

Trends in Phytochemical Research (TPR)



Original Research Article

Chemical composition of the essential oils from the aerial parts of *Malva neglecta* Wallr. from Khorramabad, Lorestan Province, Iran using solvent free microwave extraction (SFME) method

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ABSTRACT

This study deals with the investigation of the efficacy of solvent free microwave extraction (SFME) method to extract the essential oils from the aerial parts of *Malva neglecta* Wallr.. The essential oils were then injected onto an HP-5MS column of a commercially available GC/MS (Hewlett-Packard 5973), which resulted in a chromatogram consisting of 24 compounds accounting for 99.9% of the oil composition. In terms of general categories, non-terpene hydrocarbons and sesquiterpene hydrocarbons were found to be the major fractions of the chemical profiles. Moreover, hinokione was recognized as the most abundant constituent component of the essential oil comprising 40.7% of the total oil structure. Regarding our findings in this study, the SFME can be introduced as a rapid, cost-effective, and environmentally friendly extraction method for the separation of the essential oils.

ARTICLE HISTORY

Received: 07 March 2021 Revised: 19 June 2021 Accepted: 30 August 2021 ePublished: 29 September 2021

K E Y W O R D S

Essential oil Hinokione *Malva neglecta Wallr.* Non-terpene hydrocarbons Solvent free microwave extraction (SFME)

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1. Introduction

he genus Malva has about 10 herbal and shrub species native to Iran and in Persian and local traditional treatment is called "Panirak". Some species of this genus including M. armeniaca, M. leonardi, M. flexuosa, M. rotundifolia, M. Iljini, M. nicaeensis, M. parviflora and M. microcarpa, grow in Turkmenistan, Middle Asia, Mediterranean regions, the Balkan Peninsula, Anatolia, Iraq, Afghanistan and Armenia in addition to Iran (Mozaffarian, 1996). Malva neglecta Wallr. belongs to Mallow Family (Malvaceae) and is an annual herb that grows to a height of 0.6 m (2 ft). This medicinal species is known as common mallow in the United States and also button or cheese weed, cheese plant, dwarf (round leaf or garden or low or roundleaved or running) mallow, malice and round dock (Zargari, 1989). Although often considered a weed, this plant is consumed as a food worldwide (Facciola, 1990).

This is especially true for the seeds, that consiststs of 21% protein and 15.2% fat (Duke and Atchley, 1986). The plant (*M. neglecta* Wallr.) is shown to be an invasive species in the United States and commonly occurs in disturbed sites such as roadsides, railroads, waste places, gravel pits, nurseries, gardens, and cultivated fields. It is frequently found growing in yards around homes, buildings, and barns (Peterson and McKinney, 1968). Currently, promising phytochemical activities of different species of Malva genus have been reported, including potential antioxidant (Mavi et al., 2004; DellaGreca et al., 2009; Stef et al., 2009; Bouriche et al., 2010; Bouriche et al., 2011; Ghanati and Khatami, 2011; Dalar et al., 2012; Guder and Korkmaz, 2012; Tesevic et al., 2012; Turker and Dalar, 2013; Khan et al., 2016), antibacterial (Grierson and Afolayan, 1999; Keles et al., 2001; Kumarasamy et al., 2002; Shale et al., 2004; Bernardo et al., 2005; Shale et al., 2005; Cogo et al., 2010; Seyyednejad et al., 2010; Razavi et al., 2011; Walter et al., 2011; Khan et al., 2016),

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antinociceptive (Esteves et al., 2009), antiproliferative (Conforti et al., 2008), anti-inflammatory (Franzotti et al., 2000; Sleiman and Daher, 2009; Afolayan et al., 2010; Bouriche et al., 2010; Bouriche et al., 2011; Benso et al., 2015), antimicrobial (de Souza et al., 2004; Alves et al., 2009; Razavi et al., 2011; Vitullo et al., 2011; Benso et al., 2015; Khan et al., 2016), anticancer (Huang et al., 1998; Daniela et al., 2007), anti-complementary mucilage (Tomoda et al., 1989), antifungal (Wang and Bunkers, 2000; Wang et al., 2001; Andrade Pinto et al., 2010; Oshchepkova et al., 2012; Romitelli and Martins, 2013) and antiadherent properties (Alves et al., 2009). These findings account for the possibility of considering this medicinal plant as an alternative to chemical drugs that have harmful side effects. Hydrodistillation (HD) and steam distillation (SD) methods have been used for a long time to extract the essential oils from a broad spectrum of medicinal plants as conventional methods. However, these traditional procedures often need large amounts of plant samples and are also not cost-effective and timesaving (Akhlaghi et al., 2009a; Akhlaghi et al., 2011; Mohammadhosseini, 2012a, b; Mohammadhosseini, 2013; Motavalizadehkakhky et al., 2013; Shahnama et al., 2015). Therefore, development of proper alternative facile and secure techniques which are amenable to automation seems rationale and unavoidable (Nekoei and Mohammadhosseini, 2014). Over the recent decades, there has been a growing interest in using microwave radiation for the separation of the essential oils from a wide spectrum of plant organs (Hashemi-Moghaddam et al., 2015; Mohammadhosseini, 2016). The technique solvent-free microwave-assisted extraction (SFME) was developed and then patented by Chemat et al. in France (2013). This technique could be considered among the newest approaches in the extraction of the essential oils applying the microwave

beams in the conditions that no solvent and/or water are being used under the atmospheric pressure. The general SFME setup involves four parts which are: i) one reactor where only matrix to be processed is placed, ii) a standard microwave oven, iii) the refrigeration system, and iv) essence container where essential oil is recovered (see Fig. 1). In fact, the extraction process using the SFME technique accounts for dry distillation assisted by microwave beams being focused on the fresh matrix in a microwave reactor without the addition of water or any other organic solvent. Water heating of the raw material breaks the essential oil containing glands. This phase releases the essential oil, which is then driven by steam produced from matrix water. A cooling system placed outside the microwave oven allows the continuous condensation of the distillate, which consists of water and the essential oil, while the excessive water returns inside the balloon allowing maintenance of the proper humidity rate of the matrix (Chemat and Cravotto, 2013; Mohammadhosseini, 2016). The extraction process is frequently continued at 100 °C until no more essential oil is obtained. In the final step, the isolated essential oil is collected, dried with anhydrous sodium sulfate and stored at 4 °C prior to the analysis using gas chromatography coupled to mass spectrometry (GC-MS). According to the literature, the SFME technique has been successfully applied to several kinds of fresh and dry plants, including spices, aromatic herbs, and citrus fruits (Lucchesi et al., 2004a, b; Ma et al., 2012; Mohammadhosseini and Nekoei, 2014; Mohammadhosseini, 2015; Mohammadhosseini et al., 2015; Nekoei and Mohammadhosseini, 2016). To the best of our knowledge, this is the first report concerning the chemical compositions of the essential oils from the aerial parts of M. neglecta Wallr. obtained by the SFME technique.



Fig. 1. The overall scheme (left) and apparatus (right) of the solvent-free microwave extraction (SFME) technique.

2. Experimental

2.1. Plant material and botanical identification

The plant material was collected during the flowering stage in April 2020 from Khorramabad region, Lorestan

Province, Iran, at the geographical coordinates of 33.4878° N, 48.3558° E respectively at an altitude of 1147.8 meters above sea level. A voucher specimen was deposited at the Herbarium of the Faculty of Agriculture (Herbarium number 8894), Shahrood University of Technology, Shahrood, Iran, for further authentication.



2.2. Details of the SFME technique

The general characteristics associated with this technique were described in our previous report with slight modifications (Mohammadhosseini and Nekoei, 2014). Accordingly, 100-g portions of the powdered and air-dried aerial parts of *M. neglecta* Wallr. were soaked in 1000 mL of distilled water. This preliminary step was mainly to soak the external and outer layers of the plant organs prior to the extraction under the atmospheric pressure. At an optimized and fixed power of 800 W, the extraction continued for 30 minutes, and the mean yield of the oil obtained by this method was found to be 0.21% (w/w).

2.3. Chromatographic analyses

GC analyses were performed on a Shimadzu 15A gas chromatograph equipped with a split/splitless (ratio 1:30) injector and a flame ionization detector (FID), both operating at 250 °C. High purity nitrogen was utilized as the carrier gas having a flow-rate of 1 mL/min. The capillary column used was DB-5 (50 m×0.2 mm, film thickness 0.32 μ m) and its temperature was kept at 60 °C for 3 minutes, then heated to 220 °C with a 5 °C/min rate and finally kept constant at 220 °C for 5 minutes. Relative percentage amounts were calculated from peak areas using a CR5 Shimadzu CR pack without the use of correction factors. In addition, GC/MS analyses were performed using a Hewlett-Packard 5973 instrument equipped with an HP-5MS column (30 m×0.25 mm, film thickness 0.25 µm). The column temperature programming was the same with that mentioned for the GC analysis, reaching to a final temperature of 230 °C. The effluent of the GC column was directly transferred into the MS source producing the charged fragments. The flow-rate of helium as the carrier gas was regulated at 1 mL/min. The detector temperature (MS) was set at 250 °C and all the MS spectra were recorded at 70 eV (E1) over the range 30-350 amu. The optimal voltage of the electron multiplier was set at 1800 eV and scan time was fixed to be 2 scans/sec.

2.4. Qualitative and quantitative analyses

Identification and determination of the constituents of the essential oils of *M. neglecta* Wallr. (aerial parts) were tentatively made by comparison of their mass spectral fragmentation patterns and retention indices (RI) relative to C_9 - C_{25} *n*-alkanes, both with those given in the literature and those stored in the MS library (Wiley 275). For further reliability, some of the spectral patterns as well as Kovats indices based on previous findings of our research group were utilized (Mohammadhosseini et al., 2008; Akhlaghi et al., 2009b; Mohammadhosseini et al., 2010; Mohammadhosseini et al., 2011a; Mohammadhosseini et al., 2011b; Akhlaghi et al., 2012; Mohammadhosseini et al., 2012; Nekoei et al., 2012; Mohammadhosseini and Beiranvand, 2013; Mohammadhosseini et al., 2013; Hashemi-Moghaddam et al., 2014; Mohammadhosseini et al., 2016a; Mohammadhosseini et al., 2016b; Rahimi et al., 2016). Relative percentages of the components were calculated from peak areas using a Shimadzu C-R4A Chromatopac on the DB-5 column without the use of a correction factor.

3. Results and Discussion

3.1. Chemical profiles by the SFME method

The volatile constituents of the essential oils obtained from the aerial parts of *M. neglecta* Wallr. are listed in Table 1, in which both the percentages and the retention indices of the components are given. As can be seen, there are negligible differences in the numerical values of the Kovats retention indices between the calculated ones and those cited in the literature. Furthermore, a total of 24 components were obtained, accounting for 99.9% of the chemical profile (see Fig. 2). The Kovats retention indices were calculated by co-injecting a mixture of normal alkanes (n=9-24) into the column under the same conditions with the essential oil sample. As far as we know, at this time, there is no paper in the literature dealing with the characterization of the chemical composition of the essential oils of M. neglecta Wallr.. Using the SFME method (Table 1 and Fig. 2), twenty-one components could be identified in the M. neglecta Wallr. oil from its aerial parts: three oxygenated monoterpenes (5.4%), ten sesquiterpene hydrocarbons (36.6%), five oxygenated sesquiterpenes (10.1%) and three non-terpene hydrocarbons (46.1%), while only three compounds remained unidentified, totally constituting 1.8% of the essential oil composition. Specifically, the major components in the hydrodistilled oil from the aerial parts of M. neglecta Wallr. were found to be hinokione (40.7%), γ -elemene (9.4%), δ -elemene (6.8%), β-ylangene (5.8%) and bicyclogermacrene Moreover, some components including (5.1%). L-linalool (3.8%), spathulenol (3.8%), viridiflorol (3.1%), n-tetradecane (2.9%), germacrene D (2.8%), 6,10,14-trimethyl-2-pentadecanone (2.5%), β-elemene (2.0%), γ-curcumene (1.8%), δ-cadinene (1.2%), α-cadinol (1.2%), β-damascenone (1.2%), α-muurolol (1.1%), cis-muurola-3,5-diene (1.0%) occurred in the oil structure in lower quantities. On the other hand, very low quantities of natural compounds involving (E)-nerolidol (0.9%), cis-cadina-1(6),4-diene (0.7%) and α -terpineol (0.4%) were recognized in the chemical profile, each contributing to less than one percent (<1.0%) of the total oil composition.

In Fig. 3, relative percentages of the various classes of the volatile oil compounds have been compared. From this Figure, it is evident that in the SFME profile of the essential oils from the aerial parts of *M. neglecta* Wallr., non-terpene hydrocarbons constituted the main components, with the second largest class being sesquiterpene hydrocarbons. Furthermore, the third, fourth, and fifth rank in the chemical profile are due to oxygenated sesquiterpenes, oxygenated monoterpenes and non-identified compounds, respectively.





Fig. 2. The chromatogram of the volatile essential obtained from the aerial parts of *Malva neglecta* Wallr. using the SFME method.



Fig. 3. Percentages of the classes of natural compounds found in the volatiles from aerial parts of *Malva neglecta* Wallr. using the SFME method (OM= oxygenated monoterpenes; SH= sesquiterpene hydrocarbon; OS= oxygenated sesquiterpenes; NH= non-terpene hydrocarbon; NI= non-identified compounds).



Table 1

Chemical compositions of the essential oils from the aerial parts of *Malva neglecta* Wallr. obtained using the SFME method. ^a

Ser. Num.	Name	Group	Molecular formula	KI (Lit.) ^ь	KI (Cal.) ^c	Aerial parts (%)
1	L-Linalool	ОМ	C ₁₀ H ₁₈ O	1098	1100.5	3.8
2	α-Terpineol	OM	C ₁₀ H ₁₈ O	1188	1194.2	0.4
3	NI	-	-	-	1254.3	0.7
4	δ-Elemene	SH	C ₁₅ H ₂₄	1337	1328	6.8
5	β-Damascenone	OM	C ₁₃ H ₁₈ O	1391	1386.5	1.2
6	β-Elemene	SH	C ₁₅ H ₂₄	1393	1393.9	2
7	β-Ylangene	SH	C ₁₅ H ₂₄	1419	1423.4	5.8
8	γ-Elemene	SH	C ₁₅ H ₂₄	1433	1435.7	9.4
9	<i>cis</i> -Muurola-3,5-diene	SH	C ₁₅ H ₂₄	1438	1449	1
10	<i>n</i> -Tetradecane	NH	C ₁₄ H ₃₀	1400	1458.5	2.9
11	cis-Cadina-1(6),4-diene	SH	C ₁₅ H ₂₄	1463	1467.7	0.7
12	γ-Curcumene	SH	C ₁₅ H ₂₄	1482	1482.1	1.8
13	Germacrene D	SH	C ₁₅ H ₂₄	1480	1486.2	2.8
14	Bicyclogermacrene	SH	C ₁₅ H ₂₄	1494	1501.3	5.1
15	NI	-	-	-	1510	0.6
16	δ-Cadinene	SH	C ₁₅ H ₂₄	1524	1526.5	1.2
17	(E)-Nerolidol	OS	C ₁₅ H ₂₆ O	1564	1564.8	0.9
18	Spathulenol	OS	C ₁₅ H ₂₄ O	1576	1584.5	3.8
19	Viridiflorol	OS	C ₁₅ H ₂₆ O	1590	1591	3.1
20	NI	-	-	-	1628.8	0.4
21	α-Muurolol	OS	C ₁₅ H ₂₆ O	1644	1646.1	1.1
22	α-Cadinol	OS	C ₁₅ H ₂₆ O	1653	1659.7	1.2
23	6,10,14-Trimethyl-2-pentadecanone	NH	C ₁₈ H ₃₆ O	1846	1845.4	2.5
24	Hinokione	NH	C ₂₀ H ₂₈ O ₂	2549	2549	40.7
Oxygen-containing monoterpens (OM)						5.4
Sesquiterpene hydrocarbons (SH)						36.6
Oxygen-containing sesquiterpenes (OS)						10.1
Non-terpene compounds (NH)						46.1
None identified compounds (NI)						1.7
Total percentage						99.9
Yield (%, w/w-dry basis)						0.24

^a Compounds are sorted in order of their elution from an HP-5MS capillary column.

^b Kovats retention index given in the literature

 $^{\rm c}$ Kovats retention index calculated with respect to *n*-alkenes on an HP-5MS capillary column.

3.2. Chemical composition of other species of *Malva* genus

Characterization of the essential oil composition relating to another species of *Malva genus*, namely *M. sylvestris* L, has been the subject of another report. In the study of Usami and co-workers (2013), the aromaactive compounds in dry flower of *M. sylvestris* L. were characterized using gas chromatography-mass spectrometry-olfactory (GC-MS-O) analysis and odor active values (OAV) calculations. Accordingly, in the light yellow isolated oil having a sweet odor and a percentage yield of 0.039% (w/w), totally 143 volatile compounds (89.9%) were identified, in which the most abundant compounds were found to be hexadecanoic acid (10.1%), pentacosane (4.8%) and 6,10,14-trimethyl-2-pentadecanone (4.1%). In this regard, the essential oil mainly consisted of hydrocarbons (25.4%) followed by alcohols (18.8%), acids (16.7%), ethers (5.0%), ketones (7.3%), esters (12.4%), aldehydes (2.3%) and other



compounds (2.0%).

4. Concluding remarks

In this report, the essential oils from the aerial parts of M. neglecta Wallr. from Khorramabad region, Lorestan Province, Iran were separated using the solvent free microwave extraction (SFME) method. This green method is capable of isolating essential oils, faster and more effective than the classical methods like hydrodistillation. Characterization of the corresponding chemical profiles of the isolated essences has been performed for the first time in the literature. This study led to the identification of a chemical profile consisting of hinokione (40.7%), γ -elemene (9.4%), δ -elemene (6.8%), β -ylangene (5.8%) and bicyclogermacrene (5.1%) as the major constituent components. In addition, among the 21 identified compounds in the oil from the aerial parts of M. neglecta Wallr., three oxygenated monoterpenes, ten sesquiterpene hydrocarbons, five oxygenated sesquiterpenes and three non-terpene hydrocarbons, respectively contributing to 5.4%, 36.6%, 10.1% and 46.1% of the total oil composition. These compounds constitute 99.9% of the oil structure, whereas only three compounds remained unidentified altogether forming about 1.7% of the oil profile.

Acknowledgements

Financial and technical support from the Office for Research Affairs of the Islamic Azad University, Shahrood Branch is gratefully acknowledged. In addition, special thanks are due to Dr. Gholami for botanical identification and authentication of the plant sample.

References

Afolayan, A.J., Aboyade, O.M., Adedapo, A.A., Sofidiya, M.O., 2010. Anti-inflammatory and analgesic activity of the methanol extract of Malva parviflora Linn (Malvaceae) in rats. Afr .J. Biotechnol. 9, 1225-1229. Akhlaghi, H., Asl, M.R.S., Mohammadhosseini, M., 2009a. Composition of the essential oil of *Hymnocrater* platystegius in Iran. Chem. Nat. Compd. 45, 448-449. Akhlaghi, H., Nekoei, M., Mohammadhosseini, M., Motavalizadehkakhky, A., 2012. Chemical composition of the volatile oils from the flowers, stems and leaves of Prangos latiloba Korov. using the head space solid phase microextraction method prior to analysis by gas chromatography-mass spectrometry. J. Essent. Oil-Bear. Plants 15, 328-335. Akhlaghi, H., Shafaghat, A., Mohammadhosseini, M., 2009b. Chemical composition of the essential oil from leaves of Biebersteinia multifida DC. growing wild in Iran. J. Essent. Oil-Bear. Plants 12, 365-368. Akhlaghi, H., Shafaghat, A., Salimi, F., Mohammadhosseini, M., 2011. GC/MS analysis of the essential oil from wild Stachys pubescens Ten. leaves from Northwest Iran. Anal. Chem. Lett. 1, 325-327.

Alves, P.M., Guedes Queiroz, L.M., Pereira, J.V., Vieira Pereira, M.d.S., 2009. *In vitro* antimicrobial, antiadherent and antifungal activity of Brazilian medicinal plants on oral biofilm microorganisms and strains of the genus *Candida*. Rev. Soc. Bras. Med. Trop. 42, 222-224. Andrade Pinto, J.M., Souza, E.A., Oliveira, D.F., 2010. Use of plant extracts in the control of common bean anthracnose. Crop Prot. 29, 838-842.

Benso, B., Rosalen, P.L., Alencar, S.M., Murata, R.M., 2015. *Malva sylvestris* inhibits inflammatory response in oral human cells. An *in vitro* infection model. Plos One 10, e0140331.

Bernardo, W.L.d.C., Boriollo, M.F.G., Gonçalves, R.B., Höfling, J.F., 2005. *Staphylococcus aureus* ampicillinresistant from the odontological clinic environment. Rev. Inst. Med. Trop. Sao Paulo 47, 19-24.

Bouriche, H., Meziti, H., Senator, A., 2010. *In vivo* antiinflammatory and antioxidant effects of *Malva parviflora* leaf extracts, in: Safwat, M.S.A., Tantawy, A., Moaty, E.A. (Eds.), XIII International Conference on Medicinal and Aromatic Plants, pp. 23-30.

Bouriche, H., Meziti, H., Senator, A., Arnhold, J., 2011. Anti-inflammatory, free radical-scavenging, and metalchelating activities of *Malva parviflora*. Pharm. Biol. 49, 942-946.

Chemat, F., Cravotto, G., 2013. Microwave-Assisted Extraction for Bioactive Compounds, Theory and Practice. Springer, New York.

Cogo, L.L., Bastos Monteiro, C.L., Miguel, M.D., Miguel, O.G., Cunico, M.M., Ribeiro, M.L., de Camargo, E.R., Botao Kussen, G.M., Nogueira, K.d.S., Dalla Costa, L.M., 2010. Anti-Helicobacter pylori activity of plant extracts traditionally used for the treatment of gastrointestinal disorders. Braz. J. Microbiol. 41, 304-309.

Conforti, F., Ioele, G., Statti, G.A., Marrelli, M., Ragno, G., Menichini, F., 2008. Antiproliferative activity against human tumor cell lines and toxicity test on Mediterranean dietary plants. Food Chem. Toxicol. 46, 3325-3332.

Dalar, A., Turker, M., Konczak, I., 2012. Antioxidant capacity and phenolic constituents of *Malva neglecta* Wallr. and *Plantago lanceolata* L. from Eastern Anatolia region of Turkey. J. Herb. Med. 2, 42-51. Daniela, A., Pichichero, E., Canuti, L., Cicconi, R., Karou, D., D'Arcangelo, G., Canini, A., 2007. Identification of phenolic compounds from medicinal and melliferous plants and their cytotoxic activity in cancer cells. Caryologia 60, 90-95.

de Souza, G.C., Haas, A.P.S., von Poser, G.L., Schapoval, E.E.S., Elisabetsky, E., 2004. Ethnopharmacological studies of antimicrobial remedies in the south of Brazil. J. Ethnopharmacol. 90, 135-143.

DellaGreca, M., Cutillo, F., D'Abrosca, B., Fiorentino, A., Pacifico, S., Zarrelli, A., 2009. Antioxidant and radical scavenging properties of of *Malva sylvestris*. Nat. Prod. Commun. 4, 893-896.

Duke, J.A., Atchley, A.A., 1986. CRC Handbook of Proximate Analysis Tables of Higher Plants. CRC Press. Esteves, P.F., Sato, A., Esquibel, M.A., de Campos-Buzzi, F., Meira, A.V., Cechinel-Filho, V., 2009. Antinociceptive activity of *Malva sylvestris* L. Lat. Am. J. Pharm. 28, 454-456.

Facciola, S., 1990. Cornucopia: A Source Book of Edible



Plants. Kampong Publications, 1870 Sunrise Drive, Vista, CA 92084, USA.

Franzotti, E.M., Santos, C.V.F., Rodrigues, H., Mourao, R.H.V., Andrade, M.R., Antoniolli, A.R., 2000. Antiinflammatory, analgesic activity and acute toxicity of *Sida cordifolia* L. (*Malva-branca*). J. Ethnopharmacol. 72, 273-278.

Ghanati, F., Khatami, F., 2011. Polyphenols and their antioxidant activity in callus-cultured *Malva neglecta* cells under UV-B and UV-C irradiation. Planta Med. 77, 1286-1286.

Grierson, D.S., Afolayan, A.J., 1999. Antibacterial activity of some indigenous plants used for the treatment of wounds in the Eastern Cape, South Africa. J. Ethnopharmacol. 66, 103-106.

Guder, A., Korkmaz, H., 2012. Evaluation of *in-vitro* antioxidant properties of hydroalcoholic solution extracts *Urtica dioica* L., *Malva neglecta* Wallr. and their mixture. Iran. J. Pharm. Res. 11, 913-923.

Hashemi-Moghaddam, H., Mohammadhosseini, M., Basiri, M., 2015. Optimization of microwave assisted hydrodistillation on chemical compositions of the essential oils from the aerial parts of *Thymus pubescens* and comparison with conventional hydrodistllation. J. Essent. Oil-Bear. Plants 18, 884-893.

Hashemi-Moghaddam, H., Mohammadhosseini, M., Salar, M., 2014. Chemical composition of the essential oils from the hulls of *Pistacia vera* L. by using magnetic nanoparticle-assisted microwave (MW) distillation: Comparison with routine MW and conventional hydrodistillation. Anal. Methods 6, 2572-2579.

Huang, C.Y., Zeng, L.F., He, T., Wang, C.J., Hong, J.R., Zhang, X.Q., Hou, Y.H., Peng, S.S., 1998. *In vivo* and *in vitro* studies on the antitumor activities of MCP (*Malva crispa* L. powder). Biomed. Environ. Sci. 11, 297-306. Keles, O., Ak, S., Bakirel, T., Alpinar, K., 2001. Screening of some Turkish plants for antibacterial activity. Turk. J. Vet. Anim. Sci. 25, 559-565.

Khan, H., Jan, S.A., Javed, M., Shaheen, R., Khan, Z., Ahmad, A., Safi, S.Z., Imran, M., 2016. Nutritional composition, antioxidant and antimicrobial activities of selected wild edible plants. J. Food Biochem. 40, 61-70. Kumarasamy, Y., Cox, P.J., Jaspars, M., Nahar, L., Sarker, S.D., 2002. Screening seeds of Scottish plants for antibacterial activity. J. Ethnopharmacol. 83, 73-77. Lucchesi, M.E., Chemat, F., Smadja, J., 2004a. An original solvent free microwave extraction of essential oils from spices. Flav. Fragr. J. 19, 134-138. Lucchesi, M.E., Chemat, F., Smadja, J., 2004b. Solventfree microwave extraction of essential oil from aromatic herbs: comparison with conventional hydro-distillation. J. Chromatogr. A 1043, 323-327.

Ma, C.-h., Yang, L., Zu, Y.-g., Liu, T.-t., 2012. Optimization of conditions of solvent-free microwave extraction and study on antioxidant capacity of essential oil from *Schisandra chinensis* (Turcz.) Baill. Food Chem. 134, 2532-2539.

Mavi, A., Terzi, Z., Ozgen, U., Yildirim, A., Coskun, M., 2004. Antioxidant properties of some medicinal plants: *Prangos ferulacea* (Apiaceae), *Sedum sempervivoides* (Crassulaceae), *Malva neglecta* (Malvaceae), *Cruciata* taurica (Rubiaceae), Rosa pimpinellifolia (Rosaceae), Galium verum subsp verum (Rubiaceae), Urtica dioica (Urticaceae). Biol. Pharm. Bull. 27, 702-705. Mohammadhosseini, M., 2012a. Chemical profile and antibacterial activity in hydrodistilled oil from aerial parts of *Prangos ferulacea* (L.) Lindl. and prediction of gas chromatographic retention indices by using genetic algorithm multiple linear regressions. Asian J. Chem. 24, 3814-3820.

Mohammadhosseini, M., 2012b. Hydrodistilled volatile oils of the flowers of *Salvia leriifolia* Bench. and *Salvia multicaulis* Vahl. as two growing wild plants in Iran. Asian J. Chem. 24, 1432-1434.

Mohammadhosseini, M., 2013. Hydrodistilled volatile oil from stems of *Eryngium creticum* Lam. in the marginal brackish regions of Semnan province by using gas chromatography combined with mass spectrometry. Asian J. Chem. 25, 390-392.

Mohammadhosseini, M., 2015. Chemical composition of the essential oils and volatile fractions from flowers, stems and roots of *Salvia multicaulis* Vahl. by using MAHD, SFME and HS-SPME methods. J. Essent. Oil-Bear. Plants 18, 1360-1371.

Mohammadhosseini, M., 2016. A Comprehensive Review on New Methods for Processing, Separation and Identification of the Essential Oils. Islamic Azad University of Shahrood Press, Shahrood, Iran. Mohammadhosseini, M., Akbarzadeh, A., Hashemi-Moghaddam, H., Mohammadi Nafchi, A., Mashayekhi, H.A., Aryanpour, A., 2016a. Chemical composition of the essential oils from the aerial parts of *Artemisia sieberi* by using conventional hydrodistillation and microwave assisted hydrodistillation: A comparative study. J. Essent. Oil-Bear. Plants 19, 32-45.

Mohammadhosseini, M., Akbarzadeh, A., Hashemi-Moghaddam, H., Shahnama, M., Fahimi, B., Azami, S., 2016b. Gas chromatographic-mass spectrometric analysis of volatiles obtained by HS-SPME-GC-MS technique from aerial parts of *Ziziphora Capitata* L., and evaluation for biological activity. Oriental J. Chem. 32(3), 1439-1451.

Mohammadhosseini, M., Beiranvand, M., 2013. Chemical composition of the essential oil from the aerial parts of *Satureja hortensis* as a potent medical plant using traditional hydrodistillation. J. Chem. Health Risks 3, 49-60.

Mohammadhosseini, M., Mahdavi, B., Akhlaghi, H., 2013. Characterization and chemical composition of the volatile oils from aerial parts of *Eryngium bungei* Bioss. (Apiaceae) by using traditional hydrodistillation, microwave assisted hydrodistillation and head space solid phase microextraction methods prior to GC and GC/MS analyses: A comparative approach. J. Essent. Oil-Bear. Plants 16, 613-623.

Mohammadhosseini, M., Mahdavi, B., Shahnama, M., 2015. Chemical composition of essential oils from aerial parts of *Ferula gummosa* (Apiaceae) in Jajarm Region, Iran using traditional hydrodistillation and solventfree microwave extraction methods: A comparative approach. J. Essent. Oil-Bear. Plants 18, 1321-1328. Mohammadhosseini, M., Nekoei, M., 2014. Chemical



compositions of the essential oils and volatile compounds from the aerial parts of *Ferula ovina* using hydrodistillation, MAHD, SFME and HS-SPME methods. J. Essent. Oil-Bear. Plants 17, 747-757. Mohammadhosseini, M., Nekoei, M., Mashayekhi, H.A., Aboli, J., 2012. Chemical composition of the essential oil from flowers, leaves and stems of *Haplophyllum perforatum* by using head space solid phase microextraction. J. Essent. Oil-Bear. Plants 15, 506-515. Mohammadhosseini, M., Pazoki, A., Akhlaghi, H., 2008. Chemical composition of the essential oils from flowers, stems, and roots of *Salvia multicaulis* growing wild in Iran. Chem. Nat. Compd. 44, 127-128.

Mohammadhosseini, M., Pazoki, A., Zamani, H.A., Akhlaghi, H., 2011a. Chemical composition of the essential oil from aerial parts of *Ajuga chamaecistus* Ging. subsp Scopria in brackish regions of Iran. J. Essent. Oil-Bear. Plants 14, 101-105.

Mohammadhosseini, M., Pazoki, A., Zamani, H.A., Akhlaghi, H., Nekoei, M., 2010. Chemical composition of the essential oil from aerial parts of *Senicio gallicus* Chaix growing wild in Iran. J. Essent. Oil-Bear. Plants 13, 704-709.

Mohammadhosseini, M., Zamani, H.A., Akhlaghi, H., Nekoei, M., 2011b. Hydrodistilled volatile oil constituents of the aerial parts of *Prangos serpentinica* (Rech.f., Aell. Esfand.) Hernnstadt and Heyn from Iran and quantitative structure-retention relationship simulation. J. Essent. Oil-Bear. Plants 14, 559-573. Motavalizadehkakhky, A.R., Shafaghat, A., Zamani, H.A., Akhlaghi, H., Mohammadhosseini, M., Mehrzad, J., Ebrahimi, Z., 2013. Compositions and the *in vitro* antimicrobial activities of the essential oils and extracts of two *Achillea* species from Iran. J. Med. Plants Res. 7, 1280-1292.

Mozaffarian, V., 1996. A Dictionary of Iranian Plant Names. Farhang Moaser Press, Iran.

Nekoei, M., Mohammadhosseini, M., 2014. Application of HS-SPME, SDME and cold-press coupled to GC/MS to analysis the essential oils of Citrus sinensis CV. Thomson Navel and QSRR study for prediction of retention indices by stepwise and genetic algorithm-multiple linear regression approaches. Anal. Chem. Lett. 4, 93-103. Nekoei, M., Mohammadhosseini, M., 2016. Chemical compositions of the essential oils from the aerial parts of Achillea wilhelmsii using traditional hydrodistillation, microwave assisted hydrodistillation and solvent-free microwave extraction methods: Comparison with the volatile compounds obtained by headspace solid-phase microextraction. J. Essent. Oil-Bear. Plants 19, 59-75. Nekoei, M., Mohammadhosseini, M., Akhlaghi, H., 2012. Chemical composition of the volatile oils from the aerial parts of Artemisia annua L. (Asteraceae) by using head space solid phase microextraction and hydrodistillation methods prior to gas chromatographicmass spectrometric determination: A comparative investigation. J. Essent. Oil-Bear. Plants 15, 926-933. Oshchepkova, Y.I., Rogozhin, E.A., Sultanova, E.M., Oripova, M.Z., Veshkurova, O.N., Egorov, T.A., Salikhov, S.I., 2012. Cationic peptides and proteins from seeds of plants of the family Malvaceae. Chem. Nat. Compd. 48, 288-290.

Peterson, R.T., McKinney, M., 1968. A Field Guide to Wildflowers of Northeast and North Central North America. Houghton Mifflin Co., Boston, USA.

Rahimi, M., Karimi, E., Nekoei, M., Mohammadhosseini, M., 2016. Hydro-distilled volatile oil constituents from the aerial parts of *Satureja mutica* and QSRR simulation by multiple linear regression. J. Essent. Oil-Bear. Plants 19, 307-320.

Razavi, S.M., Zarrini, G., Molavi, G., Ghasemi, G., 2011. Bioactivity of *Malva sylvestris* L., a medicinal plant from Iran. Iran J. Basic Med. Sci. 14, 574-579.

Romitelli, I., Martins, M.B.G., 2013. Comparison of leaf morphology and anatomy among *Malva sylvestris* ("gerânio-aromático"), *Pelargonium graveolens* ("falsamalva") and *Pelargonium odoratissimum* ("gerânio-decheiro"). Rev. Bras. Plantas Med. 15, 91-97.

Seyyednejad, S.M., Koochak, H., Darabpour, E., Motamedi, H., 2010. A survey on *Hibiscus rosasinensis*, *Alcea rosea* L. and *Malva neglecta* Wallr as antibacterial agents. Asian Pac. J. Trop. Med. 3, 351-355. Shahnama, M., Azami, S., Mohammadhosseini, M., 2015. Characterization of the essential oil and evaluation of antibacterial activity of methanolic extract of *Stachys lavandulifolia* Vahl. Int. J. Curr. Microbiol. App. Sci 4, 275-283.

Shale, T.L., Stirk, W.A., van Staden, J., 2004. Effect of storage on antibacterial and COX-1 anti-inflammatory activity of three plants used as traditional medicines in Lesotho. South Afr. J. Bot. 70, 602-610.

Shale, T.L., Stirk, W.A., van Staden, J., 2005. Variation in antibacterial and anti-inflammatory activity of different growth forms of *Malva parviflora* and evidence for synergism of the anti-inflammatory compounds. J. Ethnopharmacol. 96, 325-330.

Sleiman, N.H., Daher, C.F., 2009. *Malva sylvestris* water extract: A potential anti-inflammatory and antiulcerogenic remedy. Planta Med. 75, PH10.

Stef, D.S., Gergen, I., Trasca, T.I., Harmanescu, M., Lavinia, S., Ramona, B., Heghedus, M.G., 2009. Total antioxidant and radical scavenging capacities for different medicinal herbs. Rom. Biotech. Lett. 14, 4704-4709.

Tesevic, V., Vajs, V., Lekic, S., Dordevic, I., Novakovic, M., Vujisic, L., Todosijevic, M., 2012. Lipid composition and antioxidant activities of the seed oil from three Malvaceae species. Arch. Biol. Sci. 64, 221-227.

Tomoda, M., Gonda, R., Shimizu, N., Yamada, H., 1989. Plant mucilages .42. An anti-complementary mucilage from the leaves of *Malva-sylvestris* var *Mauritiana*. Chem. Pharm. Bull. 37, 3029-3032.

Turker, M., Dalar, A., 2013. *In vitro* antioxidant and enzyme inhibitory properties and phenolic composition of *M. neglecta* Wallr. (Malvaceae) fruit: A traditional medicinal fruit from Eastern Anatolia. Ind. Crops Prod. 51, 376-380.

Usami, A., Kashima, Y., Marumoto, S., Miyazawa, M., 2013. Characterization of aroma-active compounds in dry flower of *Malva sylvestris* L. by GC-MS-O analysis and OAV calculations. J. Oleo Sci. 62, 563-570. Vitullo, M., Ripabelli, G., Fanelli, I., Tamburro, M., Delfine, S., Sammarco, M.L., 2011. Microbiological and toxicological quality of dried herbs. Lett. Appl. Microbiol. 52, 573-580.



Walter, C., Shinwari, Z.K., Afzal, I., Malik, R.N., 2011. Antibacterial activity in herbal products used in Pakistan. Pakistan J. Bot. 43, 155-162. Wang, X., Bunkers, G.J., 2000. Potent heterologous antifungal proteins from cheeseweed (*Malva parviflora*). Biochem. Biophys. Res. Commun. 279, 669-673. Wang, X., Bunkers, G.J., Walters, M.R., Thoma, R.S., 2001. Purification and characterization of three antifungal proteins from cheeseweed (*Malva parviflora*). Biochem. Biophys. Res. Commun. 282, 1224-1228. Zargari, A., 1989. Medicinal Plants. University of Tehran Press, Tehran (In Persian).