

Regarding *E. coli*, the antibacterial effect of the ethanolic extract of *Allium eriophyllum* was greater than that of *Tragopogon collinus* in 250 μ g/ml concentration, with a 17 mm zone diameter compared with that of 14 mm in *Tragopogon collinus*. But in determining the MIC, the extract of *Tragopogon collinus* at a concentration of 11.5 μ g/ml showed a greater inhibitory effect on E. coli compared with the extract of Allium erio*phyllum* (at a concentration of 23 μ g/ml).

The antimicrobial effects of the ethanolic extracts of Tragopogon collinus and Allium eriophyllum on E. faecalis were also significant. Tragopogon collinus extracts had similar effects in concentrations of 250 to 3.25 μ g/ml with a zone diameter of 18 mm, and the extract of this plant had antimicrobial activity on E. faecalis up to 0.97 µg/ml concentration. In contrast, Allium eriophyllum extract had the greatest effect only at a concentration of 250 µg/ml with a zone diameter of 18 mm, and with decreasing concentrations, the diameter of the growth zone decreased, and in other concentrations, it had weaker effects compared to the Tragopogon collinus extract. Examining the obtained concentrations of the ethanol extracts of Tragopogon collinus and Allium eriophyllum to determine their MICs showed that the MIC of Tragopogon collinus extract had significant inhibitory effects on E. faecalis at a concentration of 5.75 μ g/ml. On the other hand, the MIC of Allium eriophyllum extract on E. faecalis was observed at a concentration of 23 µg/ml, which had a weaker inhibitory effect on this bacterium compared to the Tragopogon collinus extract. Investigating the antibacterial effect of the ethanolic extract of Tragopogon collinus and Allium eriophyllum on P .aeruginosa, it was found that the highest effect was related to the Allium eriophyllum extract at a concentration of 250 µg/ml with a zone diameter of 16 mm, and as the concentration decreased, its effect on the bacteria decreased, and the diameter of the non-growth zone diameter decreased. The highest effect related to the extract of the Tragopogon collinus was ob-





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served on *P. aeruginosa* at a concentration of 250 μ g/ml, and the non-growth zone diameter was 15 mm. The lowest concentration of this extract had antibacterial properties on

P. aeruginosa was $31.25 \ \mu g/ml$, which showed weaker effects compared to the lowest concentration of *Allium eriophyllum* extract, at a concentration of $15.62 \ \mu g/ml$. The MIC of both plant extracts on *P. aeruginosa* was observed to be the same at a concentration of $11.5 \ \mu g/ml$.

The results of the investigation into the antibacterial effects of the extract of Tragopogon collinus and Allium eriophyllum on K. pneumoniae showed that the most antibacterial effect of the ethanolic extract of Tragopogon collinus was at a concentration of 125 µg/ml with a growth inhibition zone of 11 mm. On the other hand, the greatest antibacterial effect of the ethanolic extract of Allium eriophyllum was at a concentration of 250 μ g/ml with a growth inhibition zone of 22 mm. Moreover, the decrease in the concentration of the extract of this plant reduced the non-growth zone diameter of this bacterium. The MIC of the ethanol extract of Tragopogon collinus and Allium eriophyllum on K.pneumoniae was the same and was observed at a concentration of 11.5 μ g/ ml. In a study conducted abroad by Zurideh et al. 2008, it was shown that the methanolic extract of Nigella sativa plant had an inhibitory effect in different concentrations on two pathogenic bacteria S. aureus and P. aeruginosa, but had no antimicrobial effect on two pathogenic bacteria, E. coli and K. pneumoniae (Zuridah et al., 2008). In another study by Abere et al. (2007), it was found that the methanolic extract of the leaves of the Mitracarpus scaber plant had an inhibitory effect on the bacteria Staphylococcus aureus, K.pneumoniae, and E. coli at a concentration of 75 µg/ ml, while no antimicrobial effect was observed on *P. aeruginosa* at the same concentration. Our results showed that the ethanolic extracts of Tragopogon collinus and Allium eriophyllum had more antibacterial effects than their methanolic extracts. It should be noted that the results obtained from the investigation of the antibacterial effects of the extracts of Tragopogon collinus and Allium eriophyllum on the bacteria used in the present study showed lower effects compared

to the antibiotic gentamicin, which was used as a positive control in this research with a concentration of 1.25 μ g/ml. This is due to the use of pure and complete extracts of these plants, which contain countless mixtures of substances that are less effective than the antibiotic gentamicin, and if the combination of substances with the antimicrobial effect found in these extracts is isolated and purified, the effects will be more comparable to gentamicin.

In a general conclusion, it can be stated that the ethanolic extracts of *Tragopogon collinus* and *Allium eriophyllum* have demonstrated antibacterial activity on the studied strains in vitro, and it is necessary to conduct a wider and more extensive study in vitro so that the effective doses of these extracts on these bacteria and clinical strains, as well as their precise formulation to achieve maximum bioavailability, can be determined, and, finally, these extracts can be introduced as a new and potentially effective antimicrobial drug to the world of medicine and microbiology. This research clearly shows that medicinal plants have an antibacterial effect, which validates their use in traditional medicine.

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References

Álvarez-Martínez, F. J., Barrajón-Catalán, E., Herranz-López, M., & Micol, V. (2021)a. Antibacterial plant compounds, extracts and essential oils: An updated review on their effects and putativemechanisms of action. Phytomedicine, 90, 153626.

Álvarez-Martínez, F. J., Rodríguez, J. C., Borrás-Rocher, F., Barrajón-Catalán, E., & Micol, V. (2021)b. The antimicrobial capacity of *Cistus salviifolius* and *Punica* granatum plant extracts against clinical pathogens is related to their polyphenolic composition. Scientific reports, 11(1), 1-12.

Anand U, Jacobo-Herrera N, Altemimi A, Lakhssassi N. A Comprehensive Review on Medicinal Plants as Antimicrobial Therapeutics: Potential Avenues of Biocompatible Drug Discovery. Metabolites. 2019; 9(11):258.

Akgül H, Mohammed FS, Kına E, Uysal İ, Sevindik M,





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Doğan M. 2022. Total Antioxidant and Oxidant Status and DPPH Free radical activity of *Euphorbia eriophora*. Turk-ish Journal of Agriculture-Food Science and Technology, 10(2): 272- 275.

ATEŞ, D. A., & TURGAY, Ö. (2003). Antimicrobial activities of various medicinal and commercial plant extracts. Turkish Journal of Biology, 27(3), 157-162.

Akhond Zadeh Basti, A., (2000). Iranian Medical Plant Encyclopedia (1ST ed.). Tehran: Arjmand publication.

Burt, S. (2004). Essential oils: their antibacterial properties and potential applications in foods—a review. International Journal of Food Microbiology, 94(3), 223-253.

Capasso, L. (1998). 5300 years ago, the Ice Man used natural laxatives and antibiotics. The Lancet, 352(9143), 1864. Cowan, M. M. (1999). Plant products as antimicrobial agents. Clinical Microbiology Reviews, 12(4), 564-582.

Dias MC, Pinto DC, Silva A. 2021. Plant flavonoids: Chemical characteristics and biological activity. Molecules, 26(17): 5377.

Farzaei, M. H., Rahimi, R., Attar, F., Siavoshi, F., Saniee, P., Hajimahmoodi, M., Mirnezami, T., & Khanavi, M. (2014). Chemical composition, antioxidant and antimicrobial activity of essential oil and extracts of *Tragopogon graminifolius*, a medicinal herb from Iran. Natural Product Communications, 9(1), 121–124.

Indu, M. N., Hatha, A. A. M., Abirosh, C., Harsha, U., & Vivekanandan, G. (2006). Antimicrobial activity of some of the South-Indian spices against serotypes of *Escherichia coli, Salmonella, Listeria monocytogenes* and *Aeromonas hydrophila*. Brazilian Journal of Microbiology, 37, 153-158.

Ivanišová, E., Kačániová, M., Savitskaya, T. A., & Grinshpan, D. D. (2021). Medicinal herbs: Important source of bioactive compounds for food industry. In Herbs and Spices-New Processing Technologies. IntechOpen.

Katz, L., & Baltz, R. H. (2016). Natural product discovery: past, present, and future. Journal of Industrial Microbiology and Biotechnology, 43(2-3), 155-176.

Maleki, M., Ariaii, P., & Sharifi Soltani, M. (2021). Fortifying of probiotic yogurt with free and microencapsulated extract of *Tragopogon Collinus* and its effect on the viability of *Lactobacillus casei* and *Lactobacillus plantarum*. Food Science & Nutrition, 9(7), 3436-3448.

Masumi, S. M. (2001). Kermanshah edible wild plants and the manner of using them. Kermanshah: Kosar Cultural Institute.

Mozafari, M., Nekooeian, A. A., & Janahmadi, Z. (2017). The Antihypertensive Effects of Hydroalcoholic Extract of *Allium Eriophyllum* Leaves on Rats with Simultaneous Type 2 Diabetes and Renal Hypertension. International Cardiovascular Research Journal, 9(1).

Mohammed, F. S., Kına, E., Uysal, İ., Mencik, K., Dogan, M., Pehlivan, M., & Sevindik, M. (2022). Antioxidant and Antimicrobial Activities of Ethanol Extract of *Lepidium spinosum*. Turkish Journal of Agriculture-Food Science and Technology, 10(6), 1116-1119.

Mosavi, H., & Alizadeh, A. (2019). Phytochemical

constituents and antimicrobial activity of *Allium Eriophyllum* Var. Eriophyllum from Iran. Natural product research, 33(21), 3148-3152.

Newman, D. J., & Cragg, G. M. (2016). Natural products as sources of new drugs from 1981 to 2014. Journal of natural products, 79(3), 629-661.

Nisa MU, Huang Y, Benhamed M, Raynaud C. 2019. The plant DNA damage response: signaling pathways leading to growth inhibition and putative role in response to stress conditions. Frontiers in plant science, 10: 653.

Pierce, J., Apisarnthanarak, A., Schellack, N., Cornistein, W., Al Maani, A., Adnan, S., & Stevens, M. P. (2020). Global Antimicrobial Stewardship with a Focus on Lowand Middle-Income Countries: A position statement for the international society for infectious diseases. International Journal of Infectious Diseases, 96, 621-629.

Sasidharan S., Chen Y., Saravanan D., Sundram LYL K.M. Extraction, Isolation, and Characterization of Bioactive Compounds from Plants' Extracts. Tradit Complement Altern Med. 2011;8(1):1–10. pmid:22238476.

Yang, L., Wen, K. S., Ruan, X., Zhao, Y. X., Wei, F., & Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. Molecules, 23(4), 762.

Zangeneh, M. M., & Zangeneh, A. (2020). Biosynthesis of iron nanoparticles using *Allium eriophyllum* Boiss extract: Chemical characterization, antioxidant, cytotoxicity, antibacterial, antifungal, and cutaneous wound healing effects. Applied Organometallic Chemistry, 34(1), e5304.

Zazharskyi VV, Davydenko P, Kulishenko O, Borovik IV, Brygadyrenko VV. 2019. Antimicrobial activity of 50 plant extracts. Biosystems Diversity, 27(2): 163-169.

Zargari, A. (1993). Medicinal plants (4h vol., 5th ed.). Tehran: University of Tehran Press.

Zargari, A. (1996). Medicinal plants (5th vol., 6th ed.). Tehran: University of Tehran Press.

Zhao, H., Su, H., Ahmeda, A., Sun, Y., Li, Z., Zangeneh, M. M., ... & Moradi, R. (2022). Biosynthesis of copper nanoparticles using *Allium eriophyllum* Boiss leaf aqueous extract; characterization and analysis of their antimicrobial and cutaneous wound-healing potentials. Applied Organometallic Chemistry, 36(12), e5587.

Zuridah, H., Fairuz, A. R. M., Zakri, A. H. Z., & Rahim, M. N. A. (2008). In vitro antibacterial activity of Nigella sativa against *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Escherichia coli* and *Bacillus cereus*. Asian Journal of Plant Sciences, 7(3), 331–333.