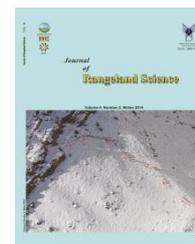




Contents available at ISC and SID

Journal homepage: www.rangeland.ir



Full Length Article:

Investigation of Heavy Metals Accumulation in Plants Growing in Contaminated Soils (Case Study: Qazvin Province, Iran)

Mahdieh Ebrahimi^A, Mohammad Jafari^B, Gholam Reza Savaghebi^C, Hossein Azarnivand^D, Ali Tavili^E, Fernando Madrid^F

^AAssistant Professor, Faculty of Water and Soil Science, University of Zabol, Iran (Corresponding Author), Email: maebrahimi2007@yahoo.com

^BProfessor, Faculty of Natural Resources, University of Tehran, Iran

^CAssociate Professor, Faculty of Agriculture, University of Tehran, Iran

^DAssociate Professor, Faculty of Natural Resources, University of Tehran, Iran

^EAssociate Professor, Faculty of Natural Resources, University of Tehran, Iran

^FTitled Superior Specialized, IRNAS-CSIC, Spain

Received on: 21/01/2013

Accepted on: 17/04/2013

Abstract. Environmental pollution with heavy metals is a global disaster that is related to human activities. This study was conducted to determine the extent of heavy metals accumulation by plant species in Lia industrial city (Qazvin, Iran) and to investigate the remediative capacity of native plant species grown in the contaminated soils. Soil and industrial wastewater sampling was done radially along transects with 300 m intervals from exit point of wastewater at three sites. In each sampling point, along 100 m transects within 5×2 m plots, the plant samples and soil samples were collected in depth of 0-20 cm and 20-40 cm from rhizosphere zone. Concentration of copper, zinc and chromium in root and shoot of 11 plant species, soil and wastewater were analyzed in three sites for mentioned metals. Bio Concentration factors and translocation factor were determined to ensure phytoremediation availability. Results showed that the concentrations of metals in the soil and wastewater greatly exceeded the threshold limit values. The contents of metals in soils ranged in the order of Cr>Zn>Cu and in wastewater were in the order of Zn>Cr>Cu, respectively. Results showed that *Scirpus maritimus* L. and *Phragmites australis* (Cav.) Trin. ex Steudel presented the highest accumulation of Zn, Cu and Cr in their root tissues which were suitable for phytostabilization (with a high BCF couple with low TF). The lowest extractable Zn (7.24 and 3.29 mgkg⁻¹ for shoot and root respectively, BCF=0.07) and extractable Cu (2.56 and 2.80 mgkg⁻¹ for shoot and root respectively, BCF=0.14) were related to *Hordeum glaucum* L. Moreover, the relatively lowest values of Cr were measured for *Taraxacum officinale* L. Results indicated that the species, which had low metal bioaccumulation in their roots and high TF, could play important roles for removal of heavy metals through phytoextraction.

Key Words: Phytoremediation, Heavy metals, BCF, TF, Lia industrial city

Introduction

Contamination of soils with toxic heavy metals is a major environmental problem that can affect both plant productivity and safety as food and feed crops (Alloway, 1990; Kabata-Pendias, 2001). In fact, heavy metals have a significant toxicity for humans, animals, microorganisms and plants (Hernandez-Ochoa *et al.*, 2005; Bodar *et al.*, 2006; Fotakis and Timbrell, 2006; Ok *et al.*, 2011). Many methods, including removal, incineration and removal followed by thermal desorption, have been used for the cleanup of contaminated soils (Joner and Leyval, 2001). In this way, phytoremediation, using plant species to restore the deteriorated soils, is a promising technology in cleanup of polluted sites due to less destructive, low cost and environmentally friendly nature (Wang *et al.*, 2012) and it can be categorized into two different approaches: (i) phytoextraction, metal accumulating species are planted on a contaminated soil and later harvested in order to remove metals from the soil (Salt *et al.*, 1995; Yoon *et al.* 2006; Usman and Mohamed, 2009), and ii) phytostabilization, metal-tolerant species are used to reduce the mobility of metals, thus, the metals are stabilized in the substrate (Salt *et al.*, 1995; Abdel-Ghani *et al.*, 2007; Antosiewicz *et al.*, 2008).

Some metallophytes are called hyperaccumulators, as these plant species can accumulate very high metal concentrations in their aerial tissues, besides normal levels found in most species (Baker and Brooks, 1989). There are also some plant species called excluders that can restrict the uptake and transport of elements between roots and shoots, maintaining low metal levels inside plant body over a wide range of external concentrations (Baker, 1981). Depending on the ability of plant species used as a phytoremediation to accumulate and tolerate heavy metals, plants are classified into three categories of

hyperaccumulators, indicators and excluders (Ghosh and Singh, 2005).

Hyperaccumulators can accumulate very high metal concentrations in their aerial tissues, besides normal levels found in most species (Baker and Brooks, 1989). Indicators can uptake and transport heavy metals to aerial tissues regularly, so that tissue concentrations are proportional to environmental concentrations (Greger, 1999; Ghosh and Singh, 2005). Excluders can restrict uptake and transport of elements between roots and shoots, maintaining low metal levels inside plant body over a wide range of external concentrations (Baker, 1981). The ability of selecting species of plants, which are either resistant to heavy metals, or can accumulate great amounts of them, would certainly facilitate reclamation of contaminated areas (Chehregani *et al.*, 2009; Lasat, 2002). Since native plant species can survive better under toxic metal stress compared to invasive plant species, it is possible to identify hyperaccumulators among native species in contaminated areas (Yoon *et al.*, 2006; Nouri *et al.*, 2011). This study was conducted to determine the concentrations of Zn, Cu and Cr in species growing in contaminated soils of Lia industrial city and to identify the remediation ability of native plant species for phytoremediation purpose.

Materials and Methods

Study area

The study area was located at Lia industrial city, Qazvin Province, Iran. This area was designed for agriculture and industrial purposes. The coldest and hottest months are February and July whose mean temperatures are, 7.2°C and 21.7°C, respectively, with an annual mean temperature of 13.9°C. The annual mean rainfall is 321.5 mm and the soil is classified as aridisols. Industrialization in this area has exposed the soil to various effluent inputs including heavy metals. This has resulted in contamination of the soils which need to be improved.

Sampling method

Sampling was carried out from May (2009) to February (2010). Sampling was done along transects with 300 m intervals in three locations due to the widespread of wastewater in the area. Samples were taken from plant, industrial wastewater and sediment. In each sampling point, based on the vegetative state and cover area, a total of 11 plant species were collected from three locations (along 100 m transects within 5×2m plots (according to dominant plant species, Bonanno *et al.*, 2010) (Table 1) with their corresponding soils. The collected plant samples were washed with tap water to remove sediments and quickly transported in plastic bags to the laboratory for analysis.

Industrial wastewater and sediment samples were collected at each sampling point. Industrial wastewater samples were collected in 0.5L clean polyethylene bottles and kept at 3°C until analysis (Bonanno *et al.*, 2010). Sediment samples were collected using a stainless steel collector at about 0–20cm and 20–40cm depths of soils from each sampling point.

Metal content analysis

The washed plant samples were divided into roots and shoots to recognize the different bioaccumulation capability. As the second step, samples were oven dried at 70 °C to a constant weight for approximately 48h and ground into fine powder (Yang *et al.*, 2009). Copper (Cu), Zinc (Zn) and Chromium (Cr) were analyzed after mineralization of 400 mg dry shoot material in a microwave oven with 5 ml of nitric acid (69% v/v), 5 ml deionized water and 2 ml H₂O₂ (30% v/v). The digest was made to 25ml final volume with deionized water, filtered (0.45 mm, Millipore) and then analyzed for Zn, Cu and Cr using ICP/OES (Du Laing *et al.*, 2003). Prior to analysis, industrial wastewater samples were passed through Whatman filters. Dried sediment samples were passed through a 2mm diameter sieve. About 100 mg dry sediment was digested with HNO₃ and

HCl (3:1) in a microwave oven. (Du Laing *et al.*, 2003). After mineralization, the samples were diluted, filtered and analyzed. Metal concentrations (Zn, Cu, Cr) of sediment samples (Table 2) and wastewater (Table 3) were measured as described for the plant samples. All the analyses were performed in five replicates.

Phytoextraction efficiency

The Bio Concentration Factor (BCF) and Translocation Factor (TF) were calculated to determine the heavy metals phytoextraction efficiency (Wilkins, 1978; Zayed *et al.*, 1998; Mattina *et al.*, 2003; Yoon *et al.*, 2006). The BCF expresses the ability of a plant to accumulate metal from soils (Metal in root DW (Dry Weight)/Metal in soil DW (Dry Weight)) (Yoon *et al.*, 2006) and TF is the capacity of a plant to transfer metal from its roots to shoots (Metal in shoot DW/Metal in root DW) (Santillan *et al.*, 2010).

Statistical analysis

All data were checked for their normality and homogeneity of variance, and where necessary, data were log-transformed before statistical analysis. Statistical significance was computed by analysis of variance (ANOVA) at $p < 0.05$. Regarding plants data, the statistical model was based on two groups (root and shoot), and aimed to show whether plant organs triggered a different accumulation pattern of a given trace element. Duncan test post hoc analysis was performed to define which specific mean pairs were significantly different. The ANOVA for industrial wastewater and sediment considered three groups (Zn, Cu, Cr) in order to detect significant different levels of concentration within the same kind of sample. The difference of metals concentration between soil sampling depths was analyzed by Student's t-test. All statistical calculations were performed using SPSS 16.0 software.

Table 1. Native plant species in surrounding area of Lia region

Family	Plant Species
Cyperaceae	<i>Scirpus maritimus</i>
Poaceae	<i>Phragmites australis, Hordeum glaucum, Bromus tectorum, Poa bulbosa</i>
Polygonaceae	<i>Polygonum lapathifolium</i>
Alismataceae	<i>Alisma plantago-aquatica</i>
Brassicaceae	<i>Cardaria draba</i>
Asteraceae	<i>Lactuca orientalis, Taraxacum officinale</i>
Euphorbiaceae	<i>Euphorbia helioscopia</i>

Results

Soil and wastewater Characteristic

Chemical analysis of soil including cation exchange capacity (Bower and Hatcher method; Bower and Hatcher, 1966), total N (Kjeldahl method; Black, 1965), organic carbon content (Walkley-Black method; Walkley and Black, 1934), EC (solid: deionized water = 1:2 w/v; Rowell, 1993), pH (1:1 soil/ water ratio; Thomas, 1996) and CaCO₃ equivalent (Drouineau and Galet method; Loeppert and Suarez, 1996) are listed in Table 2.

Acidity (pH) and Electrical Conductivity (EC) showed that the soils are categorized in salin sodic soils and are poor in terms of organic matter (less than one percent). The texture of the soils was clay to clay loam and due to low organic materials, clay particles had a

special role in the soils. The metals content in soil of the studied area greatly exceeded the threshold limit value (2758.50-74.36 mgkg⁻¹ Zn, 436.76-85.38 mgkg⁻¹ Cu and 1549.6-215 mgkg⁻¹ Cr (Table 3). The concentration of metals decreased in industrial wastewater and was in order of Zn > Cu > Cr (Table 4) the level of Zn was higher than the levels of Cu and followed by Cr. Although analyzed metal contents in soils and wastewater were highly variable, no significant variation concentration levels were observed for the metals during the experimental period among locations (sites 1, 2 and 3). Metal concentrations in the 0–20 cm depth showed higher concentrations of metals than that for the top soil (Table 3).

Table 2. Chemical and physical characteristics of soil

Site	Texture	Depth (cm)	CEC (meq)	N (%)	OC (%)	EC (dSm ⁻¹)	pH	CaCO ₃ (%)
1	Clay loam	0-20	36.00	0.10	0.12	2.24	8.41	12.28
	Clay	20-40	45.04	0.06	0.15	3.46	8.30	14.04
2	Clay loam	0-20	26.08	0.15	0.13	2.37	8.37	13.95
	Clay	20-40	47.39	0.13	0.15	3.33	8.85	16.46
3	Clay	0-20	48.69	0.08	0.13	2.78	8.42	13.27
	Clay loam	20-40	29.34	0.06	0.16	3.05	8.89	15.33

Table 3. Mean concentrations of Zn, Cu and Cr (mg kg⁻¹) in soil sediments

Site	Depth (cm)	Cr (mg kg ⁻¹)	Cu (mg kg ⁻¹)	Zn (mg kg ⁻¹)
1	0-20	1549.60 ± 9.50 a (a)	436.76 ± 4.50 a (c)	725.04 ± 7.50 a (b)
	20-40	369.36 ± 7.50 b (a)	209.16 ± 6.20 b (b)	375.70 ± 6.00 b (a)
2	0-20	674.44 ± 7.50 a (a)	269.68 ± 60.0 a (b)	265.20 ± 6.00 a (b)
	20-40	215.00 ± 6.20 b (a)	85.38 ± 3.20 b (b)	74.36 ± 0.10 b (b)
3	0-20	725.08 ± 7.50 a (b)	280.16 ± 6.00 a (c)	2758.50 ± 9.20 a (a)
	20-40	229.49 ± 6.00 b (a)	106.04 ± 3.20 b (b)	181.94 ± 6.00 b (a)

Note: Mean values are reported with SD (Standard Deviation)
 Means with the same letter for soil sampling depth (column) are not significantly different (p< 0.05)
 Means in parenthesis with the same letter for element concentration (row) are not significantly different (p< 0.05)

Table 4. Mean concentrations of metals (mg L⁻¹) in industrial wastewater

Site	Cr (mg L ⁻¹)	Cu (mg L ⁻¹)	Zn (mg L ⁻¹)
1	20.07 ± 0.05 a (b)	11.77 ± 0.02 a (c)	31.61 ± 0.02 a (a)
2	24.35 ± 0.05 a (b)	14.99 ± 0.01 a (c)	34.35 ± 0.01 a (a)
3	21.83 ± 0.01 a (b)	13.53 ± 0.06 a (c)	40.94 ± 0.40 a (a)

Note: Mean values are reported with SD (Standard Deviation)

Means with the same letter for soil sampling site (column) are not significantly different (p < 0.05)

Means in parenthesis with the same letter for element concentration (row) are not significantly different (p < 0.05)

Metal concentration in plant tissues and BCF and TF coefficients

Eleven plant species were identified in the study area (Table 1) that belong to different plant families. The data showed that the metal contents in the plant tissues differed among species at the polluted sites indicating their different capacities for metal uptake (Tables 5, 6, 7). The species with the highest Zn, Cu and Cr concentration for shoots and roots were *S. maritimus* and *P. australis*, respectively and metal concentrations were above the phytotoxic range in both species

Zinc concentration in roots of the plants varied from 3.29–238.50 mg kg⁻¹ and the level of Zn in shoots was from 7.24–93.16 mg kg⁻¹ that the highest BCF of Zn was obtained for *S. maritimus* and *P. australis* with average values of 5.4 and 4.7 and the lowest values of TF were 0.41 and 0.39 respectively. The lowest extractable Zinc with average values of 7.24 mgkg⁻¹ and 3.29 mgkg⁻¹ was

obtained for shoot and root in *H. glaucum*, respectively.

With regard to copper concentrations, in shoots of the plants, Cu varied from 2.56–43.40 mg kg⁻¹ and in root tissues, whereas the level of Cu ranged from 2.80–73.06 mg kg⁻¹ in the root tissues. The highest BCF values of Cu with average values of 3.85 and 3.05 were obtained for *S. maritimus* and *P. australis*, respectively. *P. australis* with average values of 0.74 had the lowest TF coefficient. The lowest extractable Cu with average values of 2.56 and 2.80 mgkg⁻¹ were obtained for shoot and root in *H. glaucum*, respectively (BCF=0.14). The Chromium concentrations in shoot and root ranged from 2.90–8.50 mgkg⁻¹ and 3.44 to 296 mgkg⁻¹, respectively coupled with the highest BCF values of 19.70 and 4.26 for *S. maritimus* and *P. australis*, respectively. The lowest Bio Concentration Factor (BCF) of chromium with average values of 0.22 was obtained in *T. officinale*.

Table 5. Zinc concentration (mgkg⁻¹) and coefficient in plant samples

Species	Shoot	Root	TF	BCF
<i>Phragmites australis</i>	86.59	210.35	0.41 c	4.70 a
<i>Scirpus maritimus</i>	93.16	238.50	0.39 c	5.40 a
<i>Polygonum lapathifolium</i>	51.80	31.21	1.66 b	0.70 b
<i>Cardaria draba</i>	14.45	41.16	2.80 b	0.93 b
<i>Alisma plantago-aquatica</i>	66.40	13.64	4.80a	0.31c
<i>Hordeum glaucum</i>	7.24	3.29	2.20 b	0.07e
<i>Bromus tectorum</i>	52.60	42.07	1.24 b	0.95 b
<i>Lactuca orientalis</i>	51.20	41.09	1.24 b	0.73 b
<i>Poa bulbosa</i>	34.60	19.86	1.74 b	0.45 bc
<i>Taraxacum officinale</i>	59.30	41.70	1.42 b	0.82 b
<i>Euphorbia spp</i>	66.24	45.78	1.44 b	0.63 b

Means with the same letter in each column are not significantly different (p < 0.05)

Table 6. Copper concentrations (mgkg^{-1}) and coefficient in plant samples

Species	Shoot	Root	TF	BCF
<i>Phragmites australis</i>	43.40	58.00	0.74 c	3.05 a
<i>Scirpus maritimus</i>	32.55	73.06	0.44 c	3.84 a
<i>Polygonum lapathifolium</i>	6.40	4.47	1.43 b	0.22 c
<i>Cardaria draba</i>	26.40	9.30	2.83 b	0.48 c
<i>Alisma plantago-aquatica</i>	16.71	3.40	4.90 a	0.20 c
<i>Hordeum glaucum</i>	2.56	2.80	0.90 cb	0.14 c
<i>Bromus tectorum</i>	34.40	17.50	1.96 b	0.87b
<i>Lactuca orientalis</i>	16.14	7.55	2.13 b	0.37 c
<i>Poa bulbosa</i>	3.30	4.70	0.70 b	0.23 c
<i>Taraxacum officinale</i>	25.80	22.60	1.14 b	0.86 b
<i>Euphorbia spp</i>	25.95	18.60	1.39 b	0.91b

Means with the same letter in each column are not significantly different ($p < 0.05$)

Table 7. Chromium concentrations (mgkg^{-1}) and coefficient in plant samples

Species	Shoot	Root	TF	BCF
<i>Phragmites australis</i>	50.40	63.90	0.78 c	4.26 a
<i>Scirpus maritimus</i>	85.00	296.0	0.28 d	19.70 b
<i>Polygonum lapathifolium</i>	21.03	8.50	2.47 b	0.56 c
<i>Cardaria draba</i>	12.39	10.03	1.23 b	0.66 c
<i>Alisma plantago-aquatica</i>	18.50	6.60	2.80 b	0.44 c
<i>Hordeum glaucum</i>	21.30	4.58	4.60 a	0.30 c
<i>Bromus tectorum</i>	11.70	8.27	1.41 b	0.55 c
<i>Lactuca orientalis</i>	2.90	5.80	0.50 c	0.38 c
<i>Poa bulbosa</i>	4.70	12.30	0.38 d	0.82 c
<i>Taraxacum officinale</i>	7.70	3.44	2.23 b	0.22 c
<i>Euphorbia spp</i>	12.50	9.81	1.27 b	0.65 c

Means with the same letter in each column are not significantly different ($p < 0.05$)

Discussion and Conclusion

Contaminated soil with toxic heavy metals is a serious environmental problem, which may be solved with phytoaccumulation. In this study, analyzed metal contents in soils ($100\text{--}400 \text{ mgkg}^{-1}$ Zn, Cu $60\text{--}125 \text{ mgkg}^{-1}$ and $75\text{--}100 \text{ mgkg}^{-1}$ Cr, Alloway, 1990) and wastewater (Zn $>10 \text{ mgLi}^{-1}$, Cr $> 0.5 \text{ mgLi}^{-1}$ and Cu $>1 \text{ mgLi}^{-1}$, U.S. EPA, 1993) were found to be high (Table 3).

ANOVA results of soil and wastewater samples showed that there were no significant differences among samples at three sites ($p < 0.05$); it might be due to the monotonous distribution of heavy metals at the profile of soil and relatively high availability in the sediments and wastewater. The results demonstrated that some plants could tolerate a wide range of metal concentrations in the soils. According to Alloway (1990), Zn, Cu and Cr at high threshold limits would be considered toxic to plants based on total fractions in

soil. The metals contents in the study area of Lia greatly exceeded these ranges (Tables 5, 6, 7); therefore, the 11 plant species grown in these contaminated sites exhibited high metals tolerance. The species with the highest heavy metals concentration for shoots and roots were *S. maritimus* and *P. australis*, respectively and metal concentrations were above the phytotoxic range ($100\text{--}400 \text{ mgkg}^{-1}$ Zn, $20\text{--}100 \text{ mgkg}^{-1}$ Cu and $5\text{--}30 \text{ mgkg}^{-1}$ Cr) in both species (Kabata-Pendias, 2001).

Metal concentrations in plants vary with plant species (Alloway *et al.*, 1990). BCFs were calculated to assess concentrations in roots to environmental loading. In this study, metals in *S. maritimus* and *P. australis* were accumulated in roots with concentrations greater than was found in adjacent sediments with BCF of >1 . The dominant uptake pathway of metals from the sediment was via the rhizosphere system. It is generally known that most metals tend to accumulate in the roots rather

than in shoots (Fitzgerald *et al.*, 2003), which suggests that the plants adopt either external or internal exclusion mechanisms to hinder translocation of metals to the aerial tissues (Hansel *et al.*, 2001). Khan *et al.*, (2009) in their study for the purification of industrial wastewater by macrophyte species reported the highest bioconcentration factor of chromium (3.5) for the *P. australis*. These researchers introduced species with BCF of >1 suitable for the purification of contaminated soils. TFs were calculated to enable the assessment of transport of accumulated metals from root to shoot and the ability of phytoremediation has commonly been characterized by TF (Yoon *et al.*, 2006; Usman and Mohamed 2009), in this study metals in shoots of *S. maritimus* and *P. australis* were lower than half that of root comparing with other plants. The relatively low accumulation of heavy metals in shoot tissues was probably due to the need of plants to prevent toxicity to the photosynthetic apparatus as suggested by other authors (Stoltz and Greger, 2002; Bragato *et al.*, 2006). The root of plant species are mainly responsible for heavy metal phytoextraction and plants species with TF<1 have the potential for phytostabilization (Yoon *et al.*, 2006; Usman *et al.*, 2012), because in this process the metal tolerant plant species immobilize heavy metals through absorption and accumulation by roots, adsorption onto roots or precipitation within the rhizosphere (Wong, 2003). This process also decreases metal mobility and reduce the likelihood of metals entering into the food chain. Therefore, the use of metal tolerant native flora represents an inexpensive long-term solution (Ashraf *et al.*, 2011).

Nouri *et al.* (2011) in their study showed plant species with TF<1 are useful for phytoremediation of contaminated soil with magnesium, iron, copper and zinc. *P. australis* and *S. maritimus* among the 11 plant species

investigated showed BCF values >1 and TF values <1, indicating strong potential for use in phytostabilization. *P. australis* and *S. maritimus* having the highest BCF and the lowest TF were the most promising plants species for Zn, Cu and Cr phytostabilization. Conversely, plants which have low metal bioaccumulation in their roots and high TF, might be useful for phytoextraction because it would likely increase metal bioaccumulation in its shoots. Overall, we found that *P. australis* and *S. maritimus* were the best for phytostabilization of Zn, Cu and Cr and 9 other plant species could be useful for phytoextraction of metals.

Results of this research work indicated that among the 11 sampled plant species, metal translocation into shoots appears to be very restricted in *S. maritimus* and *P. australis* so that these plant species could be used in order to stabilize the metals and reduce the metal dispersion through wind erosion and grazing animals. Further studies would be needed on investigating deeply the possible translocation of metals to tissues of plants and investigate the phytoremediation efficacy of identified plant species against various concentrations of Zn, Cu and Cr.

Acknowledgments

The authors express their thanks to University of Tehran for its facilities.

Literature Cited

- Abdel-Ghani, N. T., Hefny, M., and El-Chagbaby, G. A. F., 2007. Removal of lead from aqueous solution using low cost abundantly available adsorbents. *Jour. Envir. Sci. Tech.* 41: 67-73.
- Alloway, B. J., 1990. Heavy metals in soils. Glasgow, UK: Blackie and Son. p. 384.
- Alloway, B. J., Jackson, A. P., and Morgan, H., 1990. The accumulation of cadmium by vegetables grown on soils contaminated from a variety of sources. *Sci. Envir. Jour.* 91: 223-236.
- Antosiewicz, D. M., Escude-Duran, C., Wierzbowska, E., and Sklodowska, A., 2008. Indigenous plant species with the potential for

- the phytoremediation of arsenic and metals contaminated soil. *Jour. Water Air Soil Pollution*. 193: 197–210.
- Ashraf, M. A., Maah, M. J., and Yusoff, I., 2011. Heavy metals accumulation in plants growing in ex tin mining catchment. *Jour. Envir. Sci. Tech.* 8(2): 401-416.
- Baker, A. J. M., 1981. Accumulators and excluders strategies in the response of plants to heavy metals. *Jour. Plant Nutrition*. 31(4): 643-654.
- Baker, A. J. M., and Brooks, R. R., 1989. Terrestrial higher plants which hyper-accumulate metallic elements. A review of their distribution, ecology and phytochemistry. *Biorecovery*. 1: 81-126.
- Black, C. A., 1965. Methods of soil chemical analysis and microbiological properties. *Agronomy No. 9*. American Society of Agronomy, Madison.
- Bodar, C. W., Pronk, M. E., and Sijm, D. T., 2006. The European Union risk assessment on zinc and zinc compounds: the process and the facts. *Jour. Integ. Envir. Asses. Manag.* 1: 301–319.
- Bonanno, G., Lo Giudice, R., 2010. Heavy metal bioaccumulation by the organs of *Phragmites australis* common reed and their potential use as contamination indicators. *Jour. Ecol. Indicators*. 10: 639–645.
- Bower, C. A., and Hatcher, J. T., 1966. Simultaneous determination of surface area and cation-exchange capacity. *Soil Sci. Soc. Am. Proc.* 30: 525-527.
- Bragato, C., Brix, H., and Malagoli, M., 2006. Accumulation of nutrients and heavy metals in *Phragmites australis* (Cav.) Trin. ex Steudel and *Bolboschoenus maritimus* (L.) Palla in a constructed wetland of the Venice lagoon watershed. *Jour. Envir. Pollution*, 144: 967–975.
- Chehregani, A., Noori, M., and LariYazdi, H., 2009. Phytoremediation of heavy metal polluted soils: Screening for new accumulator plants in Angouran mine Iran and evaluation of removal ability. *Jour. Ecotoxi. Envir. Safe.* 72: 1349–1353.
- Du Laing, G., Tack, F. M. G., and Verloo, M. G., 2003. Performance of selected destruction methods for the determination of heavy metals in reed plants *Phragmites australis*. *Jour. Analyt. Chim. Acta.* 49: 191–198.
- Fitzgerald, E. J., Caffrey, J. M., Nesaratnam, S. T., and Mc Loughlin, P., 2003. Copper and lead concentrations in salt marsh plants on the Suir Estuary, Ireland. *Jour. Envir. Pollution*, 123: 67-74.
- Fotakis, G, and Timbrell, J. A., 2006. Role of trace elements in cadmium chloride uptake in hepatoma cell lines. *Jour. Toxicol letters.* 164: 97-103.
- Ghosh, M., and Singh, S. P., 2005. A review on phytoremediation of heavy metals and utilization of its byproducts. *Appl. Jour. Ecol. Envir. Res.* 3: 1-18.
- Greger, M, and T, Landberg. 1999. Use of willow in phytoremediation. *Inter. Jour. Phyto.* 12: 115-123.
- Hansel, C., Fendorf, S., Sutton, S., and Newville, M., 2001. Characterization of Fe plaque and associated metals on the roots of mine-waste impacted aquatic plants. *Jour. Envir. Sci. Tech.* 35: 3863-3868.
- Hernandez-Ochoa, I., Garcia-Vargas, G., Lopez-Carrillo, L., Rubio-Andrade, M., Moran-Martínez, J., Cebrian, M. E., and Quintanilla-Vega, B., 2005. Low lead environmental exposure alters semen quality and sperm chromatin condensation in northern Mexico. *Reprod. Toxicol.* 20: 221–228.
- Joner, E. J., and Leyval, C., 2001. Influence of arbuscular mycorrhiza on clover and ryegrass grown together in a soil spiked with polycyclic aromatic hydrocarbons. *Jour. Mycorrhiza.* 10: 155–159.
- Kabata-Pendias, A., 2001. Trace elements in soils and plants. 3rd ed. LLC, Boca Raton, FL: CRC Press.
- Khan, S., Ahmad, I., Tahir Shah, M., Rehman, S., and Khaliq, A., 2009. Use of constructed wetland for the removal of heavy metals from industrial wastewater. *Jour. environ manage.* 90(7): 3451-3457.
- Lasat, M. M., 2002. Phytoextraction of toxic metals, a review of biological mechanisms. *Jour. Envir. Quality.* 31: 109–120.
- Loeppert, R. H., and Suarez, D. L., 1996. Carbonate and gypsum. In *Methods of Soil Analysis, part 3, Chemical Methods*, Bigham JM and Bartels JM ed. SSSA, ASA, Madison, Wisconsin, Etatus-Unis, 437-474.
- Mattina, M. J., Lannucci-Berger, W., Musante, C., and White, J. C., 2003. Concurrent plant uptake of heavy metals and persistent organic pollutants from soil. *Jour. Envir. Pollution.* 124: 375-378.

- Nouri, J., Lorestani, B., Yousefi, N., Khorasani, N., Hasani, A. H., Seif, F., and Cheraghi, M., 2011. Phytoremediation potential of native plants grown in the vicinity of Ahangaran lead-zinc mine Hamedan, Iran. *Jour. Envir. Earth Sci.* 62(3): 639-644.
- Ok, Y. S., Usman, A. R. A., Lee, S. S., Abd El-Azeem, S. A. M., Choi, B. S., Hashimoto, Y., and Yang, J. E., 2011. Effects of rapeseed residue on lead and cadmium availability and uptake by rice plants in heavy metal contaminated paddy soil. *Jour. Chemosphere.* 85: 677-682.
- Rowell, D. L., 1993. *Soil Science: Methods and Applications*. Longman, Harlow, Essex.
- Salt, D. E., Blaylock, M., Kumar, N. P. B. A., Dushenkov, V., Ensley, B. D., Chet, I., and Raskin, I., 1995. Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. *Jour. Biotechnology.* 13: 468-474.
- Santillan, L. F. J., Constantino, C. A. L., Rodriguez, G. A. V., Ubilla, N. M. C., and Hernandez, R. I. B., 2010. Manganese accumulation in plants of the mining zone of Hidalgo, Mexico. *Biores. Tech.* 101 (15): 5836-5841.
- Stoltz, E., and Greger, M., 2002. Accumulation properties of As, Cd, Cu, Pb and Zn by four wetland plant species growing on submerged mine tailings. *Jour. Envir. Exp. Botany.* 47: 271-280.
- Thomas, G. W., 1996. Soil pH and soil acidity. P 475-490. In: Sparks *et al.* (eds.) *Methods of soil analysis*, part 3. Agron. Mongr. 9. 2nd ed. ASA and SSSA, Madison, WI.
- U. S. EPA. 1993. *Cleaning up the nation's waste sites; Market and Technology trends*, office of solid waste and Emergency Response, Technology Innovation office, Washington Dc, EPA, 542-R-92-012.
- Usman, A. R. A., Lee, S. S. L., Awad, Y. M., Lim, K. J., Yang, J. E., and Ok, Y. S., 2012. Soil pollution assessment and identification of hyperaccumulating plants in chromated copper arsenate CCA contaminated sites, Korea. *Jour. Chemosphere.* 872-878.
- Usman, A. R. A., and Mohamed, H. M., 2009. Effect of microbial inoculation and EDTA on the uptake and translocation of heavy metal by corn and sunflower. *Jour. Chemosphere.* 76: 893-899.
- Walkley, A., and Black, I. A., 1934. Review Examination of the Degtjareff method determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science*, 34: 29-38.
- Wang, A., Chunling, L., Renxiu, Y., Yahua, C., Zhenguo, S., and Xiangdong, L., 2012. Metal leaching along soil profiles after the EDDS application-A field study. *Jour. Envir. Pollution.* 164: 204-210.
- Wilkins, D. A., 1978. The measurement of tolerance to edaphic factors by means of root growth. *New Phyto.* 8: 623-633.
- Wong, M. H., 2003. Ecological restoration of mine degraded soils, with emphasis on metal contaminated soils. *Jour. Chemosphere*, 50(6): 775-780.
- Yang, J. E., Lee, W. Y., Ok, Y.S., and Skousen, J., 2009. Soil nutrient bioavailability and nutrient content of pine trees (*Pinus thunbergii*) in areas impacted by acid deposition in Korea. *Jour. Envir. Monit. Assess.* 157: 43-50.
- Yoon, J., Cao, X., Zhou, Q., and Ma, L. Q., 2006. Accumulation of Pb, Cu, and Zn in native plants growing on a contaminated Florida site. *Jour. Sci. total envir.* 368: 456-464.
- Zayed, A., Gowthaman, S., and Terry, N., 1998. Phyto-accumulation of trace elements by wetland plants. I. Duckweed. *Jour. Envir Quality.* 27: 715-721.

ارزیابی تجمع فلزات سنگین در گیاهان رشد یافته در خاک‌های آلوده (مطالعه موردی: قزوین، ایران)

مهديه ابراهيمی الف، محمد جعفری ب، غلامرضا ثواقبی ج، حسین آذرنيوند د، علی طویلی ه، فرناندو مادريد ز

الف استاديار، گروه مرتع و آبخيزداری، دانشگاه زابل، (نگارنده مسئول)، پست الکترونیک: maebrahimi2007@uoz.ac.ir

ب استاد، گروه احیاء مناطق خشک و کوهستانی، دانشگاه تهران

ج دانشيار، دانشکده کشاورزی، دانشگاه تهران

د دانشيار، گروه احیاء مناطق خشک و کوهستانی، دانشگاه تهران

ه دانشيار، گروه احیاء مناطق خشک و کوهستانی، دانشگاه تهران

ز فوق تخصص آلودگی، موسسه IRNAS-CSIC اسپانیا

چکیده. آلودگی محیط‌زیست به فلزات سنگین نگرانی جدی است که مربوط به فعالیت‌ها انسانی است. مطالعه حاضر به منظور تعیین میزان تجمع فلزات در گونه‌های گیاهی در منطقه صنعتی لیا (قزوین، ایران) و ارزیابی قابلیت پالایندگی گونه‌های بومی رشد یافته در خاک‌های آلوده انجام گرفت. نمونه‌برداری از خاک و پساب در سه سایت به صورت شعاعی به فواصل ۳۰۰ متر از نقطه خروجی پساب انجام شد. در هر نقطه در طول ترانسک‌های ۱۰۰ متری در پلات‌های به ابعاد ۵ در ۲ متر، نمونه‌های گیاه و نمونه‌های خاک از عمق ۰-۲۰ و ۲۰-۴۰ سانتی‌متری از منطقه ریزوسفری ریشه برداشت شد. غلظت مس، روی و کروم در ریشه و اندام هوایی ۱۱ گونه گیاهی، خاک و پساب در سه سایت اندازه‌گیری شد. فاکتور تجمع و فاکتور انتقال جهت تعیین پتانسیل گیاه‌پالایی اندازه‌گیری شد. نتایج نشان داد که غلظت فلزات در خاک و پساب بیشتر از آستانه تعیین شده است. غلظت فلزات در نمونه‌های خاک به ترتیب $Cr > Zn > Cu$ و در پساب به ترتیب $Zn > Cr > Cu$ بود. *Scirpus maritimus* L. و *Phragmites australis* (Cav.) Trin. ex Steudel حداکثر مقدار روی، مس و کروم را در ریشه داشتند که مناسب برای گیاه تثبیتی (با حداکثر BCF و حداقل TF) بودند. حداکثر مقدار روی (۷/۲۴ و ۳/۲۹ میلی‌گرم در کیلوگرم به ترتیب برای اندام هوایی و ریشه، $BCF=0.07$) و مس قابل استخراج (۲/۵۶ و ۲/۸۰ میلی‌گرم در کیلوگرم به ترتیب برای اندام هوایی و ریشه، $BCF=0.14$) مربوط به *Hordeum glaucum* L. بود. همچنین، حداکثر مقدار کروم در *Taraxacum officinale* L. تعیین شد. نتایج نشان داد که گیاهانی که حداکثر فاکتور تجمع در ریشه و حداکثر TF را دارند نقش مهمی در برداشت فلزات در فرآیند گیاه استخراجی دارند.

کلمات کلیدی: گیاه‌پالایی، فلزات سنگین، BCF، TF، منطقه صنعتی لیا