The Phytochemical Changes of Violet Flowers (*Viola cornuta*) Response to Exogenous Salicylic Acid Hormone

N. Ghorbani^{1*}, H. Moradi¹, V. Akbarpour¹, A.Ghasemnezhad²

¹ Department of Horticultural, Sari Agricultural Sciences and Natural Resources University, Sari, Iran
²Department of Horticultural, Gorgan Agricultural Sciences and Natural Resources University, Gorgan, Iran

(Received: 27 October 2013 Accepted: 24 December 2013)

ABSTRACT: Violet is one of the ornamental plants with a good value in landscaping and herbal medicine. Salicylic acid is a signaling agent involving in secondary metabolite production. The aim of this study was to evaluate the physiological responses of violet flowers to exogenous salicylic acid. This experiment was conducted in the greenhouse, as a completely randomized design. Salicylic acid was sprayed on violet plants in four levels as 0, 0.1, 0.7, 1.5 mM and three replications. Flower diameter, flower stem length, fresh weight and dry matter percentage of violet flower were measured as morphological parametes. In laboratory parameters like antioxidant activity and anthocyanin variation were recorded using spectrophotometery method. The quercetin and rutin values were determined by HPLC. Results showed that salicylic acid significantly affected on flower diameter, total antioxidant capacity, rutin and quercetin contents. Therefore data analysis provides that high levels of salicylic acid increased morphological parameters and improved chemical substance involving to secondary metabolism promotion. Furthermore, using different concentrations of the hormone is required, to achieve the best quality and quantity of plant biomass and it is also necessary to achieve the best traits of ornamental and medicinal value

KEYWORDS: Anthocyanin, Quercetin, Rutin, Salicylic acid, Violet (Viola cornuta).

INTRODUCTION

Flowers are the beautiful signs of God's creation and the best gifts for human due to their mental and nutritional value. Medicinal plants have a particular value in the provision of community health and many drugs have emerged with plant origin in different parts of the world in recent years [18,6].Violet (*Viola cornuta*) is a plant of

the Violaceae family flowering at the end of winter and early in spring [9]. Flowers, leaves, roots, seeds and even whole parts of this flower are used for medicinal purposes [7, 28]. The violet flowers contain volatile oil, rutin, syanyn, bright pigment, methyl salicylate glycosides and anthocyanins [42].

Corresponding Author: N. Ghorbani, M.Sc. student of Horticultural Sciences, Sari Agricultural Sciences and Natural Resources University, Sari, Iran. E-mail: nayyer.ghorbani@yahoo.com

Flavonoids are the largest group of natural phenolic that quercetin is one of them. Confronting with cancer cells and viruses, extensive anti oxidant activity, potent immune enhancing medicinal properties of this material is the most important attributes of them. Thus recently the study of the situation and availability of these compounds is more focused [27, 8, 37, 16, and 39]. Rutin is a glycoside of quercetin flavonoid that both of them were used in drugs inhibiting vein blood structure and were calculated as a component of multivitamins [22].

Salicylic acid (SA) is a phenolic compound that is widely found in plants and is now considered as a hormone-like substasnce which can play an important role in plant growth [31]. It has been showed that external salicylic acid application even in low concentration increase plant ability to enhance the induction of defense compounds such as phenolic compounds [20]. Salicylic acid is involved in the physiological processes that can stimulate flowering [14], increasing the synthesis of secondary metabolites [34]. Khandakr and colleagues [19] showed that the concentration of 10⁻⁵mM increased the plant height, root length, leaf number and dry weight of red Cocks comb. Salicylic acid in cucumber [23], petunia [2], calendula [11], water garden cress [30] and borage [10] improves attributes such as shoot dry weight, chlorophylls and levels of carotenoid.

The effect of salicylic acid as a growth regulator of the cell as well as its direct and indirect influence on the production of secondary metabolites has been proven in some plants. Therefore, the main objective of this research was to study the variation of some ornamental parameters and secondary metabolites of violets under different levels of salicylic acid application.

MATERIALS AND METHODS

Used Apparatuses and Materials

Violet flower F_1 seeds were purchased from Tehran Farid Company and then surface was sterilized with sodium hypochlorite 20% for planting in greenhouse. The obtained seedlings were transferred to the 2.5 liter pots containing garden soil, cocopeat, manure and sand in 1:1:1:1 ratio. Pots were orderd based on completely randomized design with 3 replications. Treatments involved 4 different concentrations of salicylic acid (control, 0.1, 0.7 & 1.5 mM). Foliar sprays of different salicylic acid concentration were performed in 3 stages.

Morphological traits measurement

Flower diameter was measured using a digital caliper and the number of flower was continuously recorded. Flower stem length was determined with ruler accurately. Flowers were picked up after reaching full size and then were weighed on digital scale with 0.001gr accuracy. After drying in oven at 40 °C for 48 hours, the dry matter was recorded.

Antioxidant capacity (free radical scavenging) determination

To measure the amount of free radical scavenging [5], 1 mm of was mixed with 1 ml of DPPH (2,2-diphenyl-1-picrylhydrazyl) at a concentration of 0.1 mM. After 15 min incubation in dark, radical activity was measured using spectrophotometer at a wavelength of 517 nm. Then the free radical scavenging was computed as the following formula, whears as recorded as sample absorbance and AC as control was used.

Free radical scavering = ((Ac-As) / Ac) * 100

Merck Hitachi7100			
Mapadauv-1800			
Fan azmagostar BM120			
A & D Company Limiled			
Euronda 4D			
Labinko L46			
Cnturion K2042			
Merck K41142931			
Sigma Aldrich 257621			
anol Merck106007			
Merck500064			
Merck551600			

Anthocyanin measurement

To obtain the concentration of anthocyanins[43], dried flowers were pulverized with acidified methanol (ratio 1:10) and extracted in the dark at 4°C. The absorbance of samples was recorded at 520nm. Anthocyanin content calculated according to the formula of A= ϵ bc, whears ϵ is extinction coefficient (3300mMcm⁻¹), A is absorbance, b = 1 cm (width cuvette) and anthocyanin content (c) is mol/g.

Phytochemical characteristics determination

Extracts was prepared according to the proposed method of Samee &Vorarat[35], with slight modification. After of extract preparation, samples were treated in an ultrasonic bath for 10 min and then centrifuged in 3500 r.p.m. The samples were passed through micro injection filter (0.45μ m) and injected to HPLC. Merck Hitachi apparatus equipped with Lacrom Pump Model 7100 – diode array detector- (285 nm) and C18 Column (length 25 cm and in diameter 4.6 mm) was used in present study. The standard calibration curve of rutin obtained was used from different concentrations of 0, 10, 50,200, and 300 ppm and for quercetin the concentrations of 0, 5, 10, 50 and 100 ppm. The correct amount of compounds was computed based on the obtained formula for each compound.

STATISTICAL ANALYSIS

Data analysis was performed using SAS software and mean comparison was performed with LSD test at the 5% probability.

RESULTS AND DISCUSSION

The results from the analysis of variance are presented in Table 1. Obtained data suggest that salicylic acid had significant effect on flower diameter, antioxidant capacity, rutin content and quercetin content.

Data showed that the highest flower diameter (5.16 cm) was observed in the plants which were treated with salicylic acid at its highest concentration (Figure 1)

Fresh weight of flower was affected by hormones. With 0.1 mmolL⁻¹, the highest fresh weight was obtained. In medicinal plants industry, dried plant organs are used commonly and there is a direct relation between dry matter and plant material quality. By applying the 0.7 mM salicylic acid, highest percentage of dried flowers (14.46%) was observed.Contrary to that lowest flower yield was achieved at control level. Induce flowering,

growth, respiration and ethylene synthesis are important

effects of salicylic acid hormone [38].

Source Change	Degrees of freedom	Flower diameter	Flower stems length	Fresh flower weight	Dry matter of flowers	Flower Anti oxidant capacity	Flower Anthocyanin	Flower Rutin	Flower Quercetin
Hormon levels	3	0.45**	0.28 ^{n.s}	0.004 ^{n.s}	1.96 ^{n.s}	57.17*	0.0008 ^{n.s}	916582.78**	8.42**
Experimental error	8	0.02	0.37	0.004	1.92	12.82	0.0004	8172.02	0.002
Coefficient of Variation%	-	3.1	15.3	14.9	10.3	6.5	14.8	9.1	0.08

Table 1. Variance analysis of of salicylic acid treatment on violets (Viola cornuta).

* & ** significant at 5 and 1% probability level and n.s is not significant.



Figurer 1. Effect of Salicylic acid on flower diameter

This regulator increases protein production and by creation new isozymes a band that induces flower buds and increases their number [21]. Stimulatory effects of salicylic acid can grow through increased growth and cell division in meristematic regions and increase the effect of this hormone on other plant hormones [36]. In the same study, salicylic acid hormone on African violets [15], and Gompherna flower [17], found similar results with this study.

The length of flower stem in plants due to better feature of flowers appearance is the important parameter in ornamental plants. In this study, the length of flower stem increased as the salicylic acid increased from 0.1 to 1.5 mmolL⁻¹(4.2 cm). Expansion and cell division is regulated by salicylic acid hormone, indeed there is a balance between growth and aging by this hormone [32]. Salicylic acid together with auxin also affect elongation and cell division [13]. This hormone also played in specific proteins synthesis that called kinase protein; these proteins also play an important role in the regulation of division, differentiation and cell morphology formation [44]. Several reports of the effect of salicylic acid on shoot and root length of the plants available. In soybean and cucumber plants it has been showed that by increasing the level of this hormone an increase in nitrate reductase activity resulted to increase in measured traits [38, 12]. Results of Mehrabian and colleagues [25] showed that compared to other concentrations 0.1 and 0.2 mM SA, increased the shoot length, root length, fresh and dry weight of maize seedlings.

One important factor in the medicinal plant is their antioxidant capacity. In this study, the effect of salicylic acid on this attribute was significant. In level of 0.7 mM treatments (Figure 2), antioxidant capacity, reached up to a maximum (54.9%). Under stress conditions plants

increase the activity of antioxidant defense enzymes to protect themselves against further damages [1]. It seems that treating by salicylic acid enhances the antioxidant



Figure 2.Effect of salicylic acid on anti oxidant capacity

Although the content of anthocyanins tend to decrease with increasing salicylic acid, but no significant difference was observed. It has been showed that salicylic acid affect phenolic accumulation by activation different enzymes like phenylalanine ammonia lyase(PAL) and chalcone synthase (CHS). Thus reduction of anthocyanin could be explained by the conversion of flavonoids to each others or more active substances [4]. Another reason for this result can be attributed to inhibition of ethylene synthesis [33]. This result is similar to Bernard and colleagues [3] in tea plant.Both Rutin (Figure 3) and quercetin (figure 4), as important flavonoids compounds, were significantly varied in different levels of salicylic acid application (Figures 3 & 4).

protection of the cell membrane, photosynthetic pigments, and causes to improve the growth index and secondary metabolites, ultimately [29].



Figure3. Effect of Salicylic acid on rutin content



Figure 4. Effect of Salicylic acid on quercetin content

Salicylic acid induces enzyme activity to increase active ingredients and flavonoids. This hormone is known as a key message component in the specific activation plant response and these responses lead to biosynthesis and accumulation of plant secondary compounds [31]. Rutin was shown to consist of bioactive compounds that may help protect plants against ultraviolet radiation or pathogens and could be used to prevent the side effects of cancer treatments, diabetes, and hypercholesteremia [24, 26]. The results of Sun and colleagues [40], rutin content of leaves of buckwheat (*Fagopyrumtartaricum*) markedly increased after treated with salicylic acid, by

increasing the activity of enzymes are involved in rutin production. Based on the role of salicylic acid in plant defense system, it seems that external salicylic acid application induces pseudo stress conditions in plant enhances the antioxidant protection via secondary metabolite accumulation [29]. Rutin and Quercetin are valuable glycoside polyphenolic compounds of violet and many of other plants that are important in medicine. in most cases a direct relationship was observed between the amount of salicylic acid application and and defense compounds accumulation [41].

CONCLUSION

The results of this study showed that salicylic acid had effective influence on morphological plant characteristics, such as size and the number of flowers. In addition, the effects of this hormone on secondary metabolites accumulation indicate that salicylic acid as a plant hormone and stress signaling compound induces pseudo stress conditions and encourages plant to start its tolerance system. If salicylic acid applied in suitable amount act as a good target in secondary metabolite production. It should be mentioned that high amount of this compound can limit the secondary product production. Based on the obtained results it can be concluded that salicylic acid in the range of 1.5mM could be appropriate for viola secondary metabolite production.

REFERENCES

1. Asada K., Takahashi M., 1987. Production and scavenging of active oxygen in chloroplasts. In Kyle, D.J., Osmond, C.B., Arntzen, D.J., eds, Photoinhibition. Elsevier, Amsterdam, 227–287.

2.Bayat H., Nemati S. H., Tehranifar A., VahdatiN., Salahvarzi Y., 2012. Effect of salycilic acid on growth and ornamental charctricticts of petunia under salt stress.Sciences and Technologys of Greenhouse Culture. 11:43-50. 3. Bernard F., Kargar Z., Shaker-Bazarnov H., Davarani S.S.H., 2003. Antagonistis effects of mannitol and SA on (+) - catechin accumulation in *Camellia sinensis* L. Calluses. Iranian Journal of Science & Technology. 27: 169-174.

4. Bernard F., Nouri M., MehrabiKushki Z., Shaker H., 2008. Comparison of physiological and biochemical responses of two separate pieces of cultivated varieties licorice to molybdenum and salicylic acid.ROSTANIHA.9 (1).89-81.

5.Ebrahimzadeh M. A., Nabavi S. M., Nabavi S. F., Bahramian F., Bekhradnia A. R., 2010. Antioxidant and ferr radical scavering activity of H.officinalis L. varangustifolius, V.odorata, B.hyrcana and C.speciosumPakistan Journal of Pharmaceutical Sciences. 23(1):.29-34.

6. Fazelian N., Asrar Z., 2011. Interaction of arsenic and salicylic acid on growth and some physiological indexes chamomile, Plant Biology, 8: 12-1.

 Flok H., Translate TavakoliSaberi M., Sedaghat M., Publication Rouzbahan, Tehran, 2005.

8. Formica J. V., Regelson W., 1995. Review of the biology of quercetin and related bioflavonoids. Food Chem Toxicol. 33:1061–1080.

9. GhasemiGhahsare, M.,Kafi M.,Scientific and practical floriculture.Publications Golbon,Vol.1, Isfahan 2007.

10. Ghorbani N., Akbarpour V., Yavari Z., 2013. Effect of salicylic acid on some growth and physiological traits of borage (*Boragoofficinalis*). Eighth Congress of Iranian Horticultural Science, Hamedan.

11. Ghorbani N., Moradi H., Akbarpour V., Ashnavar M., Yavari Z., 2013. Response of ornamental traits of two cultivars single and double Marigold (*Calendula officinalis*) in hormone salicylic acid. Second Congress of citrus, sari, iran.

12. Gutierrez-Coronado M.A., Trejo-Lopez C., Larqué-Saavedra A., 1998. Effects of salicylic acid on the growth of roots and shoots in soybean.Plant Physiology and Biochemistry. 36(8): 563-565.

13. Hashemi SH. F., Asrar Z., Pour Seyyedi SH., 2010. Effects of salicylic acid seed priming on growth and some physiological and biochemical indice cress (*Lepidiumsativum*). Journal of Plant Biology. 2(4): 1-10. 14. Hayat Q., Hayat Sh., Irfan M., Ahmad A., 2010. Effect of exogenous salicylic acid under changing enveironment, a review Enviromentaland Experimental Botany. 68: 14-25.

15. Jabbarzadeh Z., Khosh-Khui M., Salehi H., 2009. The effect of foliar-applied salicylic acid on flowering of African Violet. Australian Journal of Basic and Applied Sciences. 3(4):4693-4696.

16. Jaber R., 2002. Respiratory and allergic diseases: from upper respiratory tract infections to asthma. Primary Care. 29(2): 231-261

17. Kamali M., Kharazi S. M., Salah Varzi, Y., Tehranifar A., 2011. Effect of salicylic acid on growth and some physiological characteristics of flower bud (*Gompherna globosa* L.) under salt stress. Journal of Horticultural Science (Agricultural Sciences and Technology). 26 (1):112-104.

18. Karimi M., 2006. Effects of temperature and various chemical treatments to increase the longevity of cut flowers lily. Master's thesis, university ogguilan. Iran.

19. Khandaker, L., Masumakond, A. S. M. G. and Oba, Sh., 2011. Foliar application of salicylic acid improved the growth, yield and leafs bioactive compounds in Red Amaranth (*Amaranthus tricolor L.*). Vegeteble Crops Research Bullein. 24: 77-86.

20. KhavariNejad R. A., Mehrabian S., Asadi A., 2004. Effect of salicylic acid on anthocyanins of infected Daisy medicinal plant. Journal of Science Teacher Education University. 4(3):427-438.

21. Khurama J. P. S., Cleland C. F., 1992. Role of salicylic acid and benzoic acid in flowering of photoperiodinsensitive strain, *Lemna paucicostata*LP6.Plant Physiology. 100: 1541-1546.

22. Kreft S, Knapp M., Kreft I., 1999. Extraction of rutin from buckwheat (*FagopyrumesculentumMoench*) seeds

and determination by capillary electrophoresis, Journal of Agriculture and Food Chemistry. 47 (11):4649–4652.

23. Mardani H., Bayat H., Azizi M., 2011. Effects of salicylic acid on morphological characteristics and physiological solution sprayed cucumber seedlings under drought stress. Journal of Horticultural Science. 25 (3): 326-320.

24. Martens S., Mithofer A., 2005.Flavones and flavone synthases.Phytochemistry. 66:2399–2407.

25. Mehrabian Moghaddam N., Arvin M. J., Khajuii NejadGh.,Maghsoudi K., 2011. Effect of salicylic acid on growth and grain yield in drought stress in the field.Journal of Seed and Plant Agricultural. 2-27 (1): 55-41.

26. Mehta RG, Murillo G., Naithani R., Peng X.J., 2010. Cancer chemoprevention by natural products: how far have we come? Pharmaceutical Research. 12:950–961

27. Middleton J.R.E., Kandaswami C., 1994. The effect of plant flavonoids on mammalian cells: Implications for inflammation, heart disease and cancer. Pharmacological reviews.52 (4): 675-751.

28. Moaveni P., Medicinal Plants, Volume I, published by University of Shahreghods, 2009

29. Momeni N., Arvin M. J., KhajuiiNejad GH., Keramat B., Daneshmand F., 2012. Some effects of sodium chloride and salicylic acid on photosynthetic parameters and mineral nutrition of maize plants (*Zea mays* L.), Plant Biology. 15(5):30-15.

30. Moradi S., 2013. The effect of different concentrations of salicylic acid and putrescine on morphological and phytochemical water garden cress. Horticulture M.Sc. Thesis, University of agriculture and natural Resources, Sari.

31. Mueller M.J., Brodschelm W., Spannagl E., Zenk M.H., 1993. Signaling in the elicitation process is mediated through the octadecanoid pathway leading to jasmonic acid. Proceedings of the National Academy of Sciences of the United States of America. 90:7490–4.

32.Popova L., Ananieva V., Hristova V., Christov K., Georgieva K., Alexieva V., StoinovaZh., 2003. Salicylic acid and methyl jasmonate induced protection on photosynthesis to parquet oxidative stress. Bulgarian Journal of Plant Physiology.Special Issue.133-152.

33. Qinghua S. H., Zhujun Z., 2008. Effect of exogenous salicylic acid on manganese toxicity, element contents and antioxidative system in cucumber. Environmental and Experimental Botany. 63:317-326.

34. Raskin I., 1992. Role of salicylic acid in plantsAnnual Review of Plant Physiology and Plant Molecular Biology., 43: 439–463.

35. Samee W., Vorarat S., 2007. Simultaneous Determination of Gallic acid, Catechin, Rutin, Ellagic Acid and Quercetin in Flower Extracts of *Micheliaalba,Caesalpiniapulcherrima* and

*Nelumbonucifera*by HPLC, Thai Pharmaceutical Health Science Journal 2(2):131-137.

36. Shakirova F. M., Sakhabutdinova A. R., BozrutkovaM. V., Fatkhutdinova R. A., Fatkhutdinova, D. R., 2003. Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity.Plant Science. 164: 317-322.

37. Shoskes D.A., Zeitlin S.I., Shahed A., Rajfer J., 1999. Quercetin in men with category III chronic prostatitis: a preliminary prospective, double-blind, placebo-controlled trial. Urology. 54(6): 960-963.

38. Singh P.K., Chaturvedi V.K., Bose B., 2010. Effects of salicylic acid on seedling growth and nitrogen

metabolism in cucumber (*Cucumissativus* L.).Journal of Stress Physiology and Biochemistry. 6(3):102-113.

39.Stewart L.K., Soileau J.L., Ribnicky D., Wang Z.Q., Raskin I., Poulev A., Majewski M., Cefalu W.T., GettysT.W., 2008. Quercetin transiently increases energy expenditure but persistently decreases circulating markers of inflammation in C57BL/6J mice fed a highfat diet. Metabolism Clinical and Experimental. 57: S39-S46

40. Sun Z., Hou S., Yang W., 2012. Exogenous application of salicylic acid enhanced the rutin accumulation and influenced the expression patterns of rutin biosynthesis related genes in *Fagopyrumtartaricum*Gaertn leaves, Plant Growth Regul. 68:9–15.

41. Taiz L, Zeiger E. Plant Physiology.Benjemin Cummings Publishing Company, New York, 1999.

42.ValadAbadi S. A., Mohammad Beigi F., Daneshian J., Herbal medicines, Azad University of Shahreghodspublication, shahreghods, 2010.

43. Wanger G.J., 1979. Content and vacuole/extra vacuole distribution of neutral sugars, free amino acids and anthocyanins in protoplasts.Plant Physiology. 64: 88-93.

44. Zhang S., Klessig D. F., 1997. Salicylic acid activates a 48-KD MAP kinase in tobacco. Plant and Cell Physiology. 9: 809-824.