

The Influences of a Dietary Phytobiotic "Yucca schidigera" Supplementation on Poultry: An Updated Article Review

Review Article

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ABSTRACT

Over the last decade, antimicrobials have been banned as growth promoters and replaced by other natural alternatives. Plant-based poultry feed additives and herbal extracts have been effectively used in poultry worldwide. One of these feed additive phytogenic substances is Yucca schidigera (YS), which showed various beneficial effects in livestock production, including poultry. This article provides a comprehensive understanding of YS's overall benefits. The various influences of YS supplementation on performance parameters of broilers, layers, and breeders, as well as carcass traits, intestinal health, immunity, and blood metabolites, and its properties as an antioxidant, anti-inflammatory, and anti-stress agent in poultry, were reviewed and updated. The dietary supplementation of YS in broilers could improve the feed intake, body weight, feed conversion ratio, as well as dressing and carcass percentage and weight. In layers and breeders, YS could enhance the egg production parameters, fertility, and hatchability, respectively. An enhanced intestinal integrity, a decreased pathogenic bacterial load, and an increased beneficial bacterial count were observed after adding YS to the diets of different poultry species. Moreover, YS showed boosting of the bird's humoral immune response and stimulation of the important liver and kidney metabolites. Besides, it acts as an effective antioxidant, anti-inflammatory, and anti-stress dietary supplement for poultry production systems. In conclusion, dietary YS is the best option for the overall poultry production parameters and can be used as a good natural growth-promoting antimicrobial alternative.

KEY WORDS

antioxidant, blood metabolites, immunity, intestinal health, performance, poultry, *Yucca schidigera*.

INTRODUCTION

Although antimicrobials have played a crucial role in preventing infection as well as promoting animal production, the food and drug safety organizations restricted their use in livestock production due to their adverse effects on the development of antimicrobial resistance, thereby posing a potential threat to human health (Saleh *et al.* 2020). Therefore, it is urgent to search for antimicrobial alternatives with no adverse health effects. Over the last few decades, plant-derived extracts, which are natural bioactive compounds, have been widely and effectively used as antibiotic

alternatives in livestock production and veterinary medicine (Lillehoj *et al.* 2018; Abd El-Ghany, 2024; Abd El-Ghany, 2025). Phytogenics may provide a viable alternative to conventional antimicrobials, contributing to improved sustainability and animal welfare in the poultry industry (Abd El-Ghany and Eraky, 2019; Abd El-Ghany, 2020a; Abd El-Ghany, 2020b; Abd El-Ghany, 2022; Abd El-Ghany and Babazadeh, 2022).

Yucca schidigera (YS) (Agavaceae), commonly known as Yucca, is a plant in the family Liliaceae native to southwestern United States and northern Mexico deserts as well as China (Cheeke et al. 2006; Kucukkurt et al. 2016). It has

been recognized by indigenous Indians as a source of food and drugs due to its health-promoting effects (Patel, 2012). Abd Elmoteleb et al. (2024) reported that YS is a safe, effective, biocompatible, and cost-efficient alternative natural product. The extracts and powder of YS have been widely used in the food industry as a natural additive and flavor enhancer, while in the livestock industries, YS is used as a natural antimicrobial feed additive (Cheeke, 2000; Alghirani et al. 2021; Abd Elmoteleb et al. 2024). In humans, the plant has been used to relieve joints' pain, stop bleeding and diarrhea, and treat skin ulcers (Patel, 2012). A variety of phytochemical substances, including steroid saponins, poly-phenols, glycol, and resveratrol, are extracted from YS (Oleszek et al. 2001; Piacente et al. 2005). The plant is known as one of the main sources of saponins. Steroidal saponins are accounted for approximately 10% of the dry weight of YS stem. Saponins are present in YS extracts in a steroidal form with strong antibacterial properties. Interestingly, the polyphenolic compounds existing in YS, such as resveratrol and a number of other stilbenes (Yucca oils A, B, C, D, and E), have antimicrobial, antioxiimmunomodulatory, anti-inflammatory, carcinogenic, and health-promoting activities (Cheeke et al. 2006; Ashour et al. 2014). Moreover, the use of saponins from both Quillaja saponaria and YS showed strong immune modulatory and gut health benefits, offering valuable alternatives to traditional antibiotics (Saddoris-Clemons et al. 2024). Yucca powder and juice are used to decrease the fecal odors, ammonia, hydrogen sulfide, and some other harmful volatile compounds in domestic animal excreta (Kelly and Kohler, 2003; Gaber, 2006; Pen et al. 2006; Liang et al. 2009; Chrenková et al. 2012). The promising properties of Yucca have led to several cutting-edge studies on its use in broiler chickens' nutrition.

Feed or water supplementation of YS powder or extracts is suitable and has many benefits for poultry (Table 1). The profit per bird and benefit-cost ratio were improved after water treatment of broilers with YS (Sariozkan et al. 2015; Patoary et al. 2020). Besides, the extracts of YS have been effectively used as growth promoters (Alghirani et al. 2021; Dai et al. 2023; Sun et al. 2023; Mert, 2025) and showed immuno-stimulatory (Ranjbar et al. 2014; Su et al. 2016; Sun et al. 2017; Dai et al. 2023; Mert, 2025), antioxidant (Alagawany et al. 2016; Alagawany et al. 2018; Farag et al. 2018; Mao et al. 2023; Abd Elmoteleb et al. 2024), antimicrobial (Matusiak et al. 2016; Dai et al. 2023; Abd Elmoteleb et al. 2024), anti-inflammatory (Cheeke et al. 2006; Oelschlager et al. 2019; Mao et al. 2023; Saddoris-Clemons et al. 2024), anti-protozoa (Alfaro et al. 2007; Oelschlager et al. 2019; Bafundo et al. 2020; Kozłowski et al. 2022; Mao et al. 2023; El-Sawah et al. 2025), and antistress properties (Luo et al. 2022). Besides, YS extracts have proved to improve many physiological functions, including lipid and protein metabolism (Gupta, 2014; Alagawany et al. 2016). YS could also be used to reduce ammonia emissions from poultry farms due to the presence of glycol (Tsukahara and Ushida, 2000; Cabuk et al. 2004; Chepete et al. 2012; Vlčková et al. 2017; Saeed et al. 2018; Dai et al. 2023; Munezero et al. 2023); thus, this might help in enhancing the performance parameters. Figure 1 shows the different influences of YS on the birds' bodies. The variable modes of action of YS in the birds' bodies are illustrated in Figure 2. It is noteworthy to mention that supplementation with Yucca not only enhances general health but also supports environmental sustainability in the modern poultry industry (Mert, 2025). In particular, the reduction in ammonia emission is a vital factor that improves both animal welfare and environmental impacts (Alfaro et al. 2007).

The current review article was thus planned to explore the various influences of YS supplementation on performance parameters of broilers, layers, and breeders, as well as carcass traits, intestinal health, immunity, and blood metabolites, and its properties as an antioxidant, antiinflammatory, and anti-stress agent in poultry.

Performance parameters

The productive performance parameters are a critical metric for assessing poultry production. The effective nutrient digestion and absorption are crucial for the growth promotion, productivity, and the general health of animals.

YS plays a significant role in promoting growth and production performance in the poultry industry sector (Malecki et al. 1995; Preston et al. 1999; Cheeke and Otero, 2005; Patel, 2012; Su et al. 2016; Saeed et al. 2018). Previous studies showed that dietary supplementation with YS increased the body weight (BW), ameliorated the average daily body weight gain (BWG), and improved the feed conversion ratio (FCR) of broilers (Alfaro et al. 2007; Ranjbar et al. 2014; Sahoo et al. 2015; Sun et al. 2017; Amirbeik et al. 2018; Awad et al. 2021; Bafundo et al. 2021; Dai et al. 2023) and egg parameters in layers (Dziuk et al. 1985; Kutlu et al. 2001; Kaya et al. 2003; Ayasan et al. 2005; Tugay et al. 2005; Wang and Kim, 2011; Alagawany et al. 2016; Alagawany et al. 2018; Khaskheli et al. 2020; Al-Fehaid et al. 2022). The early study of Baker (1997) reported that broilers treated with YS extract along with aluminium chloride showed improved live BW, while the feed intake (FI) was not affected. Obianwuna et al. (2024) demonstrated that mixtures containing YS increased live BWG in broiler chickens by up to 10%. Besides, Dai et al. (2023) showed that the different levels of Yucca spp. combinations have variable effects on BWG, and determining the optimal dosage is critical.

noculation rate	Bird spp.	Effects ¹	Reference
S at 63 ppm/kg diet	Broiler chickens	/ Improved BW	Johnston et al. (1981)
0 or 120 ppm YS S powder with a 10.76% saponin concentration	Broiler chickens	 ✓ Increased growth rate and Feed efficiency ✓ No impact on FI, egg yield, feed efficiency, and 	Lundeen (2000)
levels of 0, 30, 60, and 120 ppm	Layer chicken hens	quality indices of eggs	Kutlu et al. (2001)
ppm of YS powder	Laying quails	✓ No impact on feed efficiency and egg indices	Kaya et al. (2003)
mg/kg YS extracts	Broiler chickens	✓ Improved BW ✓ Reduce ammonia concentration in intestine	Cabuk et al. (2004)
% YS scidiostat-treated or vaccinated birds	Broiler chickens	✓ Improved average daily BWG and FCR ✓ Increasing the length of duodenal villi	Alfaro et al. (2007)
, 60, and 120 mg/Kg YS + 120 mg/kg caprylic d	Layer chicken hens	✓ Improved egg weight and feed efficiency ✓ Reduced serum and yolk cholesterol concentration ✓ Decreased proliferation of <i>E. coli</i>	Wang and Kim (2011)
0 ppm of YS extract in diets	Layer chicken hens	Decrease in ammonia generation and emission by 44% and 28% for the first two days of manure storage	Chepete et al. (2012)
ppm of YS extract in diets	Laying Japanese quail	✓ No effect on egg hatchability, total eggs set, and	Ayasan (2013)
5 mg/kg YS	Broiler chickens	fertile eggs % ✓ Improved growth and health performances	Ranjbar et al. (2014)
		✓ Improved BWG, FI, and FCR efficiency ✓ Reduced mortality ✓ Increased bursa of Fabricius and relative organs'	.,
0 mg/kg caprylic acid + 100 mg/kg YS extract	Broiler chickens	weights Increased white blood cell (lymphocyte) counts Reduced caecal <i>E. coli</i> count Improved meat quality	Begum et al. (2015)
5 mg/kg YS	Broiler chickens	✓ Improved meat quanty ✓ Improved growth and health performances	Sahoo et al. (2015)
extract up to 100 mg/kg diet	Layer chicken hens	 ✓ Enhanced egg number, egg mass, and eggshell 	Alagawany et al. (2016)
mg/kg of YS	,	thickness ✓ Improved feed efficiency	
mg/kg of YS	Broiler chickens	✓ Enhanced antioxidant and immune properties ✓ No significant effect on live BWG, FI, and feed	Su et al. (2016)
0, 200, and 300 mg/kg YS extract	Broiler chickens	utilization Increased catalase activity and gene expression Improvement of liver antioxidant capacity Improved growth performance Enhanced IgM production	Sun et al. (2017)
2% YS powder 1% YS powder	Broiler chickens	✓ Decreased low-density lipoprotein concentration in serum ✓ No effect on FI and carcass traits	Amirbeik et al. (2018)
ml YS extract/L drinking water	Broiler chickens	✓ Significant reduction in total aerobic colony count Insignificant increase in lactic acid bacteria Increased antioxidant enzyme and IgM and IgG levels	Ayoub et al. (2019)
L YS/1000 L drinking water	Broiler chickens	✓ Positive effects on fat oxidation parameters ✓ Non-significant difference in drip and cooking losses of breast muscle	Benamirouche et al. (2020)
0 mg YS/kg	Broiler chickens	✓ Increased antioxidant levels ✓ Decreased protein peroxidation ✓ Minimize the ammonia gas emission	Galli et al. (2020)
nl YS to 20 L drinking water	Broiler chickens	✓ Improve carcass quality ✓ Increase production performance	Patoary et al. (2020)
0 mg/kg YS extract	Broiler chickens	✓ Improved growth performance, ileal nutrient digesti- bility, gut health, carcass characteristics, and meat quality ✓ Improved egg number, mass, and weight, Haugh	Alghirani et al. (2021)
0 g/ton and 500 g/ton YS	Layer chicken hens	unit, shell percentage, shell thickness, and FCR Boosted cellular immune response	Al-Fehaid et al. (2022)
asal diets containing: 100 mg/kg feed of YS extract. 100 mg/kg feed of caprylic acid 100 mg/kg feed of YS extract + 500 mg/kg feed of 100 mg/kg feed of YS extract + 500 mg/kg feed of 1000 mg/kg feed of caprylic acid 1000 mg/kg feed of YS extract + 1000 mg/kg feed 1000 mg/kg feed of Caprylic acid 1000 mg/kg feed of YS extract + 2000 mg/kg feed 100 mg/kg feed of YS extract + 2000 mg/kg feed 100 mg/kg feed of YS extract + 2000 mg/kg feed 100 mg/kg feed of YS extract + 2000 mg/kg feed 100 mg/kg feed of YS extract + 2000 mg/kg feed	Layer chicken hens	✓ Improved BW, FI, FCR, and egg production and egg mass (YS and caprylic acid) ✓ No effect on egg weight, yolk weight, albumin weight, egg shell thickness, yolk index, shape index, and yolk color ✓ Increased concentration of serum glucose and T3 and T4 hormones ✓ Decreased serum total cholesterol concentrations and H/L ratio	El-Shafei et al. (2022)
0 g/T YS 5 g/T (anticoccidial)	Broiler chickens	✓ Improved BW and FCR ✓ Reduced Eimeria oocysts/gram count	Kozłowski et al. (2022)
, 1, and 1.5 ml YS extract/L drinking water	Broiler chickens	✓ Reduced Elmeria oocysts/gram count ✓ Increased intestinal Lactobacillus count	Rahman et al. (2023)
g/ton Quillaja Saponaria and YS	Broiler chickens	✓ No effects on BWG, FI, carcass yield, or blood biochemistry parameters	Alfredo (2023)
al diets with <i>Yucca</i> saponin (500 mg/kg), YS 0 mg/kg), and <i>Quillaja Saponaria</i> (500 mg/kg)	Broiler chickens	 ✓ Enhanced growth performance, antioxidant and anti- inflammatory activities, and immunity ✓ Improved intestinal health and ammonia metabolism 	Dai et al. (2023)
mg/kg YS in <i>C. perfringens</i> and <i>Eimeria</i> spp. llenged birds extract 0.02% and α-1,6-galactosidase and β-	Layer chicken hens	✓ Improved productive performance and egg quality ✓ Enhanced antioxidant capacity of the jejunum ✓ Improved nutrient digestibility, carcass metrics, and	Mao et al. (2023)
-mannanase multi-carbohydrase 0.1%, and low de protein	Broiler chickens	ammonia emission ✓ No effect on growth performance	Munezero et al. (2023)
al treatment of 5, 10, and 15 mg YS/day	28-day-old pigeons	 Improved serum biochemistry, immunoglobulin contents, and intestinal morphology by regulating the composition of microbial community in the ileum 	Sun et al. (2023)
0 and 300 mg YS extract/kg + 400 and 800 mg sil oil /kg	Broiler chickens	✓ Enhanced meat quality ✓ Improved <i>Lactobacillus</i> count ✓ Mitigated the negative effects of <i>Eimeria tenella</i>	Elmeligy et al. (2025)
0 mg YS extract and its nano-emulsion in the nking water	Broiler chickens	 ✓ Mitigated the negative effects of Eimeria tenella infection (reduced oocyst counts) ✓ Improved BWG and enhanced carcass quality 	El-Sawah et al. (2025)
umate (300 mg/kg), YS (100 mg/kg), and their	Broiler chickens	✓ Improved growth performance, intestinal microbiota,	Mert (2025)

BW: body weight; BWG: body weight gain; FI: feed intake; FCR: feed conversion ratio; *E. coli: Escherichia coli*; Ig: immunoglobulin; YS: *Yucca schidigera*; H/L: heterophils/lymphocytes; T3: tri-iodothyronine and T4: thyroxine.

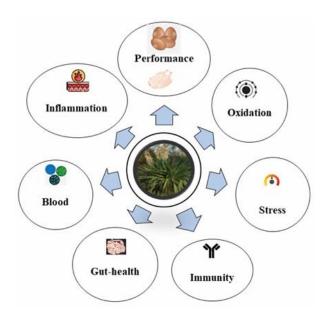


Figure 1 The impact of Yucca schidigera on body functions of the birds

It has been reported that supplementation of commercial laying flocks with YS induced a considerable favorable effect on the feed efficacy, performance productivity, finishing BW, and egg weight (Wang and Kim, 2011; Gurbuz et al. 2016). Dietary YS also showed positive influences on egg yield parameters and egg shell quality in laying Japanese quails (Ayasan et al. 2005). Similarly, Alagawany et al. (2018) found that adding YS extract to the feed of Japanese quails increased egg production, FI, and FCR. The results of the recent study of Mao et al. (2023) demonstrated that YS treatment of Clostridium perfringens (C. perfringens) and Eimeria spp. challenged layer hens induced an enhancement in the productive performance and egg quality, including increased egg weight, albumen height and weight, and yolk weight. In breeders, Enaiat et al. (2009) found that cocks of YS and YS with aluminium chloride treatments had recorded significantly high fertility %.

The growth-enhancing properties of YS are related to regulation of the intestinal microflora, improving nitrogen metabolism, digestion and absorption of nutrients, and intestinal health, as well as the anti-inflammatory, antioxidant, and immuno-modulatory capabilities (Bafundo *et al.* 2021). YS can ameliorate the intestinal morphology and increase the content of short-chain fatty acids (Dai *et al.* 2023). Supplementation with YS increased FI, most likely through reducing the circulation of cholecystokinin in the developing broilers under the stress of high ambient temperatures (Luo *et al.* 2022). Jun-Lin *et al.* (2016) demonstrated that *Yucca* supplementation decreased the intestinal pathogens, increased feed absorption, and improved FCR

by 4-6%.

In addition, the YS-treated broilers (87.26%) exhibited higher digestibility compared to the humate-treated group (86.46%), reflecting YS ability to enhance lipid absorption in the intestinal environment (Mert, 2025).

Generally, the favorable impact of phytobiotic feed additives such as YS is attributed to the presence of steroidal saponins, which contribute to the emulsification of oil fats, promotion of their digestion, and absorption of vitamins and minerals, thus improving intestinal health, decreasing microbial pressure, and preventing intestinal pathogens, perhaps leading to improved nutrient digestibility and absorption (Begum *et al.* 2015). Furthermore, steroidal saponins in YS have beneficial effects on the digestive tract through the activation of digestive enzymes, increasing the villus height and penetrability of intestinal mucosal cells, inhibiting the active mucosal transport, and facilitating the uptake of materials that are normally not absorbed (Su *et al.* 2016).

Additionally, supplementation with saponin extracts was found to reduce blood urea concentration, ammonia production, and odours from poultry droppings (Sahoo et al. 2015). Saponins are steroidal glycosides that have a strong surfactant activity which reduces the fluids' superficial tension and allows better nutrient absorption by the intestinal epithelium. Saponins can physically bind with ammonia and reduce its amount and emission (Das et al. 2012; Ayoub et al. 2019; Patoary et al. 2020). Besides, saponins increase the nutrients' absorption by decreasing the intestinal surface tension and activating the intestinal villi development, so they have a positive influence on performance indices (Jun-Ling et al. 2016). It has also been found that saponins may increase intestinal permeability and facilitate fat absorption (Cheeke, 2000), which may be attributed to an increased surface area of intestinal villi or modulation of microbial activity (Patoary et al. 2020).

The polyphenol compound present in YS also plays an important role due to its anti-inflammatory, antimicrobial, antioxidant, and free-radical-hunting characteristics, as well as its immune-enhancing properties, all of which enhance the growth performance of broilers (Su *et al.* 2016).

However, the performance parameters were not affected by adding YS to the diets of layers (Guclu, 2003; Kaya et al. 2003; Guclu and Iscan, 2004; Chepete et al. 2012; Ayasan, 2013; Sun et al. 2023) or broilers (Yeo and Kim, 1997; Cabuk et al. 2004; Begum et al. 2015). This variation may be due to the difference in the used breeds or species of poultry, the type of diet, and low levels of saponins (2.7%) and phenolic compounds (0.8%) that could not improve digestion, absorption, and nutrient utilization in the gut (Alagawany et al. 2016).

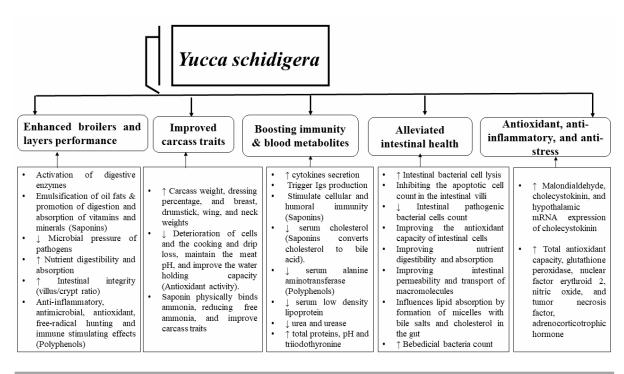


Figure 2 The modes of Yucca schidigera action in the birds body

Carcass traits

The effects of adding YS to the broiler diets (Sahoo et al. 2015; Munezero et al. 2023), drinking (Benamirouche et al. 2020; Patoary et al. 2020; Rahman et al. 2023), or spraying of litter (Guclu, 2003) on the carcass characteristics have been investigated. Sahoo et al. (2015) showed that dietary inoculation of YS increased the breast and thigh weights. Also, Rahman et al. (2023) demonstrated that broiler chickens that received YS in the drinking water showed significantly higher dressing percentage and carcass weight than control birds. Recently, the drinking water treatment of Eimeria-infected broilers with YS and nano-emulsion resulted in better meat cuts than the infected untreated group (El-Sawah et al. 2025). The inclusion of YS extract and multi-carbohydrase in low crude protein diets of broilers induced positive influences on the carcass parameters (Munezero et al. 2023). Adding 100 mg/kg of YS has proven to improve the carcass yield, especially the carcass weight, dressing percentage, and breast, drumstick, wing, and neck weights (Alghirani et al. 2021). In addition, YS treatment in broilers induced significantly higher dressing percentage and carcass weight, as well as improvement in meat pH, drip loss %, and cooking loss % (Benamirouche et al. 2020). The steroidal saponins of YS extract could physically bind ammonia, reduce the level of free ammonia, and play a significant role in the improvement of economic traits on broiler chickens (Nazeer et al. 2002; Patoary et al. 2020).

Moreover, saponins in YS demonstrate antioxidant activity to protect against the deterioration of cells, maintain the pH value of meat, decrease the cooking and drip loss, and improve the water holding capacity (Zdanowska-Zdanowska-Sąsiadek *et al.* 2019; Alghirani *et al.* 2021). Saleh *et al.* (2018) showed that chickens supplemented with 0.5, 1, and 1.5 kg/ton herbal mixture, including Yucca, resulted in a significant reduction in meat malondialdehyde (MDA). On the other hand, Begum *et al.* (2015) demonstrated that Ross chickens fed 100 mg/kg caprylic acid and 100 mg/kg YE showed no significant increase in muscle lightness compared to the control group.

Intestinal health

The reduction in the crypt depth of the intestinal mucosa reflects epithelial cells' maturation and promotes intestinal secretion (Chen and Yu, 2020). The increase in villus height-to-crypt depth ratio improves the mucosa structure and villus absorptive area, leading to enhanced intestinal digestion and absorption of nutrients (Chen and Yu, 2020). Yucca showed an intestinal villus structure-improving effect (Dengsheng et al. 2017). The addition of SY could significantly increase the intestinal villus height and the villus height-to-crypt depth ratio, reduce the crypt depth, and improve barrier function in the proximal intestine of chickens and pigeons (Alfaro et al. 2007; Alagawany et al. 2016; Bafundo et al. 2021; Sun et al. 2023). In the study of Alghirani et al. (2021), the villi height of the duodenum,

jejunum, and ileum was increased linearly with increasing levels of YS addition in the broiler chickens' diets during the starter and finisher phases. An increase in the villus height of the duodenum, jejunum, and ileum was observed in layer hens supplemented with YS extracts (Gurbuz et al. 2011). The ileal villus length was significantly improved by YS and quillaja dietary supplementation (Bafundo et al. 2021). Recently, Dai et al. (2023) found that YS increased the villus height, improved the villus height-to-crypt depth ratio, and consequently enhanced the intestinal integrity of broilers. Administration of YS could increase the activities of trypsin and lipase in the jejunal digesta, enhance the jejunal villus height-to-crypt depth ratio, and alleviate the increasing jejunal crypt depth after infection of laying hens with C. perfringens and Eimeria spp. (Mao et al. 2023). Bafundo et al. (2021) demonstrated that YS can reduce the intestinal harmful bacteria, regulate the intestinal flora, and improve the intestinal morphology. Furthermore, YS administration can mitigate the adverse effects of C. perfringens and Eimeria spp. challenge on histopathology and expression of tight-junction protein of jejunal mucosa, suggesting that YS enhanced gut barrier function (Mao et al. 2023).

The intestinal changes induced by YS are attributable to its influence on the intestinal microbial composition and mucosa, besides its ability to induce bacterial cell lysis. It has been reported that YS supplementation inhibited the apoptotic cell count in the jejunal villi of laying hens with *C. perfringens* and *Eimeria* spp. infection, thus reflecting the improved gut barrier function and protection of cell survival (Mao *et al.* 2023).

The dietary YS supplementation improved the antioxidant capacity of the proximal intestine of broilers (Alagawany et al. 2016). Similarly, Mao et al. (2023) reported that dietary addition of YS increased the total antioxidant capacity, decreased MDA level, and inhibited myeloperoxidase activity in the jejunal mucosa of *C. perfringens* and *Eimeria* spp. challenged layers. These results suggest that YS may relieve morphological damage and cell apoptosis caused by infection by improving the antioxidant capacity in the jejunal mucosa. Furthermore, YS can upregulate the expression of nuclear factor erythroid 2-related factor 2 and [NAD(P)H quinone dehydrogenase 1] in the jejunum of laying chickens infected by *C. perfringens* and *Eimeria* spp. (Mao et al. 2023).

Saponins, the major steroidal chemical elements, could ameliorate the intestinal absorption of nutrients (Sidhu and Oakenfull, 1986). Saponins of YS could improve intestinal health, decrease microbial pressure, and stabilize bowel health, but prevent intestinal disorder, which could lead to improved nutrient digestibility and absorption (Begum *et al.* 2015). Additionally, the intestinal presence of saponins

has been shown to influence the permeability and transport of macromolecules through the intestinal cells. The presence of saponins in YS influences lipid absorption by the formation of micelles with bile salts and cholesterol in the gut (Cheeke, 2000).

Broiler chickens supplemented by YS significantly induced higher energy and protein levels, indicating better energy utilization and protein digestibility (Sahoo et al. 2015; Sahoo et al. 2016; Alghirani et al. 2021). It has been found that the extract of YS in Arbor Acres chickens can positively affect the energy metabolism through the modulating hormone secretions and depressing energy compounds in the organism (Sun et al. 2017). Moreover, chickens fed 200 and 300 mg/kg YS extract showed improvement in the antioxidant capacity of liver (Sun et al. 2017). The short-chain fatty acids regulate the function of intestinal epithelial cells in order to improve the intestinal morphology and enhance the intestinal barrier and host metabolism (Martin-Gallausiaux et al. 2021). Increasing the intestinal content of short-chain fatty acids could affect ammonia emission by decreasing the fermentation intensity of animal protein in feed (Tsukahara and Ushida, 2000).

Gut flora has a significant role in influencing growth performance (Lan et al. 2020). Both the development of immunity and the preservation of the gut barrier are influenced by intestinal microbiota (Wang et al. 2023). Gut microflora interacts with the absorption of nutrients and the growth of the host's gastrointestinal tract, so it has a substantial effect on nutrition, health, and growth performance (Riyazi et al. 2015). YS can improve intestinal health by reducing the abundance of harmful intestinal flora. In YSsupplemented broiler chickens, the Lachnoclostridium concentration was significantly reduced, while Bacteroidota were significantly increased (Dai et al. 2023). The levels of tumor necrosis factor and interleukin (IL-) 6 are related to Lachnoclostridium concentration (Li et al. 2018). This indicated that YS can decrease the inflammation by reducing the Lachnoclostridium count in broilers. Moreover, a significant reduction of Desulfovibrio piger level was observed in YS-treated broiler chickens (Dai et al. 2023). Desulfovibrio spp. are sulfate-reducing bacteria that induce intestinal damage by decomposing sulfate and producing hydrogen sulfide. However, Ayoub et al. (2019) found that water treatment with YS resulted in a significant reduction in total aerobic colony count and an insignificant increase in lactic acid bacteria in broiler chickens. Similarly, Wang and Kim (2011) demonstrated that the addition of YS to the diets of laying hens did not affect Lactobacillus count.

According to its saponins content, Yucca exhibits ammonia-binding, urease activity-inhibiting, intestinal epithelial cell surface tension-reducing, antiprotozoal, antibacterial, antifungal, and antioxidant properties. Saponins show wa-

ter-and oil-soluble properties; therefore, they play an important role in the formation of micelles during fat absorption and also facilitate the passage of other nutrients through the intestinal epithelium (Franchis et al. 2002). In addition, this substance binds to the cell membranes of the intestinal pathogenic microorganisms, causing their death and exhibiting antibacterial effects (McAllister et al. 2001). Besides, saponins improve the nutrients' absorption via reducing the intestinal surface tension and triggering the intestinal villi development, thus having a positive influence on performance (Su et al. 2016). The study of Sun et al. (2023) showed that the oral supplementation of young pigeons with YS could improve the intestinal morphology by regulating the intestinal microbial community. The oral supplementation of YS increased the abundance of Patescibacteria and Desulfobacterota phyla as well as Sulfurospirillum, Solobacterium, Desulfovibrio, Desulfobulbus, Lactococcus, Parabacteroides, Acidaminococcus, and Acetobacter genera. Desulfococcus and Desulfovibrio are the main contents of sulfate-reducing bacteria, and Desulfovibrio have many enzymes acting together to resist oxidative stress (Dolla et al. 2006). In addition, the reduction of serum cholesterol levels following supplementation with YS may be controlled by the regulation of the intestinal Actinomyces genus abundance. Actinobacteria can induce secondary metabolites with structural diversity and are also the source of two-thirds of natural antibiotics and a series of anti-cancer, anti-parasitic, antifungal, and immunosuppressive drugs (Demain and Sanchez, 2009). The working principles of Yucca were explained in the study of Alghirani et al. (2021), who found that it decreased the microbial pressure by reducing the pathogenic bacteria (e.g., Enterococcus faecalis) and enhanced the growth of beneficial bacteria (e.g., Lactococcus, Streptococcus, and Parabacteroides).

These changes can alter beta diversity, enrich the microbial composition of gut, and decrease the risk of diarrhea by binding toxins, such as ammonia. Similarly, 5 to 10% of YS extract was sufficient to inhibit the growth of Enterococcus faecalis and Enterococcus avium in the manure of poultry (Matusiak et al. 2016). Hassan et al. (2010) demonstrated that quillaja and Yucca saponins may affect the activity of the intestinal Salmonella enterica ser. Typhimurium, Escherichia coli (E. coli), and Staphylococcus aureus, as well as other Gram-positive organisms. YS could be used for the treatment of salmonellosis by enhancing immune responses and activating antioxidative capacity (Abd Elmoteleb et al. 2024). Also, the study of Mert (2025) demonstrated a decrease in E. coli count, which suggests that the humate-Yucca combination may enhance economic efficiency by promoting intestinal health.

An increase in the digestion of essential minerals, improvement in the intestinal health, and reduction in the

growth of pathogenic bacteria in broilers have been reported (Ebru *et al.* 2013; Sun *et al.* 2019). The presence of antimicrobial properties of YS is due to the high content of saponins (Matusiak *et al.* 2016). Steroidal saponins are poorly absorbed from the gastrointestinal tract, hence altering the microbial fermentation in the gut. Besides, Yucca supplementation could inhibit the proliferation and growth of pathogenic bacteria by reducing the intestinal ammonia levels (Alfaro *et al.* 2007).

The anticoccidial effect of YS in chickens has been reported in numerous studies (Trejo Castro, 2002; Oviedo et al. 2003; Alfaro et al. 2007; Galli et al. 2018; Oelschlager et al. 2019; Bafundo et al. 2020; Kozłowski et al. 2022; El-Sawah et al. 2025). It has been suggested that YS can be safely used as a coccidiostat for higher economical return and reducing the costs associated with coccidiosis problems in broiler farms (Kozłowski et al. 2022). Yucca-based saponins also displayed coccidiostat actions as measured by reduced oocyte production and protection against intestinal damage induced by the Eimeria in broilers (Galli et al. 2018). Mao et al. (2023) reported that dietary supplementation of YS extract improved digestion and absorption and promoted growth with a reduction of the negative effects of coccidiosis. A combination of quillaja and Yucca saponins exhibited anticoccidial effects in growing broilers (Bafundo et al. 2021; Saddoris-Clemons et al. 2024). The treatment of Eimeria-infected broiler chickens with a mixture of YS and Ouillaja saponaria produced positive influences on growth performance, intestinal health, and immunity during both the peak and recovery phases following a coccidia infection (Saddoris-Clemons et al. 2024). A combination of coccidiostats (monensin and salinomycin) plus YS increased the intestinal crypt depth and reduced the villus-tocrypt ratio in broiler chickens (Alfaro et al. 2007). Trejo Castro (2002) demonstrated that Eimeria oocysts count and the gut lesion scores were reduced in chickens treated with YS compared to narasin, salinomycin, monensin, and nicarbazin ionophores. Moreover, YS in combination with monensin and salinomycin (anticoccidial), Stafac (antiinfective), and bacitracin methylene disalicylate was very effective in improving the growth and FCR and reducing oocyst index in chickens infected with Eimeria spp. (Oviedo et al. 2003). Similar results were obtained by Alfaro et al. (2007), who demonstrated a potent synergistic coccidiostat influence of both anti-coccidia vaccine and YS when compared to monensin and salinomycin. Recently, feed treatment of broiler chickens with YS resulted in improving the FCR and average daily gain and diminishing the oocyst per gram counts compared to the anticoccidial and challenged groups (Kozłowski et al. 2022). So, the anticoccidial effect of YS may be attributed to reducing oocyst count and enhancing the body performance of the

treated broilers (Kozłowski *et al.* 2022). The extract of YS is rich in steroidal saponins that interact with cholesterol in the protozoan cell membrane during the different developmental stages and consequently prevent the parasites' development and destroy them by lysis (Wang *et al.* 1998; Cheeke *et al.* 2006; Bafundo *et al.* 2020; Kozlowski *et al.* 2022).

The previous findings vary from those of Oelschlager *et al.* (2019), who demonstrated a reduction in interleukin (IL-1 β) with no impact on interferon- γ or IL-12 β in the duodenum at 7 days after Eimeria infection in chickens fed 250 to 500 mg/kg of a Yucca-derived saponin. This discrepancy may probably be explained by the differences in sources, quality, and composition of the tested YS and consequently the amount of steroidal saponins that are responsible for the antiprotozoal effect and the effect on the digestion process.

Immunity

YS has stimulating, inhibiting, or regulating immune effects (Dai et al. 2023) as its contents of steroidal saponin extracts can enhance cytokine secretion, trigger immunoglobulin (Ig) production, and stimulate both cellular and humoral immune responses (Song and Hu, 2009). Su et al. (2016) demonstrated that the inclusion of YS powder in broiler ration (100 or 200 mg/kg) stimulated cellular and humoral immune responses. Moreover, dietary YS can linearly increase the IgG production in the serum of laying hens (Alagawany et al. 2016) and broilers (Ayoub et al. 2019). In the study of Dai et al. (2023), high levels of IgA and IgY were detected in the serum of YS-treated broiler chickens. A high concentration of IgM was observed in 35-day-old broiler chickens treated with YS and inoculated with sheep red blood cells (Amirbeik et al. 2018). Moreover, the total protein and IgM contents in the serum were significantly higher in young pigeons treated with a dietary YS than in birds in the control non-treated group (Sun et al. 2023). The study of Mert (2025) indicated that humate and Yucca supplementation are efficient in enhancing the production of IgA and IgM levels both alone and in combination. Also, Bafundo et al. (2021) found that the combination of Yucca and Quillaja augmented intestinal health by increasing IgA and IgM levels. Increasing the intestinal lactic acid bacterial count can improve nutrient absorption and immunological response by establishing the intestinal barrier function (Mert, 2025). Alghirani et al. (2021) reported the effect of Yucca on the host's immunity as follows: it promotes the humoral immunity by elevating the serum IgM and total protein levels. Through polyphenols, Yucca exhibits antiinflammatory effects, improves antioxidant status, and reduces liver enzymes (e.g., alanine aminotransferase). By strengthening gut morphology (increasing villus height and reducing crypt depth), Yucca indirectly augments the immune barrier. YS can also boost the immune response by elevating lysozyme levels and hemagglutination inhibition antibody titer in Salmonella-infected and YS-treated broilers (Abd Elmoteleb *et al.* 2024). On the other hand, the study of Sun *et al.* (2023) demonstrated that the weight of immune organs, including the thymus glands, bursa of Fabricius, and spleen, was not affected following supplementation of young pigeons with a dietary YS.

Blood metabolites

The liver enzymes, such as serum alanine aminotransferase level, in YS-supplemented young pigeons were significantly lower than those in the control non-treated group (Sun et al. 2023). The low level of liver enzymes may be due to the richness of YS in phenolic components (Piacente et al. 2004; Bidinotto et al. 2011). The serum low-density lipoprotein level was the lowest in broiler chickens treated with 0.015% YS powder when compared with other groups (Amirbeik et al. 2018). The concentration of serum total cholesterol was significantly decreased in YS-treated layers and broilers when compared with control group (Kaya et al. 2003; Bafundo et al. 2021; Sun et al. 2023). Saponins with cholesterol form water-insoluble complexes which accelerate the liver conversion of cholesterol to bile acid, and consequently, the level of cholesterol in the serum was decreased (Sidhu and Oakenfull, 1986). The dietary supplementation of YS extract reduced urea, urease, and creatinine concentrations, maintained albumin and globulin at normal levels, and enhanced total proteins and triiodothyronine (T3) concentrations in the blood of broiler chickens (Baker, 1997; Abd Elmoteleb et al. 2024). In the study of El-Sawah et al. (2025), the results indicated that both forms of YS and its nano-emulsion were safe for the liver and kidney in terms of normal liver and kidney function tests levels. A parallel finding of Abd Elmoteleb et al. (2024) revealed that YS treatment may boost liver antioxidant activity and modify renal function owing to its antioxidantactive components, which include steroidal saponins, flavonoids, and polyphenols. Moreover, Yucca can reduce the blood urea levels by inhibiting its formation in the liver and increasing urea elimination (Abd Elmoteleb et al. 2024). In addition, the levels of T3 and thyroxine (T4) hormones were significantly increased in the humate and YSsupplemented broilers compared to the control group (Mert,

These findings suggest that humate and Yucca supplementation support metabolic processes and increase thyroid hormone production. Similar results were reported by Obianwuna *et al.* (2024), who found that *Yucca*-containing mixtures increased T3 and T4 levels by 10% and that this effect was associated with the relationship between gut health and metabolic efficiency.

Antioxidant properties

Previous studies showed the antioxidant properties of YS in poultry (Alagawany et al. 2016; Farag et al. 2018; Awad et al. 2021). The serum contents of MDA were decreased, while the total antioxidant capacity and glutathione peroxidase activity were increased in broilers supplemented with dietary YS (Dai et al. 2023). The reduction of MDA level may be due to the capability of YS to eliminate secondary reactive radicals or inhibit superoxide and hydrogen peroxide production (Enginar et al. 2006). Nearly similar results were obtained by Luo et al. (2022), who demonstrated a reduction in the plasma lipid oxidation product MDA, cholecystokinin, and hypothalamic mRNA expression of cholecystokinin, without an effect on the activities of superoxide oxidase and glutathione peroxidase in YS-treated broilers under high ambient temperature. Moreover, dietary addition of YS upregulated the expression of nuclear factor erythroid 2-related factor 2 and [NAD(P)H quinone dehydrogenase 1] in the jejunal mucosa of laying hens infected by C. perfringens and Eimeria spp. (Mao et al. 2023). The nuclear factor erythroid 2-related factor 2 signaling pathway plays a key role in regulating antioxidant enzyme functions; therefore, a lack of this factor induces a dysfunction of the antioxidant system (Cheung et al. 2014).

Supplementation with YS reduced the lipid-oxidative indicators, indicating its antioxidant capacity (Luo et al. 2022). It acts by enhancing the nuclear factor erythroid 2-related factor 2 signaling pathway (Dai et al. 2023), which is an antioxidant pathway (Wang et al. 2020), and increases glutathione peroxidase production (Gao et al. 2016). Dietary saponins from YS extracts at a level 100 mg/kg improved the antioxidant capacity in the small intestine and increased the gene expression of superoxide dismutase, glutathione peroxidase, and catalase (Sun et al. 2019). The protopanaxadiol saponin fraction may reduce the release of some inflammatory cytokines, including nitric oxide and tumor necrosis factor, blocking p38, c-Jun N-terminal kinase, and other proteins through downregulating mRNA gene expression (Yang et al. 2015).

Anti-inflammatory properties

The whole-plant Yucca powder has shown powerful antiinflammatory activity, mediated via inhibition of nuclear factor kappa B (NF-κB) activation (Cheeke *et al.* 2006). The dietary YS increased the total antioxidant capacity, decreased MDA level, and inhibited myeloperoxidase activity in the jejunal mucosa of *C. perfringens* and *Eimeria* spp. challenged layers (Mao *et al.* 2023). The myeloperoxidase is mostly expressed in inflammatory cells (particularly neutrophils) and is known to be significant in tissue damage and inflammation in different inflammatory disease conditions (Aratani, 2018). Yucca saponins can

decrease cytokine concentrations associated with inflammatory responses (Oelschlager et al. 2019; Abd Elmoteleb et al. 2024). The steroid saponin molecules in YS have a similar structure to glucocorticoid; thus, these molecules exert a glucocorticoid-like effect through glucocorticoid receptors, e.g., antioxidant (Su et al. 2016; Farag et al. 2018) and antiinflammatory properties (Cheeke et al. 2006; Dai et al. 2023). Additionally, polyphenols in Yucca can reduce the reactive oxygen species and interrupt the inflammatory response by scavenging free radicals in cells (Dai et al. 2023). The activated macrophages generate IL-6, a proinflammatory cytokine that initiates and worsens the inflammation course (Abdel-Tawwab et al. 2021). Yucca bark and whole Yucca plant powder contain resveratrol, which is known for its anti-inflammatory activity (Bhat and Pezzuto, 2002). The IL-6 mRNA gene expression is related to high resveratrol (polyphenol) levels in Yucca. Resveratrol may block the NF-κB pathway, resulting in a decrease in IL-6 transcription (Attallah et al. 2022).

The reduced plasma level of fluorescein isothiocyanate dextran in Eimeria-challenged-YS treated chickens pointed to the protective effect of the plant on the intestinal barrier and a reduction in leakiness that is often a result of inflammation-induced damage (Saddoris-Clemons *et al.* 2024). This suggested that feeding YS may help to diminish the acute inflammatory response, potentially limiting the tissue damage associated with excessive inflammation due to coccidiosis (Saddoris-Clemons *et al.* 2024).

Anti-stress properties

YS could alleviate the heat stress and improve the general health conditions of broiler chickens (Rezaei et al. 2017; Luo et al. 2022). Supplementation with YS alleviated the stress of high temperature and accordingly reduced the rectal and leg skin temperature (Luo et al. 2022). Yucca may increase the antioxidant status and alleviate the heat stress reaction in developing broilers via controlling gene expression that senses body temperature in the hypothalamus (Luo et al. 2022). Saponin component of YS has an antidepressant effect in animals (Abbas et al. 2014). For instance, intraperitoneal injection of mice with ginseng saponin (5 or 20 mg/kg BW) reduced the adrenocorticotropic hormone, which induces stress (Kim et al. 2003). In the study of Luo et al. (2022), the high temperature reduced the hypothalamic mRNA expression of transient receptor potential melastatin type 8, while treatment with YS increased the mRNA expression of both heat (transient receptor potential vanilloid receptors 1 and 4) and cool (transient receptor potential melastatin type 8) sensing receptors. These results indicate the increased YS thermoregulation capacity under heat stress, thereby attenuating the increased body temperature induced by high temperature. Moreover,

YS prevented the reduced FI of broilers exposed to heat stress conditions (Luo *et al.* 2022), which may be due to the antioxidant capacity of YS, especially in the hypothalamus (Alagawany *et al.* 2018; Luo *et al.* 2018).

CONCLUSION

The findings of this article highlight the potential of plant-based feed additives as sustainable alternatives to conventional growth promoters in poultry production systems. YS is a very suitable feed additive that has the ability to ameliorate the general health of poultry. It has advantages for improving the performance of broilers and layers, carcass traits, intestinal health, immunity, and some blood parameters and reducing serum oxidant, inflammatory, and stress factors in different poultry species. Therefore, YS could be safely used in poultry production systems for higher economical return without any adversity. A continued investigation into the use of natural feed additives for poultry will help in the development of effective and sustainable strategies for optimizing the health and production.

REFERENCES

- Abbas G., Rauf K. and Mahmood W. (2014). Saponins: The phytochemical with an emerging potential for curing clinical depression. *Nat. Prod. Res.* **29(4)**, 302-307.
- Abd El-Ghany W.A. (2020a). A review on the use of *Azolla* species in poultry production. *J. World's Poult. Res.* **10(2)**, 378-384.
- Abd El-Ghany W.A. (2020b). Microalgae in poultry field: A comprehensive perspectives. *Adv. Anim. Vet. Sci.* **8(9)**, 888-897.
- Abd El-Ghany W.A. (2022). The potential uses of silymarin, a milk thistle (*Silybum marianum*) derivative, in poultry production system. *Online J. Anim. Feed Res.* **12(1),** 46-52.
- Abd El-Ghany W.A. (2024). The influence of the phytobiotic tulsi (*Ocimum sanctum*) on broiler production and health. *Vet. Stanica.* **55(2)**, 221-230.
- Abd El-Ghany W.A. (2025). The different influences of a phytobiotic, green tea (*Camellia sinensis* L.) on the poultry health and production. *J. Appl. Vet. Sci.* **10(1)**, 32-45.
- Abd El-Ghany W.A. and Babazadeh D. (2022). Betaine: A potential nutritional metabolite in the poultry industry. *Animals*. **12(19)**, 2624-2635.
- Abd El-Ghany W.A. and Eraky R.D. (2019). Influence of dietary *Moringa oleifera* on broilers performance, intestinal microbial population and humoral immune competence. *J. Hellenic Vet. Med. Soci.* **70(4)**, 1805-1810.
- Abd Elmoteleb A.M., Hafez A.S. and Ahmed H.M. (2024). Effect of *Yucca Schidigera* extract on *Salmonella enteritidis*-infected poultry compared with fosfomycin drug. *Egyptian J. Vet. Sci.* 1, 1-10.
- Abdel-Tawwab M., Mounes H.A., Shady S.H. and Ahmed K.M. (2021). Effects of yucca, *Yucca schidigera* extract and/or yeast *Saccharomyces cerevisiae* as water additives on growth, bio-

- chemical and antioxidants/oxidant biomarkers of Niletilapia, *Oreochromis niloticus*. *Aquaculture*. **533**, 736122-736129.
- Alagawany M., Abd El-Hack M.E. and El-Kholy M.S. (2016). Productive performance, egg quality, blood constituents, immune functions, and antioxidant parameters in laying hens fed diets with different levels of *Yucca schidigera* extract. *Environ. Sci. Pollut. Res.* 23(7), 6774-6782.
- Alagawany M., Abd El-Hack M.E., Farag M.R., Elnesr S.S., El-Kholy M.S., Saadeldin I.M. and Swelum, A.A. (2018). Dietary supplementation of *Yucca schidigera* extract enhances productive and reproductive performances, blood profile, immune function, and antioxidant status in laying Japanese quails exposed to lead in the diet. *Poult. Sci.* 97(9), 3126-3137.
- Alfaro D.M., Silva A.V.F., Borges S.A., Maiorka F.A., Vargas S. and Santin E. (2007). Use of *Yucca schidigera* extract in broiler diets and its effects on performance results obtained with different coccidiosis control methods. *J. Appl. Poult. Res.* **16(2)**, 248-254.
- Alfredo J. (2023). *Quillaja saponaria* and *Yucca schidigera*: Growth performance and nutrient digestibility in broilers. *Vitam. Miner.* **12**, 256-266.
- Al-Fehaid M.F., Fathi M.M., Al-Homidan I.H. and Abou-Emera O.K. (2022). Effect of yucca (*Yucca Shidigera*) extract supplementation on egg production, egg quality and immune response of laying hens. *Egyptian Poult. Sci.* 42(4), 483-500.
- Alghirani M.M., Chung E.L.T., Sabri D.S.M., Tahir M.N.J.M., Kassim N.A., Kamalludin M.H., Nayan N., Jesse F.F.A., Sazili, A.Q. and Loh T.C. (2021). Can *Yucca schidigera* be used to enhance the growth performance, nutrient digestibility, gut histomorphology, cecal microflora, carcass characteristic, and meat quality of commercial broilers raised under tropical conditions? *Animals*. 11(8), 2276-2285.
- Amirbeik E., Hassanabadi A. and Nassiri Moghaddam H. (2018). The effect of different dietary levels of *Yucca schidigera* extract powder on performance, blood metabolites, immune system and carcass traits of broiler chickens. *Iranian J. Anim. Sci. Res.* **10(1)**, 69-80.
- Aratani Y. (2018). Myeloperoxidase: its role for host defense, inflammation, and neutrophil function. *Arch. Biochem. Bio-phys.* **640**, 47-52.
- Ashour E.A., Alagawany M., Reda F.M. and Abd El-Hac M.E. (2014). Effect of supplementation of *Yucca schidigera* extract to growing rabbit diets on growth performance, carcass characteristics, serum biochemistry and liver oxidative status. *Asian J. Anim. Vet. Adv.* **9(11)**, 732-742.
- Attallah N.G., El-Sherbeni S.A., El-Kadem A.H., Elekhnawy E., El-Masry T.A., Elmongy E.I., Altwaijry N. and Negm W.A. (2022). Elucidation of the metabolite profile of *Yucca gigantea* and assessment of its cytotoxic, antimicrobial, and anti-inflammatory activities. *Molecules*. **27(4)**, 1329-1335.
- Awad A.L., Fahim H.N. and El-shhat A.G. (2021). Effect of adding *Yucca schidigera* powder to domyati ducklings diets on some productive characteristics and economic efficiency during summer season. *Egyptian Poult. Sci. J.* **41(2)**, 265-278.
- Ayasan T. (2013). Effects of dietary *Yucca schidigera* on hatchability of Japanese quails. *Indian J. Anim. Sci.* **83(6)**, 641-644. Ayasan T., Yurtseven S., Baylan M. and Canogullari S. (2005).

- The effects of dietary *Yucca shidigera* on egg yield parameters and egg shell quality of laying Japanese quails (*Coturnix coturnix* japonica). *Int. J. Poult. Sci.* **4(3)**, 159-162.
- Ayoub M.M., Ahmed H.A., Sadek K.M., Alagawany M., Abd El-Hack M.E., Othman S.I., Allam A.A. and Abdel-Latif M.A. (2019). Effects of liquid yucca supplementation on nitrogen excretion, intestinal bacteria, biochemical and performance parameters in broilers. *Animals.* 9(12), 1097-1103.
- Bafundo K.W., Johnson A.B. and Mathis G.F. (2020). The effects of a combination of *Quillaja saponaria* and *Yucca schidigera* on *Eimeria* Spp. in broiler chickens. *Avian Dis.* **64(3)**, 300-304.
- Bafundo K.W., Männer K. and Duerr I. (2021). The combination of quillaja and yucca saponins in broilers: Effects on performance, nutrient digestibility and ileal morphometrics. *Br. Poult. Sci.* **62(4)**, 589-595.
- Baker D.H. (1997). Ideal amino acid profiles for swine and poultry and their applications in feed formulation. *BioKyowa Tech. Rev.* **9,** 1-19.
- Begum M., Hossain M.M. and Kim I.H. (2015). Effects of caprylic acid and *Yucca schidigera* extract on growth performance, relative organ weight, breast meat quality, haematological characteristics and caecal microbial shedding in mixed sex Ross 308 broiler chickens. *Vet. Med. Czech Acad. Agric. Sci.* **60(11),** 635-643.
- Benamirouche K., Baazize-Ammi D., Hezil N., Djezzar R., Niar A. and Guetarni D. (2020). Effect of probiotics and *Yucca schidigera* extract supplementation on broiler meat quality. *Acta Sci. Anim. Sci.* **42**, 48066-48076.
- Bhat K.P. and Pezzuto J.M. (2002). Cancer chemopreventive activity of resveratrol. *Ann. NY. Acad. Sci.* **957(1)**, 210-229.
- Bidinotto L.T., Costa C.A., Salvadori D.M., Costa M., Rodrigues M.A. and Barbisan L.F. (2011). Protective effects of lemongrass (*Cymbopogon citratus* STAPF) essential oil on DNA damage and carcinogenesis in female Balb/C mice. *J. Appl. Toxicol.* **31(6)**, 536-544.
- Cabuk M., Alcicek A., Bozkurt M. and Akkan S. (2004). Effect of *Yucca schidigera* and natural zeolite on broiler performance. *J. Poult. Sci. Int.* **3(10)**, 651-654.
- Cheeke P.R. (2000). Actual and potential applications of *Yucca schidigera* and *Quillaja saponaria* saponins in human and animal nutrition. *J. Anim. Sci.* 77, 1-10.
- Cheeke P.R. and Otero R. (2005). Yucca, quillaja may have role in animal nutrition. *Feedstuffs*. **77**, 11-14.
- Cheeke P.R., Piacente S. and Oleszek W. (2006). Antiinflammatory and anti-arthritic effects of *Yucca schidigera*: A review. *J. Inflamm.* **3,** 6-15.
- Chen D. and Yu B. (2020). Animal Nutrition. China Agriculture Press, Beijing, China.
- Chepete H.J., Xin H., Mendes L.B., Li H. and Bailey T.B. (2012). Ammonia emission and performance of laying hens as affected by different dosages of *Yucca schidigera* in the diet. *J. Appl. Poult. Res.* **21(3)**, 522-530.
- Cheung K.L., Lee J.H., Khor T.O., Wu T.Y., Li G.X., Chan J., Yang C.S. and Kong A.N.T. (2014). Nrf2 knockout enhances intestinal tumorigenesis in Apc (min/+) mice due to attenuation of anti-oxidative stress pathway while potentiates in

- flammation. Mol. Carcinog. 53(1), 77-84.
- Chrenková M., Chrenková M., Poláĉiková M., Formelová Z., Baláži A., Ondruška Ľ., Sirotkin A. and Chrenek P. (2012). The effect of *Yucca schidigera* extract in diet of rabbits on nutrient digestibility and qualitative parameters in caecum. *Slovak. J. Anim. Sci.* 45(3), 83-88.
- Dai Z., Wang H., Liu J., Zhang H., Li Q., Yu X., Zhang R. and Yang C. (2023). Comparison of the Effects of Yucca saponin, Yucca schidigera, and Quillaja saponaria on growth performance, immunity, antioxidant capability, and intestinal flora in broilers. Animals. 13(9), 1447-1456.
- Das T.K., Banerjee D., Chakraborty D., Pakhira M.C., Shrivastava B. and Kuhad C. (2012). Saponin: Role in animal system. *Vet. World.* **5(4)**, 248-254.
- Demain A.L. and Sanchez S. (2009). Microbial drug discovery: 80 years of progress. *J. Antibiotics (Tokyo)*. **62(1)**, 5-16.
- Dengsheng S., Xiao J., Binlin S., Junling S., Manman T. and Sumei Y. (2017). Dietary *Yucca schidigera* extract improved growth performance and liver antioxidative function in broilers. *Italian J. Anim. Sci.* **16(4)**, 677-684.
- Dolla A., Fournier M. and Dermoun Z. (2006). Oxygen defense in sulfate-reducing bacteria. *J. Biotechnol.* **126(1)**, 87-100.
- Dziuk H.E., Duke G.E., Buck R.J. and Janni K.A. (1985). Digestive parameters in young turkeys fed yucca saponin. *Poult. Sci.* **64(6)**, 1143-1147.
- Ebru O.E., Erdem E., Afsinya N.U. and Torlak E. (2013). Effect of *Yucca schidigera* spraying in different litter materials on some litter traits and breast burn of broilers at the fifth week of production. *Kaf. Univ. Vet. Fak. Derg.* **19(5)**, 749-753.
- Elmeligy E.H., Attallah S.T., Sallam E.A. and Mohammed L.S. (2025). Impact of Yucca extract and basil oil supplementation on carcass characteristics, quality of meat, and the cecal microbiota in broiler chickens. *J. Adv. Vet. Res.* **15(1)**, 20-27.
- El-Sawah A.A., Aboelhadid S.M., El-Nahass E.N. and Helal H.E. (2025). Efficacy of nanoemulsion of *Yucca shidigera* extract in controlling *Eimeria tenella* infection in broilers. *Assiut Vet. Med. J.* **71(185)**, 65-82.
- EL-Shafei A.A., AL-Gamal M.A. and Shams El-deen A.E. (2022). Effectiveness of caprylic acid and *Yucca schidigera* extract on productive and physiological performance of laying hens. *Egyptian J. Nutr. Feeds.* **25(1),** 67-83.
- Enaiat M.M.E., Eman M., Abou-Eitta A., Salem A. and Al-Kotait A.H.A. (2009). Effect of *Yucca schidigera* extract and aluminum chloride on pen's atmospheric ammonia, productive, reproductive and physiology performance of silver montazah cocks. *Egyptian J. Poult. Sci.* **29**, 337-356.
- Enginar H., Avci G., Eryavuz A., Kaya E., Kucukkurt I. and Fidan A.F. (2006). Effect of *Yucca schidigera* extract on lipid peroxidation and antioxidant activity in rabbits exposed to γ radiation. *Rev. Méd. Vét.* **157(8)**, 415-419.
- Farag M.R., Alagawany M., EI-Hack M.E.A., EI-Sayed S.A.A., Ahmed S.Y.A. and Samak D.H. (2018). *Yucca schidigera* extract modulates the lead-induced oxidative damage, nephropathy and altered inflammatory response and glucose homeostasis in Japanese quails. *Ecotoxicol. Environ. Saf.* **156**, 311-321.
- Franchis G., Kerem Z., Makkar H.P.S. and Becker K. (2002). The biological action of saponins in animal systems: A review. *Br.*

- J. Nutr. 88(6), 587-605.
- Gaber M.M. (2006). The effects of plant-protein based diets supplemented with Yucca on growth, digestibility, and chemical composition of Nile Tilapia (*Oreochromis niloticus*) ingerlings. *J. World Aquac. Soci.* **37(1)**, 74-81.
- Galli G.M., Da Silva A.S., Bottari N.B., Biazus A.H., Petrolli T., Reis J.H., Morsch V.M., Schetinger M.R.C., Piva M.M., Baggio R.A., Mendes R.E., Boiago M.M., Stefani L.M. and Machado G. (2018). Addition of yucca extract and glutamine in the diet of chicks had a protective effect against coccidiosis. *Comp. Clin. Pathol.* 27(1), 205-214.
- Galli G.M., Griss L.G., Boiago M.M., Petrolli T.G., Glombowsky P., Bissacotti B.F., Copetti P.M., da Silva A.D., Schetinger M.R., Sareta L., Mendes R.E., Mesadri J., Wagner R., Gundel S., Ourique A.F. and Da Silva A.S. (2020). Effects of curcumin and Yucca extract addition in feed of broilers on microorganism control (anticoccidial and antibacterial), health, performance and meat quality. Res. Vet. Sci. 132, 156-166.
- Gao D., Zhao M., Qi X., Liu Y., Li N., Liu Z. and Bian Y. (2016). Hypoglycemic effect of *Gynostemma pentaphyllum* saponins by enhancing the Nrf2 signaling pathway in STZ-inducing diabetic rats. Arch. Pharm. Res. 39(2), 221-230.
- Guclu B. (2003). The effects of *Yucca schidigera* extract added to quail rations on egg production, egg quality and some blood parameters. *Turkish J. Vet. Anim. Sci.* **27(3)**, 567-574.
- Guclu B. and Iscan K.M. (2004). The effects of *Yucca schidigera* extract added to quail rations on fattening performance. *J. Erciyes. Univ. Fac. Vet. Med.* **1**, 5-20.
- Gupta S. (2014). Inhibitory potential of *Yucca gloriosa* L. extract and isolated gloriosaol isomeric mixture on ovalbumin induced airway hyperresponsiveness in Balb/C mice. *Clin. Pharmacol. Biopharm.* **2(1),** 1-9.
- Gurbuz E., Balevi T., Kurtoglu V. and Oznurlu Y. (2011). Effects of adding yeast cell walls and *Yucca schidigera* extract to diets of layer chicks. *Br. Poult. Sci.* **52(5)**, 625-631.
- Gurbuz E., Balevi T., Kurtoglu V. and Oznurlu Y. (2016). Use of yeast cell walls and *Yucca shidigera* extract in layer hens' diets. *Italian J. Anim. Sci.* **10(2)**, 26-34.
- Hassan S.M., Byrd J.A., Cartwright A.L. and Bailey C.A. (2010).
 Haemolytic and antimicrobial activities differ among saponinrich extracts from guar, quillaja, yucca, and soybean. *Appl. Biochem. Biotechnol.* 162(4), 1008-1017.
- Johnston N.L., Quarles C.L., Fagerberg D.J. and Caveny D.D. (1981). Evaluation of yucca saponin on broiler performance and ammonia suppression. *Poult. Sci.* 60(10), 2289-2292.
- Jun-Ling S., Bin-Lin S., Peng-Fei Z., Deng-Sheng S., Ti-Yu L. and Su-Mei Y. (2016). Effects of Yucca extract on feed efficiency, immune and antioxidative functions in broilers. *Braz. Arch. Biol. Technol.* 59, 1-11.
- Kaya S., Erdogan Z. and Erdogan S. (2003). Effect of different dietary levels of *Yucca schidigera* powder on the performance, blood parameters and egg yolk cholesterol of laying quails. *J. Vet. Med. A Physiol. Pathol. Clin. Med.* 50(1), 14-17.
- Kelly A.M. and Kohler C.C. (2003). Effects of *Yucca schidigera* extract on growth, nitrogen retention, ammonia excretion, and toxicity in channel catfish Ictalurus punctatus and hybrid tilapia *O. mossambicus* × *O. niloticus*. *J. World Aquac. Soci*.

- **34(2),** 156-161.
- Khaskheli A.A., Khaskheli M.I. and Khaskheli A.J. (2020). Dietary influence of *Yucca schidigera* on broilers and layers: A Review. *Int. J. Vet. Sci.* **9**, 458-461.
- Kim D.H., Moon Y.S., Jung J.S., Min S.K., Son B.K. and Suh H.W. (2003). Effects of ginseng saponin administered intraperitoneally on the hypothalamo-pituitary-adrenal axis in mice. *Neurosci. Lett.* 343(1), 62-65.
- Kozłowski K., Vervenne-Zetteler P., Konieczka P., Szyma' nski Ł. and van Vilsteren A. (2022). *Yucca schidigera* improves performance and lowers oocyst counts in Eimeria challenged broilers. *Animals (Basel)*. **12(13)**, 1668-1678.
- Kucukkurt I., Akkol E.K., Karabag F., Ince S., Süntar I., Eryavuz A. and Sözbilir N.B. (2016). Determination of the regulatory properties of *Yucca schidigera* extracts on the biochemical parameters and plasma hormone levels associated with obesity. *Rev. Bras. Farmacogn.* 26(2), 246-250.
- Kutlu H.R., Görgülü M. and Ünsal I. (2001). Effects of dietary *Yucca schidigera* powder on performance and egg cholesterol content of laying hens. *J. Appl. Anim. Res.* **20**(1), 49-56.
- Lan J., Wang K., Chen G., Cao G. and Yang C. (2020). Effects of inulin and isomalto oligosaccharide on diphenoxylate-induced constipation, gastrointestinal motility-related hormones, shortchain fatty acids, and the intestinal flora in rats. *Food Function J.* 11(10), 9216-9225.
- Li W.B., Wang H.Y., Ma Y.M., Zhao X.H., Yang H., Qian J.M. and Li J.N. (2018). VSL#3 can prevent ulcerative colitis-associated carcinogenesis in mice. *World J. Gastroenterol*. **24(37)**, 4254-4262.
- Liang G., Wang X., Wang X., Li C. and Chen G. (2009). Effects of camphor familial plant extract and Yucca extracts on emission of NH₃ and H₂S in slurry of weaned pigs. *Chin. J. Anim. Sci.* **45**, 22-26.
- Lillehoj H., Liu Y., Calsamiglia S., Fernandez-Miyakawa M.E., Chi F., Cravens R.L., Oh S. and Gay C.G. (2018). Phytochemicals as antibiotic alternatives to promote growth and enhance host health. *Vet. Res.* **49(1)**, 76-75.
- Luo X., Zheng C., Xia W., Ruan D., Wang S., Zhang Y.N. and Chen W. (2018). Effects of constant or intermittent high temperature on the feed intake, hypothalamic expression of antioxidant and pro-oxidant enzymes genes in laying ducks. *J. Anim. Sci.* 96(12), 5064-5074.
- Luo J.J., Chen W., Qu H., Liu Y.Q., Luo C.L., Ji J., Shu D.M. and Wang J. (2022). Dietary supplementation with *Yucca* alleviates heat stress in growing broilers exposed to high ambient temperature. *Front. Vet. Sci.* **9**, 850715-850723.
- Lundeen T. (2000). Yucca extract improves feed efficiency, decreases abdominal fat in broilers. *Feedstuffs Repr.* **72**, 1-9.
- Malecki J.A., Ramisz L., Mardarowicz L. and Pilarczyk B. (1995). The influence of *Yucca shidigera* extract on environment and productivity of broilers. *Zeszyty Naurokowe Akademii Rolniczej w. Szczecinie, Zootechnica.* **32,** 51-56.
- Mao X., Dou Y., Fan X., Yu B., He J., Zheng P., Yu J., Luo J., Luo Y., Yan H., Wang J., Wang H. and Wang Q. (2023). The effect of dietary *Yucca schidigera* extract supplementation on productive performance, egg quality, and gut health in laying hens with *Clostridium perfringens* and coccidia challenge.

- Poult. Sci. 102(8), 102822-102832.
- Martin-Gallausiaux C., Marinelli L., Blottière H.M., Larraufie P. and Lapaque N. (2021). SCFA: Mechanisms and functional importance in the gut. *Proc. Nutr. Soc.* **80(1)**, 37-49.
- Matusiak K., Oleksy M., Borowski S., Nowak A., Korczyński M., Dobrza'nski Z. and Gutarowska B. (2016). The use of *Yucca schidigera* and microbial preparation for poultry manure deodorization and hygienization. *J. Environ. Manag.* **170**, 50-59.
- McAllister T.A., Annett C.B., Cockwill C.L., Olson M.E., Wang Y. and Hahn T.W. (2001). Studies on the use of *Yucca schidigera* to control giardiosis. *Vet. Parasitol.* **97(2)**, 85-99.
- Mert S. (2025). Effects of humate, Yucca, and humate-Yucca combination supplementation on growth performance, gut health, and immune response in broiler chickens. *Front. Vet. Sci.* **12**, 1-11.
- Munezero O., Zhang Z. and Kim I. (2023). The effects of *Yucca Schidigera* extract and multi-carbohydrase in different crude protein diets on growth performance, nutrient digestibility, carcass parameters and excreta noxious gas contents in broilers. *Brazilian J. Poult. Sci.* **25(1)**, 1-8.
- Nazeer M.S., Pasha T.N., Shahid A. and Ali Z. (2002). Effect of Yucca saponin on urease activity and development of ascites in broiler chickens. *Int. J. Poult. Sci.* 1(6), 174-178.
- Obianwuna U.E., Chang X., Oleforuh O.V.U., Onu P.N., Zhang H., Qiu K. and Wu S. (2024). Phytobiotics in poultry: Revolutionizing broiler chicken nutrition with plant-derived gut health enhancers. *J. Anim. Sci. Biotechnol.* **15**, 169-175.
- Oelschlager M.L., Rasheed M.S.A., Smith B.N., Rincker M.J. and Dilger R.N. (2019). Effects of *Yucca schidigera* derived saponin supplementation during a mixed *Eimeria* challenge in broilers. *Poult. Sci.* **98(8)**, 3212-3222.
- Oleszek W., Sitek M., Stochmal A., Piacente S., Pizza C. and Cheeke P. (2001). Resveratrol and other phenolics from the bark of *Yucca schidigera* roezl. *J. Agric. Food Chem.* **49(2)**, 747-452.
- Oviedo E.O., Parker J., Clack B. and Waetherford W. (2003). Concentrate Extract of *Yucca schidigera* (BioSupreme®) Plant as Feed Additive in Broiler Diets in the Control of Eimeria Infection. Austin State University, Nacogdoches, TX, USA.
- Patel S. (2012). Yucca: A medicinally significant genus with manifold therapeutic attributes. *Nat. Prod. Bioprospect.* **2(6)**, 231-234.
- Patoary M.U., Hossain M., Akter M. and Rubel Z.U. (2020). Effect of supplementation of *Yucca schidigera* extract on ammonia gas emission and performance of broiler chickens. *J. World Poult. Res.* **10(1)**, 57-62.
- Pen B., Sar C., Mwenya B., Kuwaki K., Morikawa R. and Takahashi J. (2006). Effects of *Yucca schidigera* and *Quillaja saponaria* extracts on *in vitro* ruminal fermentation and methane emission. *Anim. Feed Sci. Technol.* **129(3)**, 175-186.
- Piacente S., Pizza C. and Oleszek W. (2005). Saponins and phenolics of *Yucca schidigera* Roezl: Chemistry and bioactivity. *Phytochem. Rev.* **4,** 177-190.
- Piacente S., Montoro P., Oleszek W. and Pizza C. (2004). Yucca schidigera bark: phenolic constituents and antioxidant activity. J. Nat. Prod. 67(5), 882-885.
- Preston C.M., McCracken K.J. and Bedford M.R. (1999). Effects of *Yucca schdigera* extract, *Saccharomyces boulardii* and en-

- zyme supplementation of wheat-based diets on broiler performance and diet metabolisability. *Br. Poult. Sci.* **40(1)**, 39-40
- Rahman M.M., Begum S. and Mondol S.I.A.K. (2023). Impact of *Yucca schidigera* plant extract supplementation to diet on body growth, carcass characteristic's, gut microbiota, intestinal ammonia concentration and hematology in broiler chickens. *J. Sylhet Agric. Univ.* **10(1)**, 17-22.
- Ranjbar Z., Shariatmadari F. and Torshizi M.K. (2014). Effect of different levels of yucca extract and antibiotic on growth performance, immune response and blood factors of broiler chickens. *Iranian J. Med. Aromat. Plants.* **30(5)**, 675-681.
- Rezaei R., Lei J. and Wu G. (2017). Dietary supplementation with *Yucca schidigera* extract alleviates heat stress–induced growth restriction in chickens. *J. Anim. Sci.* **95(4)**, 370-371.
- Saddoris-Clemons K., Osho S., Garcia M. and Humphrey B. (2024). Effects of dietary inclusion of a proprietary combination of *Quillaja saponaria* and *Yucca schidigera* on intestinal permeability and immune response in broiler chickens during a coccidia challenge. *Animals.* **14(12)**, 1737-1745.
- Saeed M., Arain M.A., Naveed M., Alagawany M., Abd El-Hack M.E., Bhutto Z.A. and Chao S. (2018). Yucca schidigera can mitigate ammonia emissions from manure and promote poultry health and production. Environ. Sci. Pollut. Res. Int. 25(35), 35027-35033.
- Saleh A.A., Paray B.A. and Dawood M.A.O. (2020). Olive cake meal and *Bacillus licheniformis* impacted the growth performance, muscle fatty acid content, and health status of broiler chickens. *Animals.* **10(4),** 695-701.
- Sahoo S.P., Kaur D., Sethi A.P., Sharma A. and Chandra M. (2015). Evaluation of *Yucca schidigera* extract as feed additive on performance of broiler chicks in winter season. *Vet. World.* 8(4), 556-560.
- Sahoo S., Kaur D., Sethi A., Saini A. and Chandra M. (2016). Effect of dietary supplementation of *yucca schidigera* extract on the performance and litter quality of broilers in winter season. *Anim. Nutr. Feed Technol.* 16(3), 475-484.
- Sariozkan S., Konca Y., Guclu B.K., Kara K., Ildiz N., Kirkpinar F., Beyzi S.B. and Kaliber M. (2015). Effects of dietary supplementation of dried distillers grain with solubles (DDGS) and yucca (*Yucca schidigera*) on broiler performance, carcass traits, intestinal viscosity and marketing. *J. Poult. Res.* 12(1), 5-11.
- Sidhu G. and Oakenfull D.A. (1986). Mechanism for the hypocholesterolaemic activity of saponins. *Br. J. Nutr.* **55(3)**, 643-649.
- Song X. and Hu S. (2009). Adjuvant activities of saponins from traditional Chinese medicinal herbs. *Vaccine*. **27(36)**, 4883-4890.
- Su J.L., Shi B.L., Zhang P.F., Sun D.S., Li T.Y. and Yan S.M. (2016). Effects of Yucca extract on feed efficiency, immune and antioxidative functions in broilers. *Braz. Arch. Biol. Tech*nol. **59**, 1-9.
- Sun D.S., Shi B.L., Tong M.M. and Yan S.M. (2017). Improved performance and immunological responses as a result of dietary *Yucca schidigera* extract supplementation in broilers. *Italian J. Anim. Sci.* **17(2)**, 511-517.
- Sun D., Jin X., Camerlink I., Tong M., Su J., Zhao F., Yan S. and Shi B. (2019). Effects of *Yucca schidigera* extract on growth

- performance and antioxidative function of small intestine in broilers. *J. Anim. Physiol. Anim. Nutr.* **103(3)**, 738-346.
- Sun X., Wang Z., Li X., Du S., Lin D. and Shao Y. (2023). Effects of *Yucca schidigera* extract on serum biochemical parameters, humoral immune response, and intestinal health in young pigeons. *Front. Vet. Sci.* **9**, 77555-77564.
- Trejo Castro L. (2002). Report of the Evaluation of Effectiveness of the Product *Yucca schidigera* as Anticoccidial in Broiler Feed; LAB.PAR.IND2001 003; Baja Agro International, Ensenada, Mexico.
- Tsukahara T. and Ushida K. (2000). Effects of animal or plant protein diets on cecal fermentation in Guinea pigs (*Cavia porcellus*), rats (*Rattus norvegicus*) and chicks (*Gallus gallus domesticus*). Comp. Biochem. Physiol. A Mol. Integr. Physiol. 127(2), 139-146.
- Tugay A., Yurtseven S., Baylan M. and Canogullari S. (2005).
 The effects of dietary *Yucca shidigera* on egg yield parameters and egg shell quality of laying hens. *Int. J. Poult. Sci.* 4, 159-162.
- Vlčková R., Sopková D., Andrejčáková Z., Valocký I., Kádasi A., Harrath A.H., Petrilla V. and Sirotkin A.V. (2017). Dietary supplementation of yucca (*Yucca schidigera*) affects ovine ovarian functions. *Theriogenology*. 88, 158-165.
- Wang Y., Mcallister T.A., Newbold C.J., Rode L.M., Cheeke P.R. and Cheng K.J. (1998). Effects of *Yucca schidigera* extract on fermentation and degradation of steroidal saponins in the rumen simulation technique (RUSITEC). *Anim. Feed Sci. Technol.* 74(2), 143-153.
- Wang J.P. and Kim I.H. (2011). Effect of caprylic acid and *Yucca schidigera* extract on production performance, egg quality,

- blood characteristics, and excreta microflora in laying hens. *Br. Poult. Sci.* **52(6)**, 711-717.
- Wang L., Wu D., Fan Z., Li H., Li J., Zhang Y., Xu Q., Wang G. and Zhu Z. (2020). Effect of *Yucca schidigera* extract on the growth performance, intestinal antioxidant status, immune response, and tight junctions of mirror carp (*Cyprinus carpio*). *Fish Shellfish Immunol.* 103, 211-219.
- Wang Y., Zhang Y., Ren H., Fan Z., Yang X., Zhang C. and Jiang Y. (2023). Dietary *Yucca extract* and *Clostridium butyricum* promote growth performance of weaned rabbits by improving nutrient digestibility, intestinal development, and microbial composition. *Front. Vet. Sci.* 10, 1-8.
- Yang Y., Lee J., Rhee M.H., Yu T., Baek K.S., Sung N.Y., Kim Y., Yoon K., Kim J.H., Kwak Y.S., Hong S., Kim J.H. and Cho J.Y. (2015). Molecular mechanism of protopanaxadiol saponin fraction-mediated anti-inflammatory actions. *J. Ginseng. Res.* 39(1), 61-68.
- Yeo J. and Kim K.I. (1997). Effect of feeding diets containing an antibiotic, a probiotic, or yucca extract on growth and intestinal urease activity in broiler chicks. *Poult. Sci.* **76(2)**, 381-385.
- Zdanowska-Sąsiadek Z., Lipińska-Palka P., Damaziak K., Michalczuk M., Grzybek W., Kruzi'nska B., Jasi'nska K., Róg D., Kordos K. and Abek K. (2019). Antioxidant effects of phytogenic herbal-vegetable mixtures additives used in chicken feed on breast meat quality. *Anim. Sci. Pap. Rep.* 37(4), 393-408.