RESEARCH ARTICLE

Exploring the cytotoxicity of CeO₂ nanoparticles: A compendious approach

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ABSTRACT

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Keywords: Ceria nanoparticles method of synthesis enzyme mimicking activity toxicity biological application Metal oxide nanoparticles due to their antioxidant properties have attracted significant attention and exhibited good potential for use in cancer theranostics. Owing to the poor absorption in the physiological environment, they are an ideal candidate to act as nanocarriers in targeted drug delivery and bioimaging. This feature can be successfully implemented in live monitoring and imaging applications, which offer the possibilities and scope for optical, magnetic resonance, and nuclear imaging. The environment of malignant cells like the rapid proliferation of cells, specific antigen expressions, and leaky tumor vasculature can be used by the modifications in their morphology and surface functionalization. Ceria (CeO₂) nanoparticles have been fascinating in this regard. Different properties such as size, agglomeration behavior, and surface charge density facilitate the interaction of nanoparticles with cancer cells. Compared to other nanoparticles, CeO₂ nanoparticles have a potential for pharmaceutical use since they can act as a therapeutic agent in different disorders such as cancer, inflammation, and neurodegeneration, due to the ability to exhibit variable oxidation state at the nanoparticle surface. Recent literature reports the ecofriendly or 'green' synthesis of CeO₂ nanoparticles in which the biological agent acts as stabilizers for a cost-effective and feasible mode of preparation. In this review, we focus on recent literature on CeO₂ nanoparticles with an emphasis on the methods of fabrication and biomedical applications.

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INTRODUCTION

Nanotechnology has opened the doors of exciting possibilities for biomedical research. The emergence of new classes of nanomaterials has built an excellent platform of the vast opportunities by overcoming the challenges faced by conventional research methods. Owing to the exciting and unique properties of nanomaterials, they are widely employed for applications in medicine, biology, and other life sciences [1-3]. Metal oxide nanoparticles are a family of nanoparticles that gained exciting research attention, especially regarding cancer

treatment [4]. They can be successfully applied when traditional therapies, including surgery, radiotherapy, and chemotherapy, are not efficient for treating cancer.

Cerium (Ce; atomic number =58) is a rare earth element belonging to the lanthanide series of the periodic table. Among other rare elements, it is one of the most abundant oxidizing agent found in oxide form (two oxidation states; +3 and +4). Ce(III) shows an absorption maximum at 230–260 nm, while Ce(IV) shows an absorption maximum at 300–400 nm. This difference arises due to their difference in chemical or electronic environments

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of Ce(III) and Ce(IV) systems. Fig.1S shows the crystal structure of CeO₂.

 CeO_2 nanoparticles have attracted the attention of many scientists due to various applications such as ultraviolet absorption, catalysts, gas sensors, cosmetic products, biomedical applications, including pro-oxidant and antioxidant [5-8].

Considering the unique properties of CeO_2 nanoparticles, researchers have designed and fabricated functionalized CeO_2 nanoparticles with properties tuned for specific applications. There is evidence which shows CeO_2 nanoparticles are toxic to malignant cells and are used to inhibit invasion and to enhance the impact of radiation therapy and chemotherapy to cancer cells [9]. However, CeO_2 nanoparticles are not cytotoxic to healthy tissues and cells and provide conditions to protect the generation of reactive oxygen species (ROS) of various forms [10-12].

PHYSICOCHEMICAL PROPERTIES OF CEO₂ NANOPARTICLES

The variations in chemical or physical properties of the CeO₂ nanoparticles depend on their method of synthesis. The physicochemical properties affect their biological activities, i.e., whether they are inactive, antioxidant, or prooxidant [13]. In nanoscale, both oxidation states, Ce(III) and Ce(IV), are apparent at the surface. The presence of lattice defects created by Ce(III) and their compensation by oxygen reactions on the surface cause a change of CeO_2 to CeO_{2-x} . CeO_2 nanoparticles serve as a remarkable catalyst to mimic the action of natural antioxidant enzymes such as superoxide dismutase (SOD) and catalase. The oxygen defects act as catalytic sites, and by decreasing the size of the CeO₂ nanoparticles, the extent of oxygen deficits decreases [14, 15].

Most oxygen species and nitrogen are self-replicating by the biological system and CeO_2 nanoparticles. The minimum dosage can have catalytic activity for a long time [14]. Therefore, CeO_2 nanoparticles exhibit redox properties better than their bulk counterparts. Furthermore, a mixed valance state helps to scavenge reactive oxygen and nitrogen species and is also useful in chronic oxidative stress and inflammation [16].

When cerium combines with oxygen, it maintains the structure of its fluorite lattice, and this feature is common for both the bulk and the nanoscale. Cerium retains a fluorite lattice by creating a space by losing oxygen on the lattice due to the presence of Ce(III) ions [17-19]. Eight oxygen atoms surround the cerium atoms in CeO2 in the tetrahedral position of the lattice structure. The Ce(III) ions in the nano form cause intrinsic oxygen defects due to charge deficiency [20].

The surface Ce(III)/Ce(IV) ratio of the nanoparticle depends on the method of synthesis [14]. The SOD mimetic activity protects nanoparticles with a high ratio of Ce(III)/Ce(IV) against the diseases related to oxidative stress or inflammation. Besides, CeO₂ nanoparticles with a lower ratio show anticancer and antibacterial effects due to catalase enzyme mimetic activity [21]. Many other inner transition metal ions have been employed to adjust the oxygen concentration and the ratio of Ce(III)/Ce(IV) at the surface of the nanoparticles. The results also showed that the presence of metal with a higher radius (La) causes a higher rate of Ce(III)/Ce(IV) ratio to be compared to smaller metal atoms (Nd) [14].

Under *in vivo* and *in vitro* experimental models, CeO₂ nanoparticles exhibit antioxidant properties due to the self-regeneration of the surface. This is originated from a change in the cerium oxidation state from +3 to +4. Also, the oxygen-free space will alternately change the state of the CeO₂ to Ce₂O₃ [22]. On the other hand, the results indicate that CeO₂ nanoparticles cause cell death [22]. They create a pro-oxidative effect due to ROS to bring about damage to the cell and ultimately lead to apoptosis [22].

A nanosystem should be more hydrophilic and biocompatible in order to obtain good activity in a biological environment. Due to the low solubility of nanoparticles, they are often modified by encapsulating with a hydrophilic and biocompatible polymer material like Dextran. Dextran has the excellent feature like hydrophilicity, biocompatibility, and biodegradability which makes them good candidates for coating, low water soluble nanomaterials for biological applications [22-23].

METHODS OF SYNTHESIS OF CEO₂ NANOPARTICLES

The methods of synthesis play a crucial role in both the physical and chemical properties of nanoparticles. For nanoparticles with biological applications, all of the parameters in the method of preparation should be carefully controlled to maintain their physical and chemical properties in *in vivo* applications[24].

A good number of methods have already been developed for the synthesis of CeO₂ nanoparticles which include: precipitation [25-27] coprecipitation [28], microwave, [29], solvothermal [30], ball milling [31], thermal decomposition [32], spray pyrolysis [33], thermal hydrolysis [34], sol-gel [35-37], sonochemical [38, 39], and hydrothermal [40] methods. Other recent methods include green synthesis by using plant extracts and other natural products such as honey [41-44]. CeO, nanoparticles with outstanding physical and chemical properties can be obtained by an appropriate method of synthesis. Wet-chemical and microemulsion methods, for example, produce small-size CeO, nanoparticles without surface contamination [45]. The methods involving hightemperature operations produce smaller CeO, nanoparticles avoiding agglomeration in spherical morphology [46]. However, conventional methods of preparation have several disadvantages since toxic solvents and reagents need to be used, and they require high temperature and pressure and external additives as stabilizing or capping agents during the reaction [16].

Novel strategies have been developed in which natural products and biomolecules are used as stabilizing agents in the synthesis of CeO₂ nanoparticles. These 'green synthesis' methods gained research interest due to environmental compatibility, safer, reliable, and eco-friendly purpose. Further, this method is an alternative, inexpensive, and simple alternative to the traditional method. The main factors rendering green synthesis as a promising means are plant-, fungus-, polymer-, and nutrient-mediated syntheses [24].

Another interesting green method is used to synthesize nanoparticles using biological molecules such as apoferritin (cage-shaped protein). In this synthesis, apoferritin is considered as a biotemplate. The oxidation of cerium ions is enhanced and eventually causes the formation of CeO_2 nanoparticles [47]. Other than these, several studies are reported using different natural materials and nutrients, such as egg white protein and honey for green synthesis of CeO₂ nanoparticles [48].

The phytosynthesis uses plant extracts as a stabilizing medium for the synthesis of CeO_2 , but it is not suitable for biomedical applications in the present scenario [49]. The methods of producing small nanoparticles of CeO_2 using a fungus-mediation have solved this problem, which results in more stable, well dispersed, and highly

fluorescent nanoparticles. CeO_2 nanoparticles could also be prepared using the culture filtrate of *Curvularia lunata* [42]. It has recently been noted that the use of leaf extract of *Acalypha indica* and *Aloe vera* is used to synthesize CeO_2 nanoparticles [50]. The extract of *Hibiscus sabdariffa* flower also served as a chelating agent for the synthesis of CeO_2 nanoparticles [51]. The extract of these plants has been introduced as a stabilizing and a synthesizing agent in the synthesis of CeO_2 nanoparticles. Kargar et al. reported the green synthesis of CeO_2 nanoparticles stabilized by agarose polymers via the sol-gel method [52].

Factors affecting the synthesis of CeO, nanoparticles

Even though CeO2 nanoparticles contain identical chemical constituents, their biological effects vary depending on the synthetic method and factors employed during the fabrication step. The factors of synthesis in both traditional and green synthesis methods dictate the physical parameters of CeO₂ nanoparticles, including particle size, shape, dispersion level, and surface charge. Residual surfactant contamination, nature of stabilizing agent, and the Ce(III)/Ce(IV) surface ratio can influence the interactions at CeO₂ nanoparticles with the biological environment [53]. Efficient manipulation of the ratio of Ce(III)/Ce(IV) and the free space of the oxygen causes the change in biological interactions.

The counter anions in cerium salts greatly influence the size and morphology of particles. For example, anions such as SO42, or halides (Cl, Br, and I⁻) form nanorods of CeO₂, while NO₃⁻ anion forms the nanocube structure [54]. The dependence of the morphology of CeO, nanoparticles on temperature is not straightforward. CeO, nanoparticles with different morphology can be fabricated with a single reaction at the different temperatures as well as a combination of various synthetic methods. For instance, in the hydrothermal method, at room temperature, nanorods were synthesized and converted to nanotubes at 100 °C. Similarly, CeO, nanoparticles could be prepared as nanowires at a temperature of 110 °C, nanospheres at 120 °C, nanospheres and nanocubes at 140 °C, and nanocubes at both 160 and 180 °C [55].

Another important factor in the synthesis of CeO₂nanoparticles is the amount of dispersion. This determines the extent of agglomeration in aqueous solutions and other biological environments. The amount of dispersion enhances stability, along with



Fig. 1: Major biological applications of CeO, nanoparticles

the reduction of unnecessary interactions with other cells or proteins present in the vicinity. It also increases the survival time in the circulation, reduces the toxicity of nanoparticles, and provides an effective dose to minimize side effects. The dispersion rate is effectively improved by coating CeO_2 nanoparticles with biocompatible polymers, composite materials, surfactants, stabilizers, or biomacromolecules during synthesis or even after synthesis [14].

Many traditional methods are non-ecofriendly and not feasible. Generally, biocompatible coatings increase sustainability, longer retention times, and reduce the toxicity of CeO₂ NP. These coatings include polyacrylic acid [56], polyethylene glycol (PEG) [57], polyethylene mine [58], cyclodextrin [59], glucose [60], and folic acid [61]. Literature suggests that the coatings can increase the stability, influence mechanism of uptake, and intracellular localization of CeO₂ nanoparticles in different cell compartments and cytotoxicity in biological applications [62].

BIOLOGICAL APPLICATIONS OF CEO₂ NANOPARTICLES

CeO₂ nanoparticles have many uses in the biological fields, but there are many concerns about their consequences on health and the environment. The impact of CeO₂ nanoparticles on human health has attracted scientists and researchers. The inhalation of CeO₂ nanoparticles will affect the lungs and lymph nodes. When it enters the circulatory system, it may be distributed in other organs like the liver, kidney, and spleen. Eventually, the excretion of cerium in the feces takes place [22]. For this, it is essential to study their interaction

with physiological activity and toxicity in the human body. The main areas of biological activities reported for CeO_2 nanoparticles are represented in Fig.1.

Antibacterial activity

 CeO_2 nanoparticles have antibacterial activity against *P. pseudomonas aeruginosa*. Research confirms that by increasing the concentration of CeO_2 nanoparticles, the growth of *P. aeruginosa* (NCIM-2242) is inhibited. Hydrothermally synthesized CeO2-ZnO nanocomposite were studied for activity against Streptococcus mutans. The authors observed that minimum inhibitory concentration (MIC) against the microorganisms was 0.22 mg / mL[63].

Genotoxicity

The literature shows the investigation of the mechanism of molecular toxicity of CeO, nanoparticles on lung adenocarcinoma cells (A549). These nanoparticles have changed the morphology of cancer cells A549. Due to the presence of CeO₂ nanoparticles and ROS, damage to the DNA and cell cycle stopped causing the death of the A549 cell [64]. Another genotoxicity study on female albino Wistar rats was performed by comet and chromosomal aberration and micronucleus tests. The final results indicated that in high doses of CeO₂, nanoparticles would damage DNA in the liver and peripheral blood leukocytes (PBLs)[43]. Cytotoxic and genotoxic study of CeO, nanoparticles in a human neuroblastoma cell line (IMR32) were also reported. CeO, nanoparticles caused cytotoxicity, which was confirmed by lactate dehydrogenase assays and 3-[4,5-dimethylthiazol2-yl-2, 5-diphenyl tetrazolium bromide. The results showed that ROS was involved in the toxicity of CeO_2 nanoparticles [65].

Neurotoxicity

Because of the presence of the blood-brain barrier (BBB), the targeted drug delivery is the hardest task in neural drugs since the BBB acts as a selective filter for these molecules. Investigation of the responsive action of the brain was performed that instructs the administration of suramin into the intracerebral region. The results showed that CeO_2 nanoparticles labeled as fluorescence deposited into the liver and spleen when they entered the body of the mice [22].

Antioxidant activity

Anti-oxidant properties appear when CeO_2 nanoparticles are combined with levan. Levan acted as a reducing and stabilizing agent. Levan combined with nanoparticles were favorable towards the disease related to ROS [66]. Recent work has shown that CeO_2 nanoparticles with a size of 20 nm increase longevity and maintains function expression in brain cell cultures. In a size of 10 nm, cell death is reduced with CeO_2 nanoparticles and UV and H_2O_2 . Therefore, the results indicate that the antioxidant activity of the CeO_2 nanoparticles is dependent on the size [67].

In vitro studies

A large number of *in vitro* studies were available, which reports the cytotoxic action of CeO₂ nanoparticles in various cancer cell lines. A study found that CeO₂ nanoparticles are toxic to lung cancer cell lines. Sulforhodamine B was used to examine the cell viability at different concentrations and times. The significant cellular loss was observed with regard to the dose of nanoparticles and the incubation time with CeO₂ nanoparticles. Cytotoxicity assay of CeO₂ nanoparticles was carried on human fibrosarcoma (HT- 1080) cells and breast cancer (MCF-7). There were no cell deaths in the concentration range of 20-200 µg mL⁻¹ when CeO₂ nanoparticles were incubated with cancer cells [68].

There has been an investigation on the protective effects of CeO_2 nanoparticles on primary human skin fibroblasts [69]. The CeO_2 nanoparticles were found to be internalized and showed strong ROS scavenging activity, and the viability of the fibroblasts was not affected. CeO₂

nanoparticles affect the mitochondrial activity and lead to more productive ATP. Sack et al. reviewed cancer treatment using the redox activity of CeO, nanoparticles in combination with doxorubicin. A comparison of the antitumor activity of CeO₂ nanoparticles with doxorubicin shows that doxorubicin has antitumor activity and ROS formation and oxidative damage against human melanoma cells (A375 cells) are noticeable. The use of the combination of doxorubicin and nanostera exhibits better anti-tumor activity than when it is used alone [70]. In another study, Sack-Zschauer et al. have shown that CeO, nanoparticles have destroyed the cancer cells of the glioma and protected healthy cells. CeO, nanoparticles have cytotoxicity activity on astrocytoma (grade III glioma), while on microvascular endothelial cells, no influence could be marked [71]. One of the major features of CeO2 nanoparticles is the activity in cancer cells. Enough evidences were available in literature showing the potent activity of CeO2 in various types of malignant cells. For example, the viability of lung cancer cells got decreased with increasing the concentration and exposure time of CeO2 NPs. The mechanism was reported to be oxidative induced free radical formation[110]. Lack of toxicity in normal cell lines(L929) with significant toxicity against prostate cancer cells (PC-3) were also an interesting result confirmed using MTT assay[111]. Apart from direct activity against cancer cells, CeO2 NPs have found to enhance the effectiveness of cancer treatment strategies. A good example of this was reported in pancreatic cell lines in which the authors confirmed that CeO2 NPs can act as sensitizer in radiation treatment associated with pancreatic cancer which increases the therapeutic outcome [112]. The role of CeO2 NPs have been thoroughly reviewed in literature which further discuss the mode of action and possibilities for cancer therapeutics. These studies indicate that CeO2 NPs have the ability to selectively attack malignant cells through increasing the sensitivity to other treatments and also inducing toxicity with differentiating them from healthy cells [113]. This differential cytotoxcity of CeO2 in malignant cells gives future possibilities to use as a nanotherapeutic agent in cancer treatment [114]. Similar results were obtained by coupling conventional Chemotherapeutic drugs such as Doxorubicin (DOX) with CeO2 NPs for enhanced synergistic effect [115]. Recently the efficiency of CeO2 as an agent in near-infrared light controlled Photo dynamic therapy (PDT) against hypoxia cancer is also reported which gives further scope in this area[116].

In vivo studies

Literature shows that CeO₂ nanoparticles can affect *in vivo* models such as the rat, mice, and non-rodent models in different ways. There are a good number of reports on the liver, lungs, spleen, kidneys, and brain to comprehend the activity of CeO₂ nanoparticles [72-74]. Also, there have been studies on plant crops like rice [75], wheat, sunflower, pumpkin [76], tomato [77], kidney bean [78], radish [79], cucumber [80], and Rubia cordifolia [81] to determine the absorption rate of CeO₂ nanoparticles. The absorption rate in the roots is higher than in other parts of the plant. This is due to various factors, such as the size of nanoparticle [80], the extent of agglomeration [76, 80], and concentration [77, 78].

Caenorhabditis elegans have been exposed to various levels of nanoscale coated surfaces in an *in* vivo model. The different surfaces coated with CeO, nanoparticles showed a difference in absorption [82]. Kwon et al. showed that the introduction of CeO₂ nanoparticles within mitochondria is a successful treatment for neurodegenerative diseases [83]. The synthesis of triphenylphosphoniumconjugated CeO, nanoparticles has also been reported, which is located in the mitochondria of different cells. This can be used to treat diseases associated with mitochondria, Alzheimer's, and other neurodegenerative diseases. Hijaz et al. reported that combining folic acid-CaPs with cisplatin results in the death of ovarian cancer xenograft nude cells [84]. Also, folic acid- CeO, nanoparticles reduce vimentin, which indicates the break of the metastasis of ovarian cancer. Reports also demonstrate the cellular mechanism and the catalytic activity of CeO, nanoparticles to prevent the reduction of photoreceptor cells. The results showed that CeO, nanoparticles led to a reduction in apoptotic cells and lipid peroxidation in the retina [85].

Physiological studies

The co-precipitation method was used for the synthesis of CeO_2 nanoparticles, and the biological effects of synthesized particles were studied using the intraperitoneal technique, where nanoparticles spread throughout all tissues. The results showed that the antioxidant effects of CeO₂ nanoparticles depend strongly on the dosage [86]. The antioxidant effects of CeO_2 nanoparticles were also reported wherein it was demonstrated that CeO_2 nanoparticles help reducing weight gain and lower the plasma levels of leptin, insulin, glucose, and triglycerides [87].

 CeO_2 nanoparticles were used for prophylactic treatment of hepatic ischemia reperfusion injury in Sprague Dawley rats. The CeO_2 nanoparticles caused a decrease in hepatocyte necrosis, alanine aminotransaminase, macrophage-derived chemokine, lactate dehydrogenase, myoglobin, macrophage inflammatory protein-2, keratinocyte chemoattractant (KC)/human growth-regulated oncogene (GRO), and plasminogen activator inhibitor-1 [88].

Enzyme Mimetic Activities

Superoxide Dismutase and Catalase Mimetic Activity

In mammalian cells, natural aerobic metabolism produces free radicals, called superoxide radicals. They are produced due to. The radicals of this kind serve as signaling molecules and have been an important factor in the oxidative response to pathogens. The extra amount of these radicals is destroyed by enzymes SOD [89]. A ratio higher than Ce(III)/Ce(IV) in CeO₂ nanoparticles shows more SOD-mimetic activity than lower values. Catalase causes H_2O_2 to be lost, and an oxidizing agent is harmful [90]. Low extent of Ce(III)/ Ce(IV) ratio shows catalase-mimetic activity more compared to higher ratio of Ce(III)/Ce(IV) [91].

Phosphatase Mimetic Activity

Phosphatases are the kind of enzymes used for the hydrolysis of esters into phosphate ions to remove phosphate groups from their substrates [92]. The relatively low Ce(III)/Ce(IV) ratio in CeO₂ nanoparticles can mimic phosphatase for artificial phosphate substrates [93] and bio-relevant substrates, such as ATP [94].

Radical/ROS Scavenging Activity

Due to the redox properties, rare metal ions such as cerium ion can serve as oxygen free radical scavenger. ROS comprises free radicals and non-radicals and is more reactive than molecular oxygen. Free radicals are less stable and more reactive and can have a significant effect on the biochemistry and metabolism of living environments. ROS are produced as an ancillary

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		-	
Method of Preparation	The diameter of particles (nm)	Morphology	Reference
Precipitation	15	Spherical	28
Hydrothermal	5	Octahedral	40
Solvothermal	8	Polyhedral	30
Spray Pyrolysis	17	Cubic	33
Plant- mediated	36	Spherical	115
Fungus- mediated	5	Spherical	50
Polymer- mediated	2	Spherical	116
Nutrient-mediated	25	Spherical	48

Table 1. Summary of major synthesis methods and features of CeO₂ nanoparticles

product in aerobic activity and are associated with oxidative stress. Increasing levels in the body cause many diseases. However, ROS plays the role of signaling molecules in physiological processes. Given these points, antioxidants are molecules that can destroy or inhibit the production of ROS. Excess ROS levels create oxidative stress, depletion of cellular antioxidants, and, thus, oxidation of the biomolecules like proteins or lipids of cellular membranes, carbohydrates, DNA, and ultimately causing cell damage and cell death.

The smaller CeO₂ nanoparticles achieve more cellular absorption due to volumes larger than their surface and faster kinetics. Kumari et al. reported that CeO₂ nanoparticles with a size of 25 nm to the neuroblastoma cells were more toxic than particles with a size of 3 μ m. [65].

Synthesis of various forms of nanoparticles, such as spheres, pillars, rods, cubes, helices, wires, polygonal, octahedral, has been reported.[95-98]. Due to the type of nanoparticle structure, they have different properties of different chemical, electrical, optical, and magnetic properties. Therefore, they can interact with different cells and biological molecules. For example, nanoparticles in polygonal, cube, or rod morphology have sharp edges and can cause mechanical damage to cells [99]. The smaller nanoparticles have a larger surface-to-large ratio for interactions with cells. The smaller nanoparticles have a higher Ce(III) content, which eliminates intracellular and extracellular ROS [100].

Studies have shown that the toxicity of CeO_2 nanoparticles to human hepatoma cells of SMMC⁻ 7721 is not related to the size and morphology of cerium oxide nanoparticles [101]. This can be due to aggregation, which reduces their toxicity. Therefore, the mechanism of interaction does not only depend on CeO_2 nanoparticles but also depends on the type of cell and the culture medium.

The hydroxyl radical is one of the most

biologically active free radical [102]. CeO_2 nanoparticles are capable of removing hydroxyl radicals at certain sizes. CeO_2 nanoparticles show a neuroprotective effect on adult rat spinal cord neurons.

INFLUENCE OF MORPHOLOGY ON THE ACTIVITY CEO, NANOPARTICLES

Various methods of preparation have influences on the features of nanoparticles formed, such as morphology, size, etc. as shown in Table 1. Also, surface charge density on the nanoparticle plays a crucial role in many applications such as *in vitro* and *in vivo* processes.

The Influence of morphology on the cytotoxicity of CeO2 nanoparticles has been investigated and reported in literature. A good example of this exploration was carried out by fabricating CeO2 nanoparticles having a different morphology such as rod shaped and cubic nanoparticles. Toxicity was evaluated using important parameters such as LDH release, ROS production and TNF-a production in Macrophages from RAW264.7 cell line [117]. The authors were able to reach a conclusion that cytotoxicity is significantly enhanced in a dosedependent manner by rod-like nanoparticles. But a contrary result was also reported in which rod shaped CeO2 nanoparticles were found less cytotoxic compared to Cubic shaped nanoparticles in HepG2 cells [118]. These results indicate the morphological influence may depend significantly on the nature of cell line exposed to CeO2 nanoparticles and needs further exploration to reveal more information about toxicity pathways.

The surface charge of CeO_2 nanoparticles is essential in cell targeting, cell adhesion, uptake, subcellular distribution, and cytotoxicity. The surface charge can be modified by using appropriate acid, base buffers, or coatings with polymer, biomolecule, ligand, or surfactant and stabilizer. The uptake of nanoparticles by cells is, in general, associated with two steps: a binding step on the cell membrane and an ultimate internalization step. The first stage is influenced by the surface charge of cell membranes and CeO₂ nanoparticles. The higher the surface charge of the CeO₂ nanoparticles, the stronger is the bond through the electrostatic interaction with the membrane of the cell [103]. In general, the cell surface has a negative charge, so nanoparticles interact with a positive charge by electrostatic interaction. It has also been reported that negative CeO₂ nanoparticles can be located at specific cationic locations at the cell surface.

Research shows that Ce(III) is related to the toxicity of CeO₂ nanoparticles [104]. The higher Ce(III) levels show toxicity effects and *vice versa* in the proposed animal model. Further CeO₂ nanoparticles with higher Ce(IV) on their surfacedisplayed catalase mimetic activity. This destroys H_2O_2 to molecular oxygen and protects the cells against the toxic ROS. CeO₂ nanoparticles with higher Ce(III) on their surface shows the efficacy to remove radicals of superoxide and produce H_2O_2 , which is toxic to the cells.

CeO₂ nanoparticles act as direct antioxidants and act as radical scavenging agents as their own since the interaction of superoxide radical, hydroxyl radical, and hydrogen peroxide, and ultimately oxidative stress causes cell death. The level of a quantitative assessment of total ROS, malondialdehyde, α -tocopherol, glutathione, and lactate dehydrogenase shows a reduction in the level of glutathione and α - tocopherol indicating cytotoxicity. Free radicals are generated due to interaction with nanoparticles and enhancement in oxidative stress, yielding a high level of lactate dehydrogenase and malondialdehyde, which causes cell membrane damage and lipid peroxidation.

Ovalbumin and lysozyme are two proteins in the egg which can be effectively used as stabilizing agent in synthesis CeO_2 nanoparticles. The mechanism involves electrostatic interactions between cerium ions, and opposite- charged protein leads to controlled and stable growth of CeO₂ nanoparticles [105].

The activity of CeO_2 nanoparticles in a microbial environment has shown that nanoparticles cannot penetrate the cells of bacteria and algae. For this reason, the toxic effects of CeO_2 nanoparticles have been suggested to be caused by their direct connection to the cell wall of algae and bacteria[106]. CeO₂ nanoparticles cause a change in the nutrient transport functions of the membrane bacterial cells and algae, which results in mechanical damage and membrane disruption, or generation of ROS and oxidative stress [107].

Sadhu et al. observed that high concentrations of CeO, nanoparticles in BY-2 cells cause cellular toxicity and can affect metabolic activity [108]. Ce(III) and ROS concentration-dependent accumulation was observed for all CeO, NPs. Significant DNA damage and changes in the antioxidant defense system were observed for concentration 50g/mL and 250g/mL. Another study reports the synthesis of CeO, nanoparticles and a comparison of the toxicity of CeO, powder on nano and a bulk scale on cancer cells and normal healthy cells. MTT assay test was performed for SKBR3 (human breast cancer cell line), A431 (Human epidermis carcinoma cell line), and C2Cl2 (ATCC mouse skeletal muscle cell line). The toxicity of CeO₂ nanoparticles is significantly higher than the bulk form, and their toxicity is higher on cancer cells. This issue is further observed at higher concentrations [109].

CONCLUSION

Metal oxide nanoparticles are hotspots of nanotechnology research. They are relevant and received attention in almost all thrust areas of research like drug delivery, bioimaging, and microbiology, etc. Compounds of inner transition metals are a unique family in nanomaterials due to their unique chemical properties like spectral behavior, magnetic property, etc. In this review, we have discussed CeO, nanoparticle by focusing on their various biological applications and synthetic methods. Since cytotoxicity remains as a challenge of application of nanomaterials, special attention has been given to explain cytotoxicity of CeO, nanoparticles. This may pave new possibilities for applications of CeO₂ nanoparticles by summarizing the major outcome of research and various aspects of toxicity.

SCOPE AND FUTURE PERSPECTIVES

CeO₂ nanoparticles stand out the unique class of candidates due to their chemical properties and lack of significant cytotoxicity. Over the past years, new methods for controlling dispersion, reducing the accumulation and reduction of protein interactions, including coatings with compatible biocompatible and biodegradable compounds, are designed. Green synthesis methods are promising for the production of CeO₂ nanoparticles and their biological applications among the various methods reported for the synthesis of CeO₂ nanoparticles. Also, studies have to be conducted to determine the influences of cell types and cellular features on the cellular toxicity of CeO, nanoparticles. All these developments can be effectively applied to new real-life applications that are yet to be explored. Even though the possibilities of these materials are still not explored, their cytotoxicity should be carefully analyzed in different media and systems of administration. Also, features like vitro conditions, medium pH, protein, and salt concentration should be taken into consideration. The characteristic CeO₂ nanoparticles should also be performed at each stage of their application periodically and in vitro / in vivo tests to better explain the changes in toxicity.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interests regarding the publication of this manuscript.

REFERENCES

- Loh KP, Ho D, Chiu GNC, Leong DT, Pastorin G, Chow EK-H. Clinical Applications of Carbon Nanomaterials in Diagnostics and Therapy. Advanced Materials. 2018;30(47):1802368.
- Sivasankarapillai VS, Jose J, Shanavas MS, Marathakam A, Uddin MS, Mathew B. Silicon Quantum Dots: Promising Theranostic Probes for the Future. Current Drug Targets. 2019;20(12):1255-63.
- Yao J, Wang H, Chen M, Yang M. Recent advances in graphenebased nanomaterials: properties, toxicity and applications in chemistry, biology and medicine. Microchimica Acta. 2019;186(6).
- 4. Wason M, Lu H, Yu L, Lahiri S, Mukherjee D, Shen C, et al. Cerium Oxide Nanoparticles Sensitize Pancreatic Cancer to Radiation Therapy through Oxidative Activation of the JNK Apoptotic Pathway. Cancers. 2018;10(9):303.
- Li C, Sun Y, Hess F, Djerdj I, Sann J, Voepel P, et al. Catalytic HCl oxidation reaction: Stabilizing effect of Zr-doping on CeO2 nano-rods. Applied Catalysis B: Environmental. 2018;239:628-35.
- Parwaiz S, Khan MM, Pradhan D. CeO2-based nanocomposites: An advanced alternative to TiO2 and ZnO in sunscreens. Materials Express. 2019;9(3):185-202.
- Motaung DE, Mhlongo GH, Makgwane PR, Dhonge BP, Cummings FR, Swart HC, et al. Ultra-high sensitive and selective H2 gas sensor manifested by interface of n-n heterostructure of CeO2-SnO2 nanoparticles. Sensors and

Actuators B: Chemical. 2018;254:984-95.

- Khorrami MB, Sadeghnia HR, Pasdar A, Ghayour-Mobarhan M, Riahi-Zanjani B, Hashemzadeh A, et al. Antioxidant and toxicity studies of biosynthesized cerium oxide nanoparticles in rats. International Journal of Nanomedicine. 2019;Volume 14:2915-26.
- Li C, Zhao W, Liu B, Xu G, Liu L, Lv H, et al. Cytotoxicity of Ultrafine Monodispersed Nanoceria on Human Gastric Cancer Cells. Journal of Biomedical Nanotechnology. 2014;10(7):1231-41.
- Renu G, Rani VVD, Nair SV, Subramanian KRV, Lakshmanan V-K. Development of Cerium Oxide Nanoparticles and Its Cytotoxicity in Prostate Cancer Cells. Advanced Science Letters. 2012;6(1):17-25.
- Gao Y, Gao F, Chen K, Ma J-l. Cerium oxide nanoparticles in cancer. OncoTargets and Therapy. 2014:835.
- 12. Abbas F, Jan T, Iqbal J, Haider Naqvi MS, Ahmad I. Inhibition of Neuroblastoma cancer cells viability by ferromagnetic Mn doped CeO2 monodisperse nanoparticles mediated through reactive oxygen species. Materials Chemistry and Physics. 2016;173:146-51.
- Mortazavi Milani Z, Charbgoo F, Darroudi M. Impact of physicochemical properties of cerium oxide nanoparticles on their toxicity effects. Ceramics International. 2017;43(17):14572-81.
- Chen B-H, Stephen Inbaraj B. Various physicochemical and surface properties controlling the bioactivity of cerium oxide nanoparticles. Critical Reviews in Biotechnology. 2018;38(7):1003-24.
- Das S, Dowding JM, Klump KE, McGinnis JF, Self W, Seal S. Cerium oxide nanoparticles: applications and prospects in nanomedicine. Nanomedicine. 2013;8(9):1483-508.
- Charbgoo F, Ahmad M, Darroudi M. Cerium oxide nanoparticles: green synthesis and biological applications. International Journal of Nanomedicine. 2017;Volume 12:1401-13.
- F. Esch, S. Fabris, L. Zhou, T. Montini, C. Africh, P. Fornasiero, G. Comelli, R. Rosei, Electron localization determines defect formation on ceria substrates, Science, 309 (2005) 752-755.
- Tsai Y-Y, Oca-Cossio J, Lin S-M, Woan K, Yu P-C, Sigmund W. Reactive oxygen species scavenging properties of ZrO2–CeO2solid solution nanoparticles. Nanomedicine. 2008;3(5):637-45.
- Fronzi M, Soon A, Delley B, Traversa E, Stampfl C. Stability and morphology of cerium oxide surfaces in an oxidizing environment: A first-principles investigation. The Journal of Chemical Physics. 2009;131(10):104701.
- Suzuki T, Kosacki I, Anderson HU, Colomban P. Electrical Conductivity and Lattice Defects in Nanocrystalline Cerium Oxide Thin Films. Journal of the American Ceramic Society. 2004;84(9):2007-14.
- Gupta A, Das S, Neal CJ, Seal S. Controlling the surface chemistry of cerium oxide nanoparticles for biological applications. Journal of Materials Chemistry B. 2016;4(19):3195-202.
- Rajeshkumar S, Naik P. Synthesis and biomedical applications of Cerium oxide nanoparticles – A Review. Biotechnology Reports. 2018;17:1-5.
- Perez JM, Asati A, Nath S, Kaittanis C. Synthesis of Biocompatible Dextran-Coated Nanoceria with pH-Dependent Antioxidant Properties. Small. 2008;4(5):552-6.
- 24. Dhall A, Self W. Cerium Oxide Nanoparticles: A Brief Review

of Their Synthesis Methods and Biomedical Applications. Antioxidants. 2018;7(8):97.

- Y. Liu, J. Zuo, X. Ren, L. Yong, Synthesis, and character of cerium oxide (CeO₂) nanoparticles by the precipitation method, Metalurgija, 53 (2014) 463-465.
- Zhang QL, Yang ZM, Ding BJ. Synthesis of Cerium Oxide Nanoparticles by the Precipitation Method. Materials Science Forum. 2009;610-613:233-8.
- 27. P. Kavitha, Synthesis and characterization of nanoceria by using rapid precipitation method, PARIPEX-Indian J. Res. , 4 (2016).
- 28. D.A. Pelletier, A.K. Suresh, G.A. Holton, C.K. McKeown, W. Wang, B. Gu, N.P. Mortensen, D.P. Allison, D.C. Joy, M.R. Allison, Effects of engineered cerium oxide nanoparticles on bacterial growth and viability, Appl. Environ. Microbiol., 76 (2010) 7981-7989.
- 29. Shirke BS, Patil AA, Hankare PP, Garadkar KM. Synthesis of cerium oxide nanoparticles by microwave technique using propylene glycol as a stabilizing agent. Journal of Materials Science: Materials in Electronics. 2010;22(2):200-3.
- 30. Zhang H, He X, Zhang Z, Zhang P, Li Y, Ma Y, et al. Nano-CeO2Exhibits Adverse Effects at Environmental Relevant Concentrations. Environmental Science & Technology. 2011;45(8):3725-30.
- Yadav TP, Srivastava ON. Synthesis of nanocrystalline cerium oxide by high energy ball milling. Ceramics International. 2012;38(7):5783-9.
- Wang Y, Mori T, Li J-G, Ikegami T. Low-Temperature Synthesis of Praseodymium-Doped Ceria Nanopowders. Journal of the American Ceramic Society. 2004;85(12):3105-7.
- 33. Demokritou P, Gass S, Pyrgiotakis G, Cohen JM, Goldsmith W, McKinney W, et al. Anin vivoandin vitrotoxicological characterisation of realistic nanoscale CeO2inhalation exposures. Nanotoxicology. 2012;7(8):1338-50.
- 34. Hirano M, Fukuda Y, Iwata H, Hotta Y, Inagaki M. Preparation and Spherical Agglomeration of Crystalline Cerium(IV) Oxide Nanoparticles by Thermal Hydrolysis. Journal of the American Ceramic Society. 2004;83(5):1287-9.
- 35. He H-W, Wu X-Q, Ren W, Shi P, Yao X, Song Z-T. Synthesis of crystalline cerium dioxide hydrosol by a sol–gel method. Ceramics International. 2012;38:S501-S4.
- 36. Darroudi M, Sarani M, Kazemi Oskuee R, Khorsand Zak A, Hosseini HA, Gholami L. Green synthesis and evaluation of metabolic activity of starch mediated nanoceria. Ceramics International. 2014;40(1):2041-5.
- Darroudi M, Sarani M, Kazemi Oskuee R, Khorsand Zak A, Amiri MS. Nanoceria: Gum mediated synthesis and in vitro viability assay. Ceramics International. 2014;40(2):2863-8.
- Pinjari DV, Pandit AB. Room temperature synthesis of crystalline CeO2 nanopowder: Advantage of sonochemical method over conventional method. Ultrasonics Sonochemistry. 2011;18(5):1118-23.
- 39. Yin L, Wang Y, Pang G, Koltypin Y, Gedanken A. Sonochemical Synthesis of Cerium Oxide Nanoparticles— Effect of Additives and Quantum Size Effect. Journal of Colloid and Interface Science. 2002;246(1):78-84.
- Rojas S, Gispert JD, Abad S, Buaki-Sogo M, Victor VM, Garcia H, et al. In Vivo Biodistribution of Amino-Functionalized Ceria Nanoparticles in Rats Using Positron Emission Tomography. Molecular Pharmaceutics. 2012;9(12):3543-50.

- 41. Arumugam A, Karthikeyan C, Haja Hameed AS, Gopinath K, Gowri S, Karthika V. Synthesis of cerium oxide nanoparticles using Gloriosa superba L. leaf extract and their structural, optical and antibacterial properties. Materials Science and Engineering: C. 2015;49:408-15.
- 42. S. Munusamy, K. Bhakyaraj, L. Vijayalakshmi, A. Stephen, V. Narayanan, Synthesis and characterization of cerium oxide nanoparticles using Curvularia lunata and their antibacterial properties, Int J Innovative Res Sci Eng, 2 (2014) 318-323.
- 43. Kumari M, Kumari SI, Kamal SSK, Grover P. Genotoxicity assessment of cerium oxide nanoparticles in female Wistar rats after acute oral exposure. Mutation Research/Genetic Toxicology and Environmental Mutagenesis. 2014;775-776:7-19.
- 44. M. Aliahmad, A. Rahdar, F. Sadeghfar, S. Bagheri, M.R. Hajinezhad, Synthesis and Biochemical effects of magnetite nanoparticle by surfactant-free electrochemical method in an aqueous system: The current density effect, Nanomedicine Research Journal, 1 (2016) 39-46.
- 45. Das S, Singh S, Dowding JM, Oommen S, Kumar A, Sayle TXT, et al. The induction of angiogenesis by cerium oxide nanoparticles through the modulation of oxygen in intracellular environments. Biomaterials. 2012;33(31):7746-55.
- 46. Hardas SS, Butterfield DA, Sultana R, Tseng MT, Dan M, Florence RL, et al. Brain Distribution and Toxicological Evaluation of a Systemically Delivered Engineered Nanoscale Ceria. Toxicological Sciences. 2010;116(2):562-76.
- 47. Liu X, Wei W, Yuan Q, Zhang X, Li N, Du Y, et al. Apoferritin– CeO2nano-truffle that has excellent artificial redox enzyme activity. Chem Commun. 2012;48(26):3155-7.
- Kargar H, Ghazavi H, Darroudi M. Size-controlled and bio-directed synthesis of ceria nanopowders and their in vitro cytotoxicity effects. Ceramics International. 2015;41(3):4123-8.
- 49. Gagnon J, Fromm KM. Toxicity and Protective Effects of Cerium Oxide Nanoparticles (Nanoceria) Depending on Their Preparation Method, Particle Size, Cell Type, and Exposure Route. European Journal of Inorganic Chemistry. 2015;2015(27):4510-7.
- 50. G.S. Priya, A. Kanneganti, K.A. Kumar, K.V. Rao, S. Bykkam, Biosynthesis of Cerium oxide nanoparticles using Aloe barbadensis miller gel, Int J Sci Res Publ, 4 (2014) 199-224.
- Thovhogi N, Diallo A, Gurib-Fakim A, Maaza M. Nanoparticles green synthesis by Hibiscus Sabdariffa flower extract: Main physical properties. Journal of Alloys and Compounds. 2015;647:392-6.
- Kargar H, Ghasemi F, Darroudi M. Bioorganic polymerbased synthesis of cerium oxide nanoparticles and their cell viability assays. Ceramics International. 2015;41(1):1589-94.
- 53. Alili L, Sack M, von Montfort C, Giri S, Das S, Carroll KS, et al. Downregulation of Tumor Growth and Invasion by Redox-Active Nanoparticles. Antioxidants & Redox Signaling. 2013;19(8):765-78.
- 54. Wu Q, Zhang F, Xiao P, Tao H, Wang X, Hu Z, et al. Great Influence of Anions for Controllable Synthesis of CeO2 Nanostructures: From Nanorods to Nanocubes. The Journal of Physical Chemistry C. 2008;112(44):17076-80.
- 55. Pan C, Zhang D, Shi L, Fang J. Template-Free Synthesis, Controlled Conversion, and CO Oxidation Properties

of CeO2 Nanorods, Nanotubes, Nanowires, and Nanocubes. European Journal of Inorganic Chemistry. 2008;2008(15):2429-36.

- 56. Sehgal A, Lalatonne Y, Berret JF, Morvan M. Precipitation– Redispersion of Cerium Oxide Nanoparticles with Poly(acrylic acid): Toward Stable Dispersions. Langmuir. 2005;21(20):9359-64.
- 57. Karakoti AS, Singh S, Kumar A, Malinska M, Kuchibhatla SVNT, Wozniak K, et al. PEGylated Nanoceria as Radical Scavenger with Tunable Redox Chemistry. Journal of the American Chemical Society. 2009;131(40):14144-5.
- 58. Lee SS, Song W, Cho M, Puppala HL, Nguyen P, Zhu H, et al. Antioxidant Properties of Cerium Oxide Nanocrystals as a Function of Nanocrystal Diameter and Surface Coating. ACS Nano. 2013;7(11):9693-703.
- 59. Xu C, Lin Y, Wang J, Wu L, Wei W, Ren J, et al. Nanoceria-Triggered Synergetic Drug Release Based on CeO2-Capped Mesoporous Silica Host-Guest Interactions and Switchable Enzymatic Activity and Cellular Effects of CeO2. Advanced Healthcare Materials. 2013;2(12):1591-9.
- 60. Li M, Shi P, Xu C, Ren J, Qu X. Cerium oxide caged metal chelator: anti-aggregation and anti-oxidation integrated H2O2-responsive controlled drug release for potential Alzheimer's disease treatment. Chemical Science. 2013;4(6):2536.
- Deshpande S, Patil S, Kuchibhatla SVNT, Seal S. Size dependency variation in lattice parameter and valency states in nanocrystalline cerium oxide. Applied Physics Letters. 2005;87(13):133113.
- Vincent A, Babu S, Heckert E, Dowding J, Hirst SM, Inerbaev TM, et al. Protonated Nanoparticle Surface Governing Ligand Tethering and Cellular Targeting. ACS Nano. 2009;3(5):1203-11.
- 63. dos Santos C, Passos Farias I, Reis Albuquerque A, e Silva P, Costa One G, Sampaio F. Antimicrobial activity of nano cerium oxide (IV) (CeO2) against Streptococcus mutans. BMC Proceedings. 2014;8(Suppl 4):P48.
- 64. Mittal S, Pandey AK. Cerium Oxide Nanoparticles Induced Toxicity in Human Lung Cells: Role of ROS Mediated DNA Damage and Apoptosis. BioMed Research International. 2014;2014:1-14.
- 65. Kumari M, Singh SP, Chinde S, Rahman MF, Mahboob M, Grover P. Toxicity Study of Cerium Oxide Nanoparticles in Human Neuroblastoma Cells. International Journal of Toxicology. 2014;33(2):86-97.
- Kim S-J, Chung BH. Antioxidant activity of levan coated cerium oxide nanoparticles. Carbohydrate Polymers. 2016;150:400-7.
- 67. N. Singh, E. Amateis, J.E. Mahaney, K. Meehan, B.A. Rzigalinski, The antioxidant activity of cerium oxide nanoparticles is size-dependent and blocks Aβ1-42-induced free radical production and neurotoxicity, Federation of American Societies for Experimental Biology, 2008.
- Akhtar MJ, Ahamed M, Alhadlaq HA, Khan MAM, Alrokayan SA. Glutathione replenishing potential of CeO 2 nanoparticles in human breast and fibrosarcoma cells. Journal of Colloid and Interface Science. 2015;453:21-7.
- 69. Pezzini I, Marino A, Del Turco S, Nesti C, Doccini S, Cappello V, et al. Cerium oxide nanoparticles: the regenerative redox machine in bioenergetic imbalance. Nanomedicine. 2017;12(4):403-16.
- 70. Sack M, Alili L, Karaman E, Das S, Gupta A, Seal S, et al. Combination of Conventional Chemotherapeutics with

Redox-Active Cerium Oxide Nanoparticles--A Novel Aspect in Cancer Therapy. Molecular Cancer Therapeutics. 2014;13(7):1740-9.

- Zschauer M S, S B, P B. Cerium Oxide Nanoparticles as Novel Tool in Glioma Treatment: An In vitro Study. Journal of Nanomedicine & Nanotechnology. 2017;08(06).
- 72. Dan M, Tseng M, Wu, Unrine, Grulke, Yokel. Brain microvascular endothelial cell association and distribution of a 5 nm ceria engineered nanomaterial. International Journal of Nanomedicine. 2012:4023.
- 73. Portioli C, Benati D, Pii Y, Bernardi P, Crucianelli M, Santucci S, et al. Short-Term Biodistribution of Cerium Oxide Nanoparticles in Mice: Focus on Brain Parenchyma. Nanoscience and Nanotechnology Letters. 2013;5(11):1174-81.
- 74. Yang L, Sundaresan G, Sun M, Jose P, Hoffman D, McDonagh PR, et al. Intrinsically radiolabeled multifunctional cerium oxide nanoparticles for in vivo studies. Journal of Materials Chemistry B. 2013;1(10):1421.
- 75. Rico CM, Hong J, Morales MI, Zhao L, Barrios AC, Zhang J-Y, et al. Effect of Cerium Oxide Nanoparticles on Rice: A Study Involving the Antioxidant Defense System and In Vivo Fluorescence Imaging. Environmental Science & Technology. 2013;47(11):5635-42.
- 76. Schwabe F, Tanner S, Schulin R, Rotzetter A, Stark W, von Quadt A, et al. Dissolved cerium contributes to uptake of Ce in the presence of differently sized CeO2-nanoparticles by three crop plants. Metallomics. 2015;7(3):466-77.
- 77. López-Moreno ML, de la Rosa G, Hernández-Viezcas JA, Peralta-Videa JR, Gardea-Torresdey JL. X-ray Absorption Spectroscopy (XAS) Corroboration of the Uptake and Storage of CeO2Nanoparticles and Assessment of Their Differential Toxicity in Four Edible Plant Species. Journal of Agricultural and Food Chemistry. 2010;58(6):3689-93.
- Majumdar S, Almeida IC, Arigi EA, Choi H, VerBerkmoes NC, Trujillo-Reyes J, et al. Environmental Effects of Nanoceria on Seed Production of Common Bean (Phaseolus vulgaris): A Proteomic Analysis. Environmental Science & Technology. 2015;49(22):13283-93.
- 79. Zhang W, Ebbs SD, Musante C, White JC, Gao C, Ma X. Uptake and Accumulation of Bulk and Nanosized Cerium Oxide Particles and Ionic Cerium by Radish (Raphanus sativus L.). Journal of Agricultural and Food Chemistry. 2015;63(2):382-90.
- Zhang Z, He X, Zhang H, Ma Y, Zhang P, Ding Y, et al. Uptake and distribution of ceria nanoparticles in cucumber plants. Metallomics. 2011;3(8):816.
- 81. Sisubalan N, Ramkumar VS, Pugazhendhi A, Karthikeyan C, Indira K, Gopinath K, et al. ROS-mediated cytotoxic activity of ZnO and CeO2 nanoparticles synthesized using the Rubia cordifolia L. leaf extract on MG-63 human osteosarcoma cell lines. Environmental Science and Pollution Research. 2017;25(11):10482-92.
- 82. Collin B, Oostveen E, Tsyusko OV, Unrine JM. Influence of Natural Organic Matter and Surface Charge on the Toxicity and Bioaccumulation of Functionalized Ceria Nanoparticles in Caenorhabditis elegans. Environmental Science & Technology. 2014;48(2):1280-9.
- Kwon HJ, Cha M-Y, Kim D, Kim DK, Soh M, Shin K, et al. Mitochondria-Targeting Ceria Nanoparticles as Antioxidants for Alzheimer's Disease. ACS Nano. 2016;10(2):2860-70.
- 84. M. Hijaz, S. Das, I. Mert, A. Gupta, Z. Al-Wahab, C. Tebbe,

S. Dar, J. Chhina, S. Giri, A. Munkarah, Folic acid tagged nanoceria as a novel therapeutic agent in ovarian cancer, BMC Cancer, 16 (2016) 220.

- Wong LL, Pye QN, Chen L, Seal S, McGinnis JF. Defining the Catalytic Activity of Nanoceria in the P23H-1 Rat, a Photoreceptor Degeneration Model. PLOS ONE. 2015;10(3):e0121977.
- 86. Rahdar A, Aliahmad M, Hajinezhad MR, Samani M. Xanthan gum-stabilized nano-ceria: Green chemistry based synthesis, characterization, study of biochemical alterations induced by intraperitoneal doses of nanoparticles in rat. Journal of Molecular Structure. 2018;1173:166-72.
- 87. Rocca A, Moscato S, Ronca F, Nitti S, Mattoli V, Giorgi M, et al. Pilot in vivo investigation of cerium oxide nanoparticles as a novel anti-obesity pharmaceutical formulation. Nanomedicine: Nanotechnology, Biology and Medicine. 2015;11(7):1725-34.
- Manne Nandini DPK, Arvapalli R, Graffeo Vincent A, Bandarupalli Venkata VK, Shokuhfar T, Patel S, et al. Prophylactic Treatment with Cerium Oxide Nanoparticles Attenuate Hepatic Ischemia Reperfusion Injury in Sprague Dawley Rats. Cellular Physiology and Biochemistry. 2017;42(5):1837-46.
- Heckert EG, Karakoti AS, Seal S, Self WT. The role of cerium redox state in the SOD mimetic activity of nanoceria. Biomaterials. 2008;29(18):2705-9.
- 90. Nicholls P. Classical catalase: Ancient and modern. Archives of Biochemistry and Biophysics. 2012;525(2):95-101.
- Pirmohamed T, Dowding JM, Singh S, Wasserman B, Heckert E, Karakoti AS, et al. Nanoceria exhibit redox state-dependent catalase mimetic activity. Chemical Communications. 2010;46(16):2736.
- 92. Cohen P. The origins of protein phosphorylation. Nature Cell Biology. 2002;4(5):E127-E30.
- 93. Kuchma MH, Komanski CB, Colon J, Teblum A, Masunov AE, Alvarado B, et al. Phosphate ester hydrolysis of biologically relevant molecules by cerium oxide nanoparticles. Nanomedicine: Nanotechnology, Biology and Medicine. 2010;6(6):738-44.
- 94. Dhall A, Burns A, Dowding J, Das S, Seal S, Self W. Characterizing the phosphatase mimetic activity of cerium oxide nanoparticles and distinguishing its active site from that for catalase mimetic activity using anionic inhibitors. Environmental Science: Nano. 2017;4(8):1742-9.
- Xue Y, Luan Q, Yang D, Yao X, Zhou K. Direct Evidence for Hydroxyl Radical Scavenging Activity of Cerium Oxide Nanoparticles. The Journal of Physical Chemistry C. 2011;115(11):4433-8.
- Knott AB, Bossy-Wetzel E. Nitric Oxide in Health and Disease of the Nervous System. Antioxidants & Redox Signaling, 2009;11(3):541-53.
- 97. Bauer LA, Birenbaum NS, Meyer GJ. Biological applications of high aspect ratio nanoparticles. Journal of Materials Chemistry. 2004;14(4):517.
- 98. Ji Z, Wang X, Zhang H, Lin S, Meng H, Sun B, et al. Designed Synthesis of CeO2 Nanorods and Nanowires for Studying Toxicological Effects of High Aspect Ratio Nanomaterials. ACS Nano. 2012;6(6):5366-80.
- Prabaharan DMDM, Sadaiyandi K, Mahendran M, Sagadevan S. Structural, Optical, Morphological and Dielectric Properties of Cerium Oxide Nanoparticles. Materials Research. 2016;19(2):478-82.
- 100. Lord MS, Jung M, Teoh WY, Gunawan C, Vassie JA,

Amal R, et al. Cellular uptake and reactive oxygen species modulation of cerium oxide nanoparticles in human monocyte cell line U937. Biomaterials. 2012;33(31):7915-24.

- 101. Cheng G, Guo W, Han L, Chen E, Kong L, Wang L, et al. Cerium oxide nanoparticles induce cytotoxicity in human hepatoma SMMC-7721 cells via oxidative stress and the activation of MAPK signaling pathways. Toxicology in Vitro. 2013;27(3):1082-8.
- 102. Lipinski B. Hydroxyl Radical and Its Scavengers in Health and Disease. Oxidative Medicine and Cellular Longevity. 2011;2011:1-9.
- 103. Fröhlich E. The role of surface charge in cellular uptake and cytotoxicity of medical nanoparticles. International Journal of Nanomedicine. 2012:5577.
- 104. Pulido-Reyes G, Rodea-Palomares I, Das S, Sakthivel TS, Leganes F, Rosal R, et al. Untangling the biological effects of cerium oxide nanoparticles: the role of surface valence states. Scientific Reports. 2015;5(1).
- 105. Singh AV, Bandgar BM, Kasture M, Prasad BLV, Sastry M. Synthesis of gold, silver and their alloy nanoparticles using bovine serum albumin as foaming and stabilizing agent. Journal of Materials Chemistry. 2005;15(48):5115.
- 106. K.V. Hoecke, J.T. Quik, J. Mankiewicz-Boczek, K.A.D. Schamphelaere, A. Elsaesser, P.V.d. Meeren, C. Barnes, G. McKerr, C.V. Howard, D.V.D. Meent, Fate and effects of CeO₂ nanoparticles in aquatic ecotoxicity tests, Environmental science & technology, 43 (2009) 4537-4546.
- 107. Xia T, Kovochich M, Liong M, Mädler L, Gilbert B, Shi H, et al. Comparison of the Mechanism of Toxicity of Zinc Oxide and Cerium Oxide Nanoparticles Based on Dissolution and Oxidative Stress Properties. ACS Nano. 2008;2(10):2121-34.
- 108. Sadhu A, Ghosh I, Moriyasu Y, Mukherjee A, Bandyopadhyay M. Role of cerium oxide nanoparticleinduced autophagy as a safeguard to exogenous H2O2mediated DNA damage in tobacco BY-2 cells. Mutagenesis. 2018;33(2):161-77.
- 109. F. Soltani, K. Yavari, M. Sadeghi, A. BAHRAMI SAMANI, S. SHIRVANI ARANI, Toxicity of nano and bulk forms of cerium oxide in different cell lines, Iranian Journal of Pharmacology and Therapeutics, 16 (2018) 1-6.
- 110. Lin W, Huang Y-w, Zhou X-D, Ma Y. Toxicity of Cerium Oxide Nanoparticles in Human Lung Cancer Cells. International Journal of Toxicology. 2006;25(6):451-7.
- 111. Renu G, Rani VVD, Nair SV, Subramanian KRV, Lakshmanan V-K. Development of Cerium Oxide Nanoparticles and Its Cytotoxicity in Prostate Cancer Cells. Advanced Science Letters. 2012;6(1):17-25.
- 112. Wason MS, Colon J, Das S, Seal S, Turkson J, Zhao J, et al. Sensitization of pancreatic cancer cells to radiation by cerium oxide nanoparticle-induced ROS production. Nanomedicine: Nanotechnology, Biology and Medicine. 2013;9(4):558-69.
- 113. Wason MS, Zhao J. Cerium oxide nanoparticles: potential applications for cancer and other diseases. American journal of translational research. 2013;5(2):126.
- 114. Gao Y, Gao F, Chen K, Ma J-l. Cerium oxide nanoparticles in cancer. OncoTargets and Therapy. 2014:835.
- 115. Sack M, Alili L, Karaman E, Das S, Gupta A, Seal S, et al. Combination of Conventional Chemotherapeutics with Redox-Active Cerium Oxide Nanoparticles--A Novel Aspect in Cancer Therapy. Molecular Cancer Therapeutics. 2014;13(7):1740-9.

Z. Yaghoobi et al. / the cytotoxicity of CeO, nanoparticles: A compendious approach

- 116. Yao C, Wang W, Wang P, Zhao M, Li X, Zhang F. Near-Infrared Upconversion Mesoporous Cerium Oxide Hollow Biophotocatalyst for Concurrent pH-/H2 O2 -Responsive O2 -Evolving Synergetic Cancer Therapy. Advanced Materials. 2018;30(7):1704833.
- 117. Forest V, Leclerc L, Hochepied J-F, Trouvé A, Sarry G, Pourchez J. Impact of cerium oxide nanoparticles shape on their in vitro cellular toxicity. Toxicology in Vitro. 2017;38:136-41.
- 118. Wang L, Ai W, Zhai Y, Li H, Zhou K, Chen H. Effects of Nano-CeO2 with Different Nanocrystal Morphologies on Cytotoxicity in HepG2 Cells. International Journal of Environmental Research and Public Health. 2015;12(9):10806-19.
- 119. Malavasi L, Fisher CAJ, Islam MS. Oxide-ion and proton conducting electrolyte materials for clean energy applications: structural and mechanistic features. Chemical Society Reviews. 2010;39(11):4370.