



Biomarker application in the recognition of the geochemical characteristics of crude oils from the five depobelts of the Niger Delta basin, Nigeria

Timothy Chibuikwe Anyanwu*¹, Bassey Offiong Ekpo², Boniface Aleruchi Oriji³

1. African Centre of Excellence, World Bank Centre for Oilfield Chemicals Research, University of Port Harcourt, Nigeria

2. Exploration Research and Services Section, Research & Development, NNPC, Port Harcourt, Nigeria

3. Institute of Petroleum Studies, University of Port Harcourt, Rivers State, Nigeria

Received 11 Feb 2021; accepted 17 November 2021

Abstract

Biomarker fingerprints of crude oils are useful indicators of origin of organic matter input in source rocks and depositional conditions which are useful indices for petroleum systems development within a hydrocarbon producing horizon. Twenty five (25) crude oil samples from the five depobelts of the Niger Delta basin, Nigeria were studied to describe their biomarker fingerprints, provide information on the origin of organic matter input in the source rock(s), determine depositional environmental conditions and thermal maturity of the crude oils. The study was based on biomarkers (steranes and hopanes), normal alkanes and acyclic isoprenoids (pristine and phytane) obtained from the gas chromatography–mass spectrometry (GC-MS) analyses performed on the saturated fractions of the crude oils. The results of pristine/phytane (Pr/Ph) ratios, Pr/n-C₁₇ ratios, Ph/n-C₁₈ ratios, C₂₉/C₂₇ sterane ratios, sterane/hopane ratios, %C₂₇, %C₂₈ and %C₂₉ regular steranes, oleanane index, waxiness index, Ts/Tm ratios and Carbon Preference Index (CPI) values, indicated that the analyzed crude oils belong to the same family of oil and originated from terrigenous clastic source rock (s) containing land plant organic matter with minor marine organic matter input, deposited under oxic to sub-oxic paleoenvironmental conditions. The Pr/n-C₁₇ versus Ph/n-C₁₈ cross plot indicated marine algal type II and a mixed type II/III kerogen. The C₃₂H:22S/(22S+22R) values together with Ts/(Ts + Tm), C₂₉:20S/20S + 20R, oleanane index and CPI values indicated that most of the crude oils have reached thermal equilibrium with high thermal maturity levels.

Keywords: Niger Delta, depobelts, source rocks, organic matter, Biomarker

1. Introduction

The Niger Delta (Fig 1) is a hydrocarbon province with an estimated recovery of about 40 billion barrels of crude oil and over 40 trillion cubic feet of gas reserves (Adegoke et al. 2017). It is the major oil bearing province in Africa, making up about 70% of the total hydrocarbon reserves of the entire sub-Saharan Africa (Reijers 2011; Adegoke et al. 2017). The Niger Delta is the Cenozoic offlapping siliciclastic succession situated in the Gulf of Guinea, central part of West Africa (Fig 2). The delta developed on top of the Cretaceous marine strata of the Anambra Basin (Reijers et al. 1997 Corredor et al. 2005; Adegoke et al. 2017), as a major embankment that extended outwards in the process of subsidence and cooling of the oceanic crusts resulting from the separation of the African and the South American plates (Adegoke et al. 2017). The whole of the sedimentary wedge in the Niger Delta were laid in five discrete offlapping siliciclastic sedimentation cycles of fluvio-marine systems known as depobelts. The mechanisms of sedimentation in these depobelts have been described using the escalator regression model (Knox and Omatsola 1989). These depobelts are self-contained entities with respect to hydrocarbon distribution, stratigraphy and structural framework (Adegoke et al. 2017).

The stratigraphic successions in each of these depobelts begins with marine clays and ends with coarse grained sands both vertically and laterally basinward. The presence of major regional faults makes stratigraphic correlation within these depobelts difficult. Geochemical studies in the Niger Delta indicates that the hydrocarbons originated from land plants and other structureless organic matters of Type III Kerogen in Late Eocene to Pliocene aged rocks, deposited in the outer neritic to bathyal environments (Bustin 1988; Onyia et al. 2002). A recent study (Anyanwu et al. 2021) identified subtle geochemical differences among crude oils from the coastal and offshore Niger Delta, where the coastal crude oils appeared to be sourced from more of planktonic/bacterial organic materials, the offshore oils originated mainly from planktonic/land plants organic materials. Ekpo et al. (2018) using biomarkers from twenty-four crude oil samples from western offshore Niger Delta showed that the oils originated from same source rocks, deposited under an oxic paleoenvironmental condition with a mixture of marine and terrigenous organic inputs. Onojake et al. (2013) reported that crude oil samples from two fields in the Niger Delta originated from terrestrial organic sources and deposited in an oxygenated environment. Akinlua and Ajayi (2009) investigated the origin of crude oils from the central Niger Delta and indicated that the oils originated from organic matter of both terrestrial and marine sources deposited in an oxidized environment.

*Corresponding author.

E-mail address (es): anyanwutimothy@gmail.com

Similarly, Sonibare et al. (2008) analyzed ten crude oil samples from the onshore and offshore Niger delta and concluded that the oils originated from a mixed origin (marine and terrestrial kerogen). A detailed understanding of the petroleum systems within a hydrocarbon producing basin is an important aspect of petroleum exploration. Geochemical studies indicate that the Niger Delta petroleum belongs to one family (Bustin 1988; Onyia et al. 2002; GhasemShirazi et al. 2014; Sharifi Teshnizi et al. 2021), of a single petroleum system (Ekweozor et al. 1979). There is need to study the

petroleum system in each of the five depobelts of the Niger Delta in order to ascertain if there are subtle geochemical differences and relationships. In this study, standard oil geochemical parameters were used to characterize crude oil samples from the five depobelts of the Niger Delta (Fig 2). The objective was to describe the biomarker fingerprints in the crude oils in order to provide information on the origin of organic matter input in the source rock(s), determine the depositional environmental conditions and thermal maturity of the crude oils.

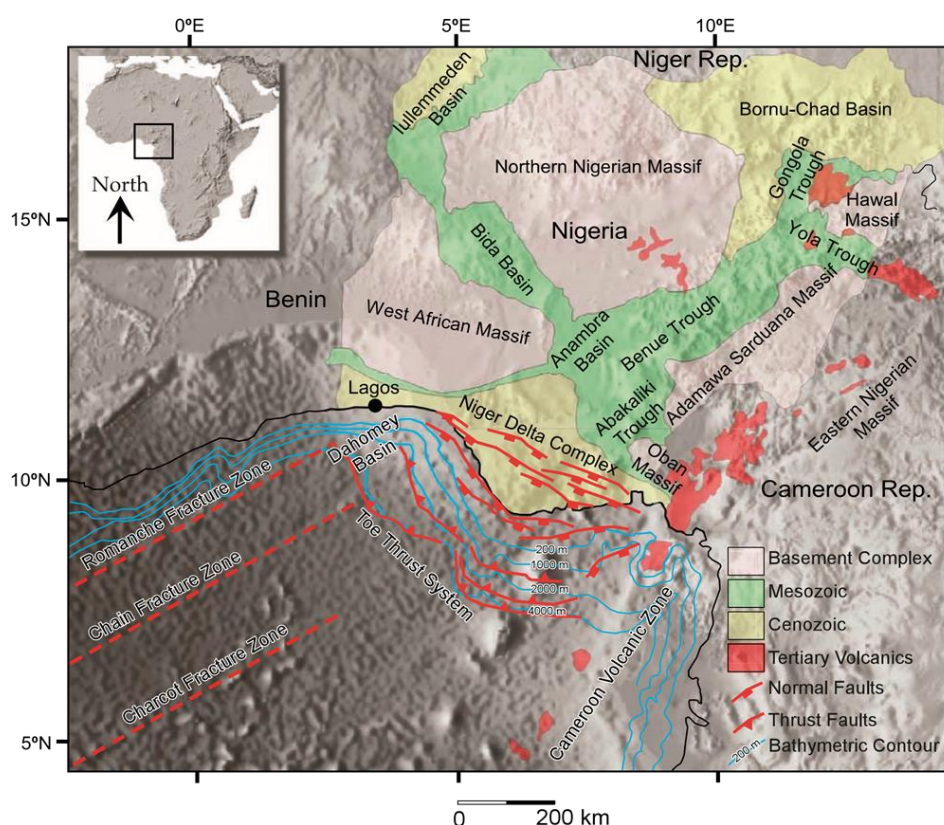


Fig 1. Regional location map of the Niger Delta showing the major sedimentary basins, tectonics and structural features: The topographic and bathymetric features are indicated as (shaded relief) gray-scale image (Corredor et al. 2005).

2. Geology of the Niger Delta basin

The break-up of Gondwana supercontinent is the fundamental events that lead to the evolution of the Niger Delta basin. The origin of this basin is linked to the same processes that formed the Benue Trough, which evolved from the failed arm of rift Triple Junction in the late Jurassic when convection currents in the asthenosphere caused the break-up of the Gondwana Supercontinent (Burke 1972; Olade 1975). The Benue Trough is a conglomerate of series of pull-apart rifts, resulting from sinistral wrench faulting, structural readjustments (that occurred in the Santonian as a result of dissimilarities in the rate of Atlantic spreading in the Mesozoic) (Burke 1972; Genik 1993). This thermo-tectonic events lead to the folding, uplift and changes of depositional axis in the southern part of the Benue Trough. This resulted to subsidence in the west, south and southeast of the

Abakaliki Anticlinorium and the prolongation of the Anambra Basin towards the Calabar area (Adegoke et al. 2017). At this time Anambra Basin was filled with Campanian to Early Paleocene sediments, as subsidence continued in the southern parts of the Benue Trough due to thermal contraction of the lithospheric plates (Turcotte 1977; Onuoha 1981), the Niger Delta Basin evolved (Adegoke et al. 2017; Jehangir Khan et al. 2021). The Benue Trough, the Anambra Basin, and the Niger Delta Basin are a conglomerate of basins stacked vertically in the southern part, and constrained lateral by the fracture zone positions (Fig 3; Fig 4; Fig 5).

The stratigraphy of the Niger Delta basin is made up of the Cretaceous to Holocene clastic sediments which overlie oceanic crust and some remnants of the continental plate (Fig 6). Although the Cretaceous strata can only be inferred from the exposed lithological units

of the closest basin, which is the Anambra Basin, because these strata have not been penetrated beneath the Niger Delta (Reijers et al. 1997; Corredor et al. 2005). Stratigraphy of the Tertiary Niger Delta is subdivided into three main Formations, these Formations represents depositional environments that are prograding (Short and

Stauble 1965; Doust and Omatsola 1990; Kulke 1995). The Akata Formation occupies the basal part of the Tertiary Niger Delta. This Formation is marine in origin, with a thickness that ranges from 2000m at the distal end of the delta to about 7000m underneath the continental shelf (Doust and Omatsola 1990; Corredor et al. 2005).

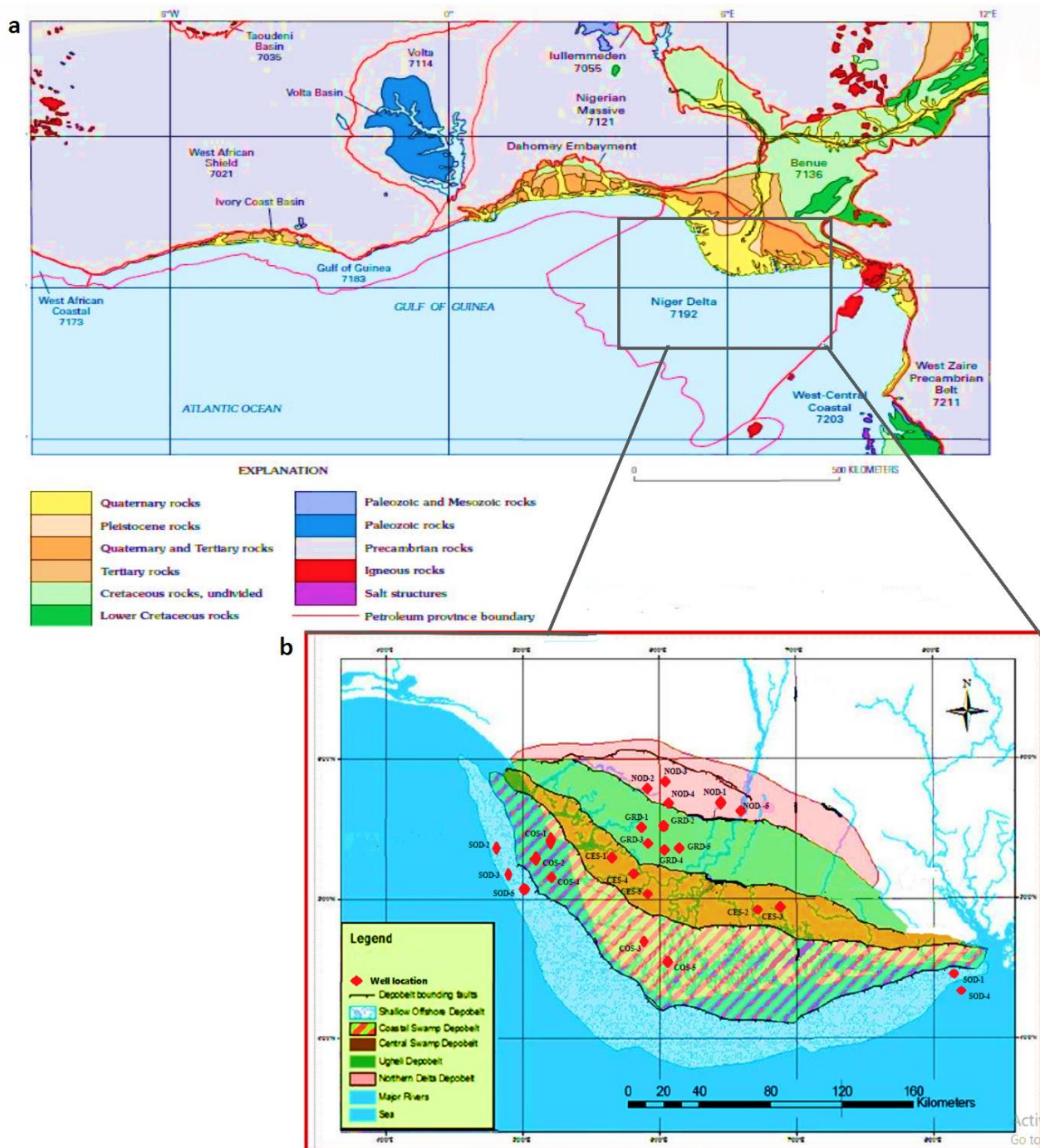


Fig 2. Location of the study area: A. Generalized geologic map of the Gulf of Guinea and surrounding areas (Brownfield and Charpentier 2006); B. Outline map of the Niger Delta. Red Stars show the locations of the twenty five wells in the five depobelts from which crude oil samples were collected for this study (modified from Okosun and Osterloff 2013).

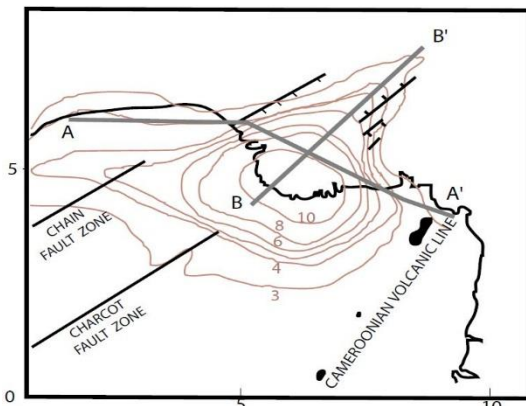


Fig 3. An extension of the Fracture Zones (Chain and Charcot Fracture Zones) into African Plate (Adegoke et al. 2017).

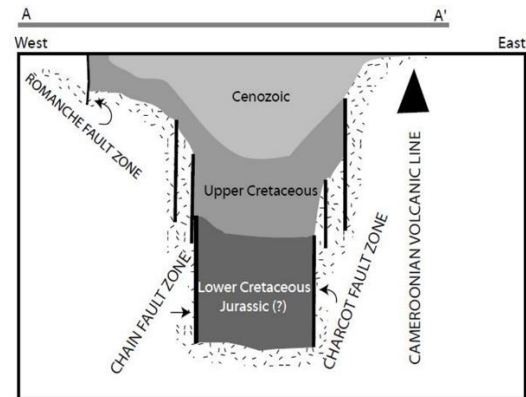


Fig 4. Cross section AA' in Fig 3 showing the stacking pattern of basins in southern Nigeria (the southern Benue Trough, Anambra Basin, and the Niger Delta Basin) (Adegoke et al. 2017).

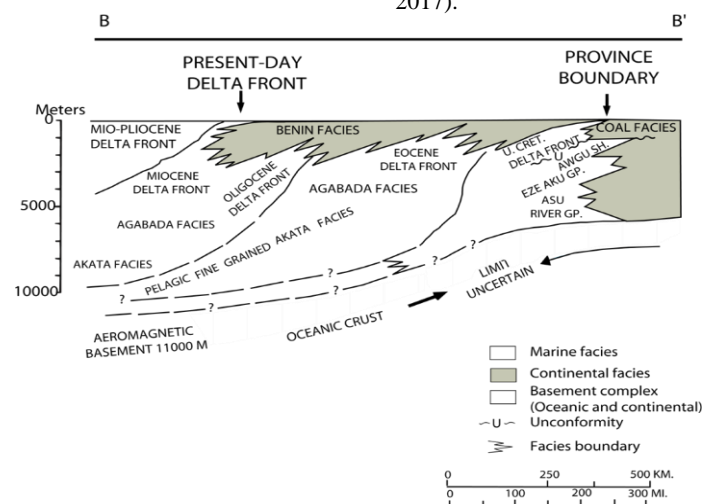


Fig 5. Northeast to Southwest cross section (BB' section in Fig 3) showing the vertical and lateral lithofacies successions starting from the southern Benue Trough, through the Anambra Basin to the Niger Delta (Adegoke et al. 2017).

The Akata Formation has a thickness of about 5000m in the deep-water fold and thrust belts, due to repetitions of structures in this zone (Bilotti et al. 2005; Corredor et al. 2005). This Formation is made up of thick sequences of shales believed to be the major source rocks with minor amounts of turbidite sands representing possible reservoirs in deeper water regions. The Akata Formation generally lacks internal reflections on seismic sections (Fig 6). Overlying the Akata Formation, is the Agbada Formation which represents the major petroleum-bearing portion in the Niger Delta. This Formation ranged from Eocene to recent, and is made up of paralic clastic strata of more than 3500m thick, representing the true deltaic unit of the sequence (Corredor et al. 2005). The Benin Formation overlies the Agbada Formation, and it is made up of late Eocene to recent (Holocene) deposits of continental origin. This includes the alluvial and coastal-plain sediments of up to 2000m in thickness (Avbovbo 1978; Corredor et al. 2005).

During the Paleocene to Earliest Eocene, marine shales accumulated over much of the Niger Delta resulting to

the deposition of the Early – Middle Eocene coarse grained materials of the Agbada Formation in the Northern depobelt while the marine Akata Formation shales were deposited further offshore (Reijers et al. 1997). Paleontological studies of well samples from the Northern depobelt indicated normal marine saline waters, oxygenated conditions and normal temperature conditions of sediment deposition (Ukpong and Anyanwu 2018a; Ukpong and Anyanwu 2018b; Salari and Yazdi 2017). The development of syn-sedimentary growth faults close to the coastlines enabled sands and fines to be trapped along the coast resulting in the progradation of the delta during the Oligocene and Earliest Miocene. The deposition of sediments was very much affected by the long shore currents which helped in the distribution of the sediments and the formation of a thick clastic sequence referred to as the Greater Ughelli depobelt (Reijers et al. 1997), having an abundance composition of the siliciclastic sediments of the Agbada Formation deposits (Ukpong et al. 2017).

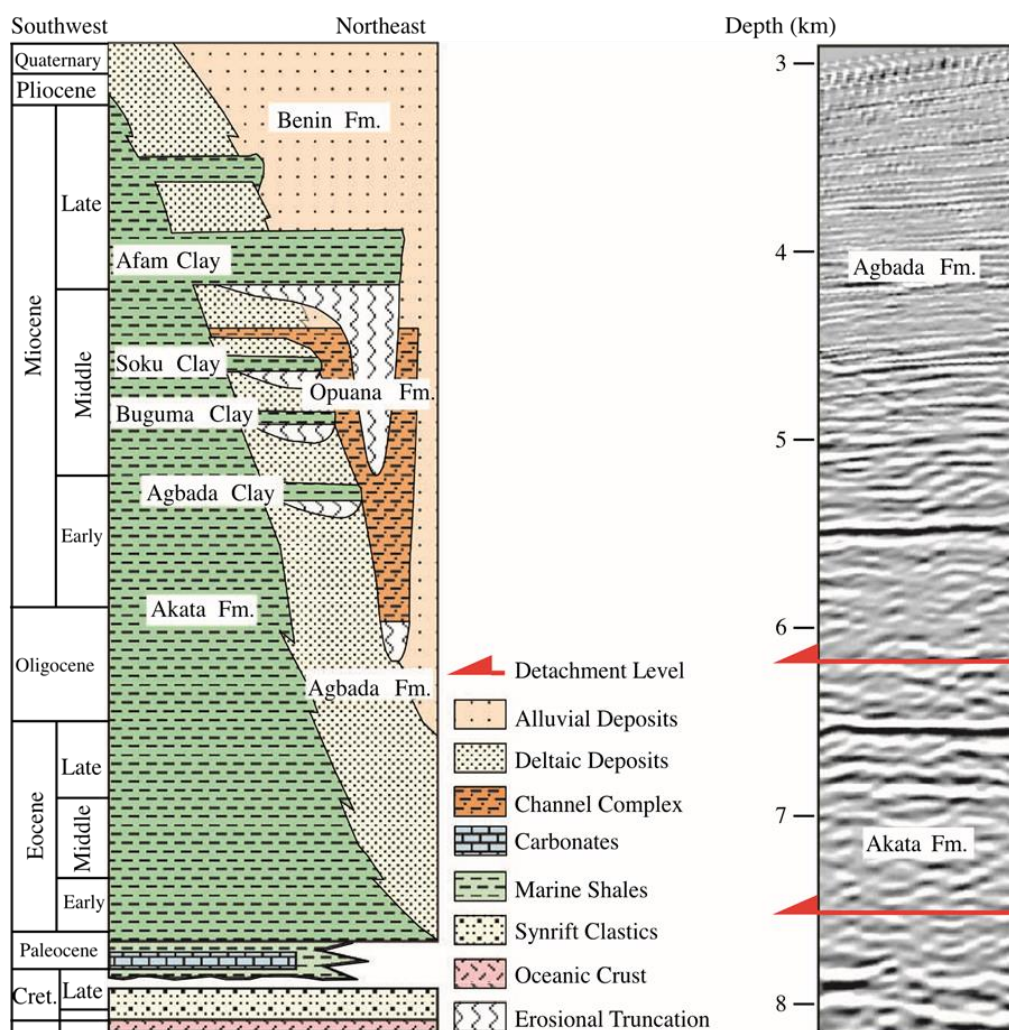


Fig 6. Diagrammatic description of the regional stratigraphy of the Niger Delta basin and the variable density-seismic display of the major stratigraphic sections (Corredor et al. 2005).

The paleohigh of the Abakaliki uplift which was active during this time separated the developing Niger Delta into different units each with its own characteristic paleodrainage pattern resulting to sediments deposition in the Central Swamp and the northern part of the present-day Coastal Swamp depobelts (Reijers et al. 1997). The Coastal Swamp and Shallow Offshore depobelts were formed during periods of steady delta progradation in the Late Miocene to Pliocene ages (Reijers et al. 1997).

3. Materials and methods

A suit of twenty five (25) crude oil samples from twenty five (25) oil producing wells were randomly collected from the five depobelts of Niger Delta basin (Fig 2), for this study. The crude oils were sampled from the wellheads under atmospheric conditions. Fractionation of the crude oil samples into saturates, aromatics and NSOs was carried out following standard procedures involving column chromatography with hexane, toluene and methanol respectively (Eneogwe et al. 2002). Evaporation of solvents were done in a stream of

nitrogen. Gas chromatography-mass spectrometry (GC-MS) analyses were carried out on the saturated hydrocarbon fractions by using an Agilent HP5 ms column. The oven operating temperature condition for the GC MS was held at 55 °C for 1 min, 25°C/min. to 320°C held for 35 min. Sample injection was by splitless mode at an injector temperature of 290 °C, the electron ionization mode was 70 eV. The carrier gas was helium (He) at constant flow condition. The EI Scan mode was used to analyse the samples with source and quadrupole temperatures of 350°C and 150°C respectively. The data was processed using Chem Station G1701BA software, peak integration was done using the RTE integrator. Peak identification was based on retention times and comparison with set standards, literatures and elution order of compounds (Peters et al. 2005; Wang et al. 2006). The peak area of individual compounds was transferred to an Excel spreadsheet for calculation of ratios, preparation of tables and plotting.

4. Results and Discussion

4.1. n-Alkanes and acyclic isoprenoids

The n-alkanes distribution is very useful in determining source input of organic matter (Duan and Ma 2001). The representative chromatograms ion (m/z 71) of the saturated fractions showing n-alkanes and acyclic isoprenoids distribution are shown on Fig 7. The acyclic isoprenoids in the form of pristane and phytane occur in appreciable amounts in the crude oil samples. The major source of pristane and phytane is the phytol side chain of chlorophyll (a) in phototrophic organisms and bacteriochlorophyll (a) and (b) in purple sulfur bacteria (Powell and McKirdy 1973). Pristane (Pr) and phytane (Ph) ratios less than one ($Pr/Ph < 1$) in crude oil show anoxic source-rock deposition environment, while $Pr/Ph > 1$ show oxic conditions of deposition (Didyk et al. 1978). Very high values of greater than or equal to three (≥ 3) indicate high oxidizing conditions associated with sources from terrestrial organic matter input (ten Haven et al. 1987; Peters et al. 2005). The Pr/Ph ratio for the crude oils from the Northern depobelt ranged from 0.75 to 3.87, 0.99 to 1.79 for Greater Ughelli oils, 0.63 to 2.08 for Central swamp oils, 1.01 to 1.57 for Coastal swamp oils and 0.62 to 4.61 for Shallow offshore oils (Table 1). The distribution of the isoprenoid ratios suggests an oxic to suboxic depositional conditions in all the depobelts. The cross plot of the $Pr/n-C_{17}$ versus $Ph/n-C_{18}$ was used to identify the organic matter contributions and

depositional environments of the crude oils (Fig 8). The $Pr/n-C_{17}$ ratio ranged from 0.12- 12.53 while the $Ph/n-C_{18}$ values ranged from 0.17-8.10 in all the depobelts. The values observed for the Pr/Ph ratios as well as the relationship between $Pr/n-C_{17}$ versus $Ph/n-C_{18}$ observed in almost all the oils from the five depobelts suggest major source input from mixed marine-terrestrial organic matter of Type II/III kerogen and marine algal organic matter of Type II Kerogen deposited under oxic-suboxic paleoenvironmental conditions (Fig 8; Peters et al. 1999). The distribution patterns of the n-alkanes and isoprenoids on the m/z 71 chromatograms (Fig 7) shows biodegradation effect on the crude oils. The conspicuous chromatographic baseline hump represents the unresolved complex mixture (Peters et al. 2005). The sampled crude oils showed varying degrees of biodegradation (Fig 7). Biodegradation of crude oils is a function of bacterial alteration of the crude oil compositions (Peters et al. 2005; El-Sheshtawy et al. 2020). El-Sheshtawy et al (2020) demonstrated the effect of bacterial alteration on two shale samples from the Campanian Oil Shale of Egypt. Intense bacterial alteration of organic materials under sulfidic/anoxic conditions encourages the enrichment of metals such as Molybdenum (Mo), Nickel (Ni), and Vanadium (V) (Farouk et al. 2020), heteroatomic compounds (NSO's) with a corresponding removal of n-alkanes and saturated hydrocarbons (Peters et al. 2005).

Table 1. n-Alkanes and acyclic isoprenoids ratios of the crude oils from the five depobelts of the Niger Delta

SAMPLES	Pr/Ph	Pr/(Pr + Ph)	Pr/n-C ₁₇	Ph/n-C ₁₈	CPI	Waxi. Index
Northern Depobelt						
NOD-1	3.87	0.79	2.95	0.43	1.17	1.51
NOD-2	0.75	0.43	0.19	1.08	1.14	0.76
NOD-3	1.76	0.64	1.12	0.71	1.05	3.23
NOD-4	1.9	0.65	0.74	0.46	1.03	1.67
NOD-5	-	-	-	-	-	-
Greater Ughelli						
GRD-1	0.99	0.5	0.32	0.46	1.1	1.08
GRD-2	1.71	0.63	0.25	0.31	1.1	1.24
GRD-3	1.04	0.51	0.26	0.53	0.99	0.69
GRD-4	1.69	0.59	0.23	0.33	1.11	1.22
GRD-5	1.01	0.53	0.33	0.5	1.13	1.1
Central Swamp						
CES-1	2.08	0.68	0.9	1.2	0.92	0.05
CES-2	0.65	0.4	0.51	2.6	1.02	1.4
CES-3	0.63	0.39	0.5	2.59	1.01	1.41
CES-4	0.98	0.5	1.08	0.41	1.06	0.37
CES-5	2.01	0.66	0.89	1.22	1.03	0.09
Coastal Swamp						
COS-1	1.21	0.55	0.83	1.32	0.99	3.09
COS-2	1.01	0.5	0.54	0.6	0.68	1.31
COS-3	1.23	0.54	0.8	1.29	1.01	2.9
COS-4	1.57	0.61	0.66	0.73	0.93	1.42
COS-5	1.35	0.56	0.82	1.33	0.98	3.1
Shallow Offshore						
SOD-1	0.62	0.38	0.25	9.23	1.21	1.79
SOD-2	2.85	0.74	0.63	0.25	1.24	1.78
SOD-3	1.63	0.62	3.51	1.85	0.87	5.43
SOD-4	4.61	0.82	12.53	0.17	0.87	0.09
SOD-5	0.63	0.41	0.28	8.1	1.22	1.8

*Pr- Pristane, Ph- Phytane

* Pr/(Pr + Ph) - Pristine/ (Pristine + Phytane)

*CPI- Carbon Preference Index = Odd number n-alkanes/even numbered n-alkanes (Bray and Evans, 1961).

*Waxiness Index (Wax. Index) = $\sum (n-C_{21} - n-C_{31}) / \sum (n-C_{15} - n-C_{20})$

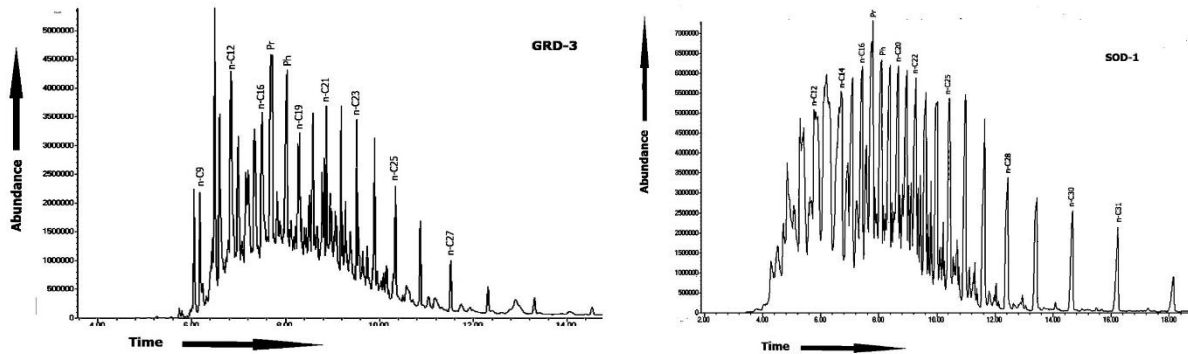


Fig 7. Representative mass chromatograms (71 m/z ion) for the crude oil samples from the five depobelts of the Niger Delta. Pr = pristane and Ph = phytane.

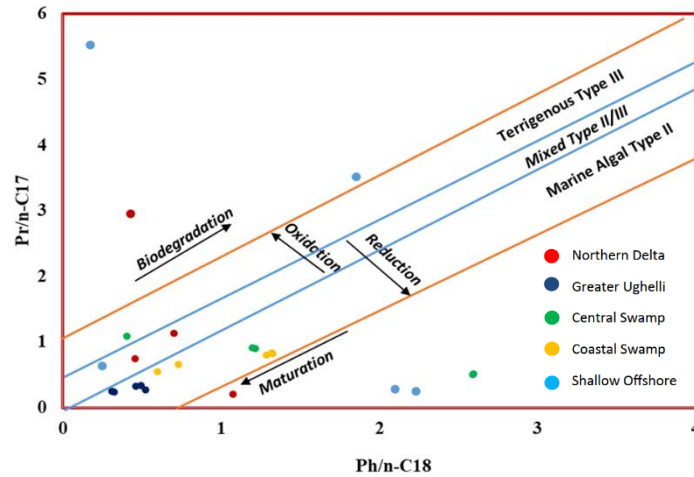


Fig 8. Plot of pristane/n-C17 versus phytane/n-C18 used to infer paleoenvironment and the precursor organic matter in the source rock (Peters et al. 1999). The increase in thermal maturity is shown using the arrow pointing towards the lower left while increasing biodegradation is demonstrated using the arrow pointing to the upper right.

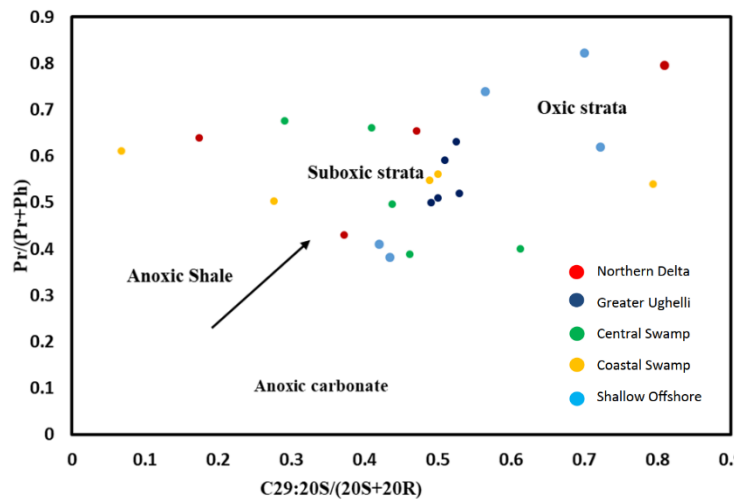


Fig 9. Cross plot of $C_{29}:20S/(20S+20R)$ versus $Pr/(Pr + Ph)$ ratios for the crude oil samples from the five depobelts of the Niger Delta. The arrow show direction of thermal maturation

The Pr/n-C₁₇ versus Ph/n-C₁₈ relationships in the crude oils suggests major terrestrial input. The calculated Pr/(Pr + Ph) values correlated well with the C₂₉: 20S/(20S+20R) values for the Niger Delta oils (Fig 9). The ratio of Pr/(Pr + Ph) will increase with an increase in the clay content (acidity), and is related to the redox potentials of the environment during sediment deposition (Moldowan et al. 1986; Ten Haven et al. 1987; Moldowan et al. 1994).

The calculated Carbon preference index (CPI) values were also used to interpret the depositional environmental conditions (Peters et al. 2005). The Niger Delta oils contain relatively low to moderate ranges of Carbon preference index values. CPI values ranged from 1.03 to 1.17 for the Northern depobelt oils, 0.99 to 1.13 for the Greater Ughelli oils, 0.92 to 1.06 for the Central Swamp oils, 0.68 to 1.01 for the Coastal Swamp oils and 0.87 to 1.24 for the Shallow Offshore oils (Table 1). Cross plot of CPI versus Pr/Ph ratios of the analyzed crude oil samples from the Niger Delta (Fig 10), indicates more oxidizing environmental condition than a reducing condition of deposition (Waples 1983). This agreed with the correlation between pristane/(pristane + phytane) and C₂₉: 20S/(20S+20R) (Fig 9), which indicated an oxic to suboxic paleoenvironmental conditions of deposition (Moldowan et al. 1994). The degree of waxiness which is usually expresses as $\Sigma (n-C_{21} - n-C_{31}) / \Sigma (n-C_{15} - n-C_{20})$ was applied to determine the amount of terrestrial organic matter input in the crude oils source rocks. This technique is based on the idea that land derived organic materials is a major contributor to higher molecular weight n-paraffin in crude oil and therefore can be useful as a source parameter (Hedberg 1968; El Diasty and Moldowan 2012). The Niger Delta oils contains waxy oils in abundance (waxiness >1) (Table 1). The waxiness index ranged from 0.76 to 3.23 for crude oils from the Northern

depobelt, 0.69 to 1.24 for Greater Ughelli oils, 0.05 to 1.491 for Central Swamp oils, 1.31 to 3.10 for Coastal Swamp oils and 0.09 to 5.43 for the Shallow Offshore oils. The cross plot of Pr/Ph ratio versus waxiness index indicates an increasing composition of terrestrial plant organic materials and oxygen rich conditions of deposition for the analyzed crude oil samples (Fig 11).

4.2 Sterane biomarker fingerprints

Steranes measured using GC-MS and monitored with ion (m/z 217+218) are important biomarker groups that are capable of retaining the characteristic structure of precursor organic compounds in petroleum hydrocarbons. Thus, they can yield very important information useful to interpret genetic relationships and depositional conditions of the organic matters in crude oil. The representative GC-MS chromatograms of steranes distribution are shown in Fig 12a to Fig 12c. Table 2 shows the diagnostic biomarker (steranes and hopanes) ratios of the crude oils from the five depobelts of the Niger Delta basin. The peaks identification is shown on Table 3. The steranes are known to be associated with the sterols found in eukaryotic organisms such as algae and higher plants but are absent in the prokaryotes. A study of recent marine and continental sediments showed that the distribution of C₂₇, C₂₈, and C₂₉-sterol homologs on a ternary plots could be used to indicate ecosystems and source input of organic matter in crude oil and source rocks (Huang and Meinschein 1979). The dominance of C₂₇ steranes were used to indicate source input from marine algae while predominance of C₂₉ sterols were used to indicate source inputs from land plant and from cyanobacteria (Huang and Meinschein 1979). As shown in the chromatograms (Fig 12a to Fig 12c), the C₂₇, C₂₈, and C₂₉-sterol homologs are almost of equal proportion in all the analyzed oil samples.

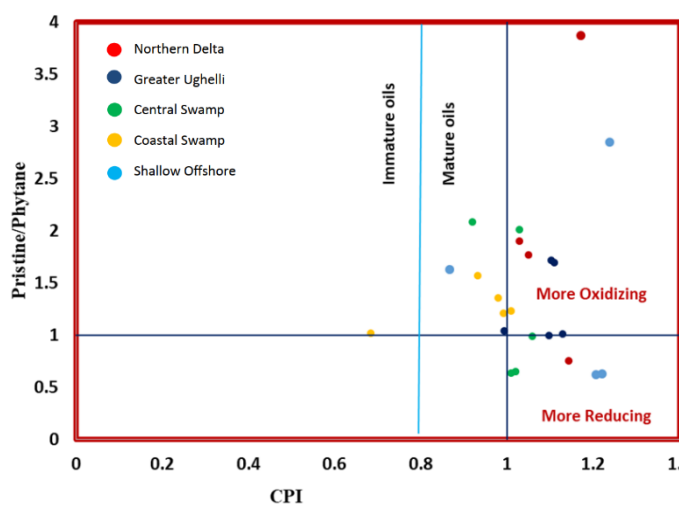


Fig 10. Cross plot of CPI versus Pristine/Phytane ratios for the crude oil samples from the five depobelts of the Niger Delta.

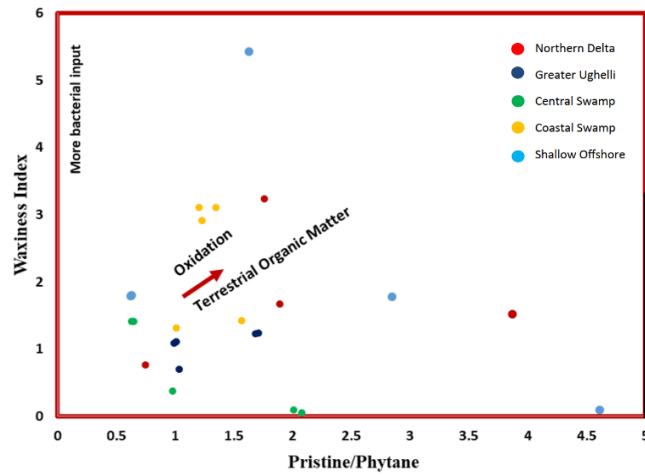


Fig 11. Cross plot of pristane/phytane ratio versus waxiness index (El Diasty and Moldowan 2012).

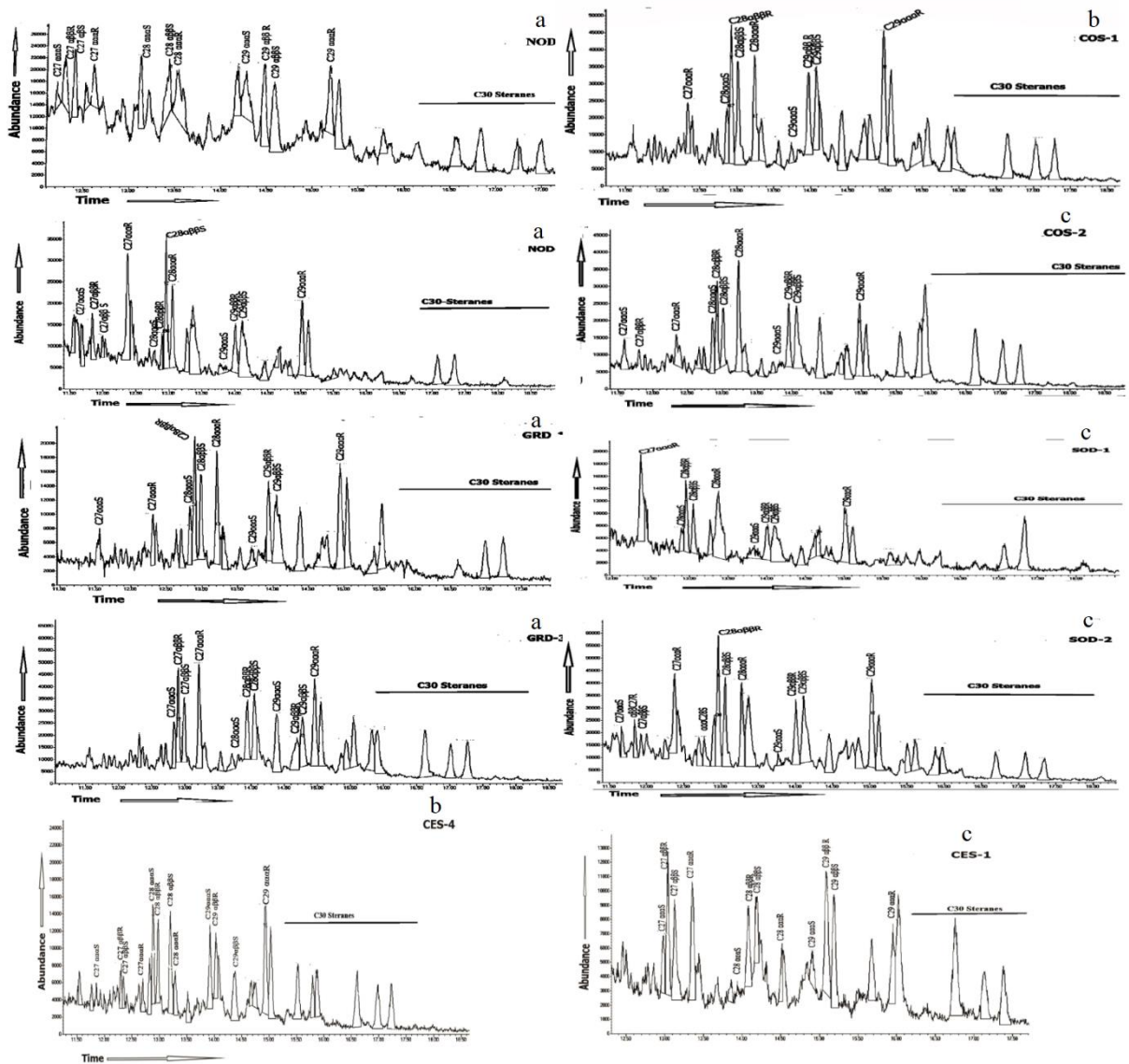


Fig. 12 Representative mass chromatograms 217+218 m/z ion for crude oil samples a) from the Northern (NOD) and Greater Ughelli (GRD) depobelts of the Niger Delta b) from the Central Swamp depobelt (CES) of the Niger Delta c) from the Coastal Swamp (COS) and Shallow Offshore (SOD) depobelts of the Niger Delta

The ternary plot (Fig 13), showed that this crude oils originated from a siliciclastic mixed marine and terrestrial sources, with major organic matter input from eukaryotic organisms (marine algae and land plants) and planktonic bacterial (Huang and Meinschein 1979; Peters et al. 2005). This observation agrees with Ekpo et al. (2018) that used biomarker ratios from twenty-four crude oil samples from western offshore Niger Delta to show that the oils originated from a mixture of marine and terrigenous organic matter input. The distribution of the percentage regular steranes (C_{27} , C_{28} and C_{29}) also show that the crude oils are genetically related, belonging to one source family (Fig 12a to Fig 12c; Table 2).

High values (>1) of $\%C_{29}/\%C_{27}$ sterane ratios are used to indicate origin from land plants (terrigenous environment) while lower values (<1) indicate origin from marine planktonic bacterial (Peters et al. 2005; Peters and Moldowan 1993). The $\%C_{29}/\%C_{27}$ regular sterane ratios ranged from 1.00 to 1.70 for the Northern depobelt oils, 1.12 to 1.53 for Greater Ughelli oils, 1.00

to 3.78 for Central Swamp oils, 0.46 to 1.69 for Coastal Swamp oils and 0.95 to 1.84 for the Shallow Offshore oils (Table 2). These values observed for the $\%C_{29}/\%C_{27}$ regular sterane ratios indicates source organic matter origin from land plants (terrigenous environment), except for some crude oil samples from the Coastal Swamp that showed lower values (>1) for $\%C_{29}/\%C_{27}$ sterane ratios (Table 2). High concentrations of steranes and higher steranes/hopanes ratio (≥ 1) are used to indicate input from marine organic matters of planktonic and benthonic algal origin (Peters et al. 2005). Similarly, low steranes and lower steranes/hopanes ratio (≤ 1) are used to indicate terrestrial input or microbially reworked organic matter (Tissot and Welte 1984; Peters et al. 2005). The ratio of regular steranes/17 α -hopanes observed for the analyzed crude oil samples were very low (≤ 1) (Table 2). This indicates major source input from terrestrial materials or microbial reworked organic matter (Tissot and Welte 1984; Peters et al. 2005).

Table 2. Diagnostic biomarker (Steranes and Hopanes) ratios of crude oils from the five depobelts of the Niger Delta basin

SAMPLES	C28/C29 Steranes	C29/C28 Steranes	C29/C27 Steranes	%C29	%C28	%C27	C29: 20S/ (20S+20R)	Ster./Hop	Ts/ (Ts+Tm)	Ts/Tm	Olea. index	C31H/ C30H	Mor./ C30Hop	C32H: 22S/ (22S + 22R)
Northern Depobelt														
NOD-1	0.94	1.07	1.67	39.4	36.94	23.66	0.81	0.14	0.66	2.28	0.89	0.34	0.21	0.60
NOD-2	0.39	2.57	1.7	50.54	19.65	29.81	0.37	0.06	0.54	1.16	0.39	0.34	0.17	0.58
NOD-3	0.91	1.1	1.28	37.25	33.72	29.03	0.17	0.19	0.7	2.36	0.97	0.33	0.2	0.58
NOD-4	0.7	1.4	1.00	37.0	25.85	37.09	0.4	0.13	0.59	1.41	0.75	0.36	0.13	0.57
NOD-5	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Greater Ughelli														
GRD-1	0.93	1.08	1.51	38.6	35.87	25.53	0.49	0.19	0.54	1.17	0.75	0.36	0.16	0.54
GRD-2	0.79	1.27	1.12	37.34	29.41	33.25	0.53	0.14	0.48	0.91	0.84	0.34	0.16	0.56
GRD-3	0.92	1.06	1.52	40.6	33.87	25.53	0.5	0.18	0.44	0.78	0.71	0.32	0.16	0.59
GRD-4	0.81	1.25	1.19	39.32	27.41	33.27	0.51	0.15	0.49	0.89	0.83	0.34	0.15	0.56
GRD-5	0.9	1.04	1.53	40.5	32.97	26.53	0.52	0.18	0.53	0.16	0.76	0.35	0.16	0.52
Central Swamp														
CES-1	0.52	1.93	1.01	39.9	20.71	39.38	0.29	0.16	0.45	0.81	0.91	0.32	0.16	0.57
CES-2	0.62	1.61	1.00	38.16	23.73	38.11	0.61	0.2	0.83	4.78	0.95	0.34	0.19	0.57
CES-3	0.76	1.31	1.19	38.4	29.22	32.39	0.46	0.36	0.5	0.99	0.64	0.39	0.24	0.57
CES-4	0.56	1.79	3.78	54.81	30.68	14.51	0.44	0.15	0.47	0.88	0.89	0.33	0.17	0.64
CES-5	0.53	1.9	1.03	40.11	19.61	39.37	0.41	0.19	0.48	0.82	0.93	0.35	0.16	0.55
Coastal Swamp														
COS-1	0.58	1.73	0.99	38.64	22.38	38.97	0.49	0.21	0.52	1.08	0.9	0.35	0.19	0.55
COS-2	0.54	1.85	1.13	41.29	22.29	36.42	0.28	0.16	0.43	0.74	0.9	0.34	0.19	0.58
COS-3	1.14	0.88	0.46	23.29	26.58	50.13	0.79	0.51	0.61	1.54	0.7	0.23	0.12	0.61
COS-4	0.45	2.23	1.69	49.03	21.94	29.03	0.07	0.31	0.5	1	0.39	0.33	0.17	0.59
COS-5	0.56	1.69	1.02	40.74	20.38	38.87	0.5	0.22	0.51	1.06	1.01	0.33	0.18	0.56
Shallow Offshore														
SOD-1	1.07	0.93	1.06	33.2	35.57	31.23	0.43	0.09	0.6	1.47	0.43	0.29	0.23	0.60
SOD-2	1.53	0.66	0.95	27.98	42.69	29.33	0.56	0.28	0.52	1.09	0.99	0.29	0.26	0.59
SOD-3	0.53	1.89	1.84	48.23	25.49	26.27	0.72	0.07	0.51	1.06	4.67	0.27	0.17	0.58
SOD-4	0.55	1.91	1.6	50.24	22.48	27.27	0.7	0.09	0.53	1.08	4.76	0.3	0.18	0.56
SOD-5	1.06	0.86	1.08	40.1	28.67	31.33	0.42	0.1	0.55	1.5	0.42	0.3	0.22	0.6

* C_{29} : 20S/(20S+20R) - C_{29} α 20S stigmastane/(C_{29} α 20S stigmastane + C_{29} α 20R stigmastane)

* Ster./Hop. - Regular Steranes /17 α -hopanes

* Ster - Steranes, * Hop- Hopane, * $\%C_{27}$ Steranes, * $\%C_{28}$ Steranes, * $\%C_{29}$ Steranes, *Oleanane index (Olea. Index) - α -oleanane/ C_{30} 17 α (H)-hopane

*Ts/(Ts+Tm):- 18 α (H)-trisorhopane/(18 α (H)-trisorhopane + 17 α (H)-trisorhopane)

*Ts/Tm: 18 α (H)-trisorhopane/17 α (H)-trisorhopane

* $C_{31}H/C_{30}H$ - C_{31} -17 α (H), 21b(H)-30 homohopane (22S+22R)/2/ (C_{30} 17 α (H)-hopane)

*Mor./ $C_{30}Hop$ - C_{30} -17b(H), 21a(H)-moretane/ C_{30} 17 α (H)-hopane

* $C_{29}/C_{30}H$ - C_{29} Tm 17 α (H)21b(H)-norhopane/ C_{30} 17 α (H)-hopane

* $C_{32}H$: 22S/(22S + 22R) - C_{32} -17 α (H), 21b(H)-30 bishomohopane (22S)/ (C_{32} -17 α (H), 21b(H)-30 bishomohopane (22S)+ C_{32} -17 α (H), 21b(H)-30 bishomohopane (22R)

4.3 Hopane (m/z 191) biomarker fingerprints

Just like the steranes, hopanes biomarkers are very important because they retain the characteristic skeletons of the original organic compounds in petroleum and source rocks. The hopanes are monitored on mass chromatogram ion (m/z=191), and are commonly used to determine genetic relationships between crude oils and their source rock depositional environment (Hunt 1996). The mass chromatograms showing the distribution of the hopanes are shown on Fig 14a to Fig 14e. The hopane biomarker ratios (Table 2) suggested source deposition in an oxic to sub-oxic environmental conditions. The hopanes originates from bacteriohopanetetrol and polyfunctional C₃₅ hopanoids of prokaryotic microorganisms (Ourisson et al. 1984; Peters et al. 2005). The ratio of Ts (trisnorneohopane) to Tm (trisnorhopane) more than (0.5) has been found to increase as the shale ratio increases, also Ts/Tm ratios begins to decrease later in the course of thermal maturity (Hunt 1996; Van Grass 1990). The Ts/Tm ratios for the Northern depobelt oils ranged from 1.16 to 2.36, 0.16 to 1.17 for the Greater Ughelli oils, 0.81 to 4.78 for Central Swamp oils, 0.54 to 1.54 for the Coastal Swamp oils and 1.06 to 1.50 for the Shallow Offshore oils. The Ts/Tm ratios for the Niger Delta oils showed high values greater than 0.5 (Ts/Tm > 0.5) for all the crude oil samples, except two oil samples (GRD- 1 and GRD-5) in the Greater Ughelli depobelt. This values indicated thermally matured shale source rocks for the crude oils (Hunt 1996; Van Grass 1990). The oleanane index is expressed in this study as 18 α (H)-oleanane /C₃₀ 17 α (H)-hopane. Moderate values for oleanane index was observed in the crude oils from the Northern depobelt (0.39-0.97), the oleanane index for the Greater Ughelli ranged from 0.71 to 0.84, 0.64 to 0.95 and 0.39 to 1.01 for the Central Swamp and Coastal Swamp respectively, and 0.42 to 4.76 for the Shallow offshore depobelt (Table 2). The abundance of oleanane as well as high oleanane index values have been reported

in the Tertiary Niger Delta crude oils and source rock (Ekweozor et al. 1979; Udo and Ekweozor 1990; Onojake and Abrakasa 2021). This is an indication of an abundance of higher-plant macerals organic matter (Udo and Ekweozor 1990). A cross plot of oleanane index against the biostratigraphic ages of the depobelts showed gradual increase in the oleanane fingerprints from the older depobelts to the younger ones (Fig 15). Oleanane occurrences in crude oils have been widely used to indicate source inputs from higher-plants of Cretaceous and younger ages. This biological markers are derived from betulins and taraxerene (ten Haven and Rullkotter 1988; Grantham et al. 1983), including other pentacyclic triterpenoids (Whitehead 1974). The increasing oleanane fingerprints from the older depobelts to the younger ones appear to have resulted from increasing terrestrial organic matter input and/or thermal maturity of the source rocks (Moldowan et al. 1994). The Ts/(Tm+Ts) ratio is known to be controlled to some extent by the redox potential of the sediments during the time deposition, and it is also dependent on maturity. The Ts is relatively more stable than the Tm, and as such the Ts/(Ts+Tm) ratios increases with an increase in the thermal maturity as demonstrated using the relationship of Ts/(Ts + Tm) ratio and C₂₉ 20S/(20S + 20R) sterane ratio (Fig 16).

The Ts (Ts+Tm) ratios and the C₂₉: 20S/(20S+20R) ratio are maturity indicators, C₂₉: 20S/(20S+20R) ratio increases from 0.0 to about 0.5 (0.52-0.55= attending equilibrium) with continues increase in maturity, this variation in ratio might be a function of isomerization and/or the greater stabilization of the 20S epimer when compared with the 20R epimer (Hunt 1996; Dahi et al. 1999). The C₂₉: 20S/(20S+20R) ratio approaches equilibrium value at/or before the peak of the oil generative window, this is quite distinct from the Ts / (Ts+Tm) ratio which approaches its endpoint around the end of the oil generative window (Peters and Moldowan 1993).

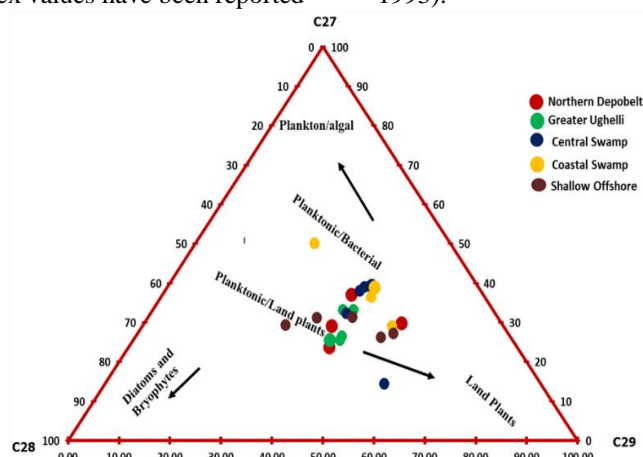


Fig 13. Ternary plot showing the distribution of steranes (C₂₇, C₂₈ & C₂₉) for the analysed crude oils from the five depobelts of the Niger Delta (modified after Huang and Meinschein 1979).

Table 3. Identification of peaks on m/z 191 (hopanes) and m/z 217+218 (steranes) mass chromatograms

Peak	Compound name	Peak	Compound name
Ts	18 α (H)- trisnorneohopane	C27 $\alpha\beta\beta$ R	C ₂₇ 13 β (H) 17 α (H) Diacholestane (20R)
Tm	17 α (H)-trisnorhopane	C27 $\alpha\beta\beta$ S	C ₂₇ 5 α (H) 14 β (H) 17 β (H) cholestane (20S)
$\alpha\beta$ C29Hop	C ₂₉ Tm 17 α (H)21 β (H)-norhopane	C27 $\alpha\alpha\alpha$ R	C ₂₇ 5 α (H) 14 α (H) 17 α (H) cholestane (20R)
C29Ts	C ₂₉ 18 α (H) norneohopane (29Ts)	C28 $\alpha\alpha\alpha$ S	C ₂₈ 5 α (H) 14 α (H) 17 α (H) ergostane (20S)
$\beta\alpha$ C29Diahop	C ₂₉ 17 α (H), 21 β (H)-25-dinorhopane	C28 $\alpha\beta\beta$ R	C ₂₈ 5 α (H) 14 β (H) 17 β (H) cholestane (20R)
α -ole	18 α (H) -oleanane	C28 $\alpha\beta\beta$ S	C ₂₈ 5 α (H) 14 β (H) 17 β (H) cholestane (20S)
$\alpha\beta$ C30Hop	C ₃₀ 17 α (H)-hopane	C28 $\alpha\alpha\alpha$ R	C ₂₈ 5 α (H) 14 α (H) 17 α (H) ergostane (20R)
$\beta\alpha$ C30Mor	C ₃₀ 17 β (H), 21 α (H)-moretane	C29 $\alpha\alpha\alpha$ S	C ₂₉ 5 α (H) 14 α (H) 17 α (H) stigmastane (20S)
$\alpha\beta$ C31SHop	C ₃₁ 17 α (H), 21 β (H)-30 homohopane (22S)	C29 $\alpha\beta\beta$ R	C ₂₉ 5 α (H) 14 β (H) 17 β (H) stigmastane (20R)
$\alpha\beta$ C31RHop	C ₃₁ 17 α (H), 21 β (H)-30 homohopane (22R)	C29 $\alpha\beta\beta$ S	C ₂₉ 5 α (H) 14 β (H) 17 β (H) stigmastane (20S)
$\alpha\beta$ C32SHop	C ₃₂ 17 α (H), 21 β (H)-30 bishomohopane (22S)	C29 $\alpha\alpha\alpha$ R	C ₂₉ 5 α (H) 14 α (H) 17 α (H) stigmastane (20R)
$\alpha\beta$ C32RHop	C ₃₂ 17 α (H), 21 β (H)-30 bishomohopane (22R)		
C27 $\alpha\alpha\alpha$ S	C ₂₇ 5 α (H) 14 α (H) 17 α (H) cholestane (20S)		

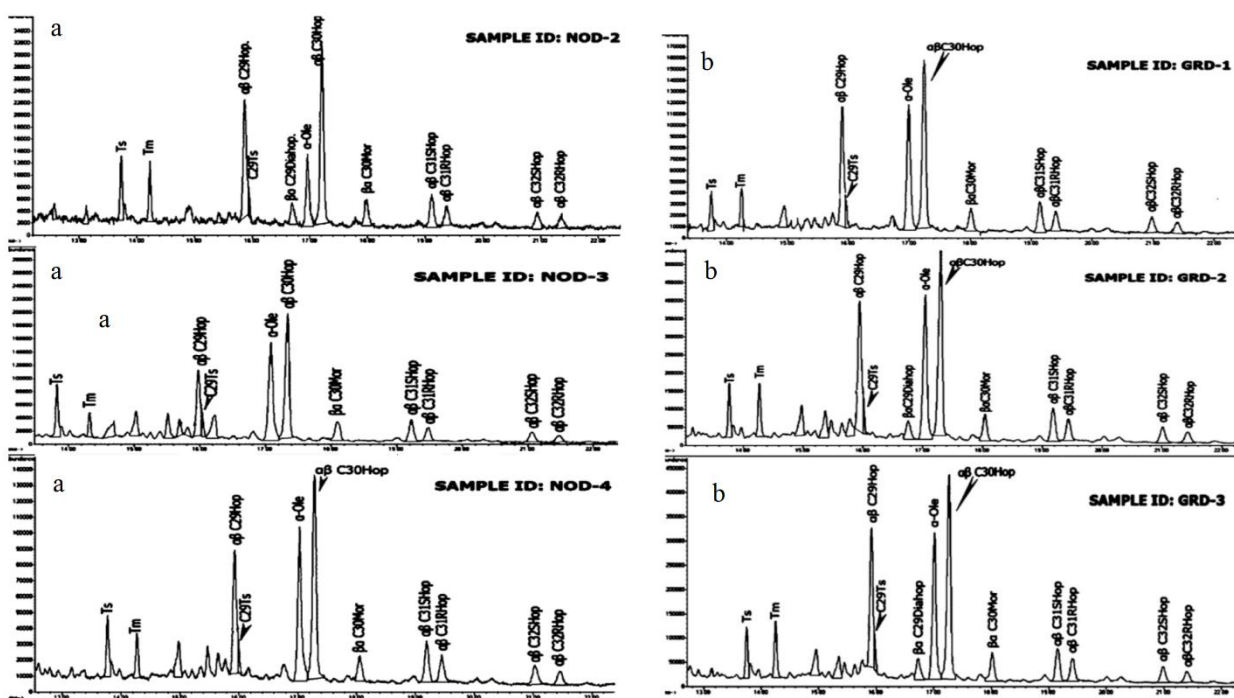


Fig 14. Representative mass chromatograms (m/z 191) showing the distribution of hopanes in the analyzed crude oils from the a) Northern Depobelt of the Niger Delta b) Greater Ughelli depobelt of the Niger Delta

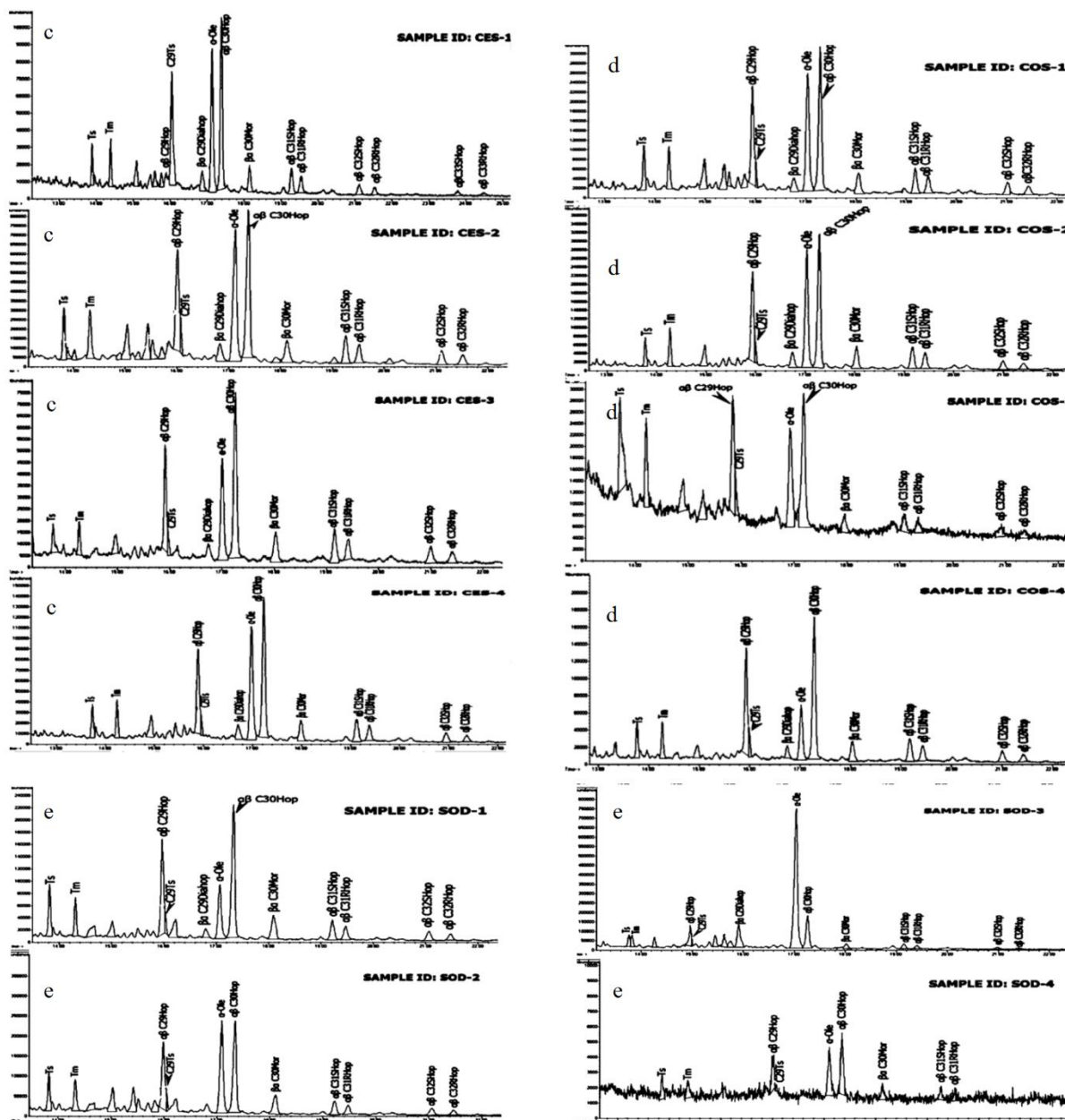


Fig 14. Continued. Representative mass chromatograms (m/z 191) showing the distribution of hopanes in the analyzed crude oils from the c) Central Swamp depobelt of the Niger Delta d) Coastal swamp depobelt of the Niger Delta e) Shallow offshore depobelt of the Niger Delta

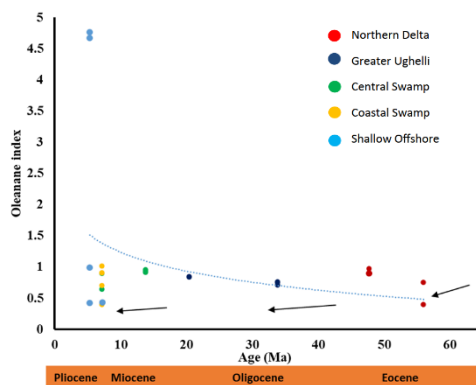


Fig 15. Oleanane index (vertical lines) from the twenty five analyzed representative crude oil samples from the five depobelts of the Niger Delta, horizontal axis indicate the geological/biostratigraphic ages of the depobelts (Reijers et al. 1997); the arrows show the direction of delta progradation and depobelts evolution.

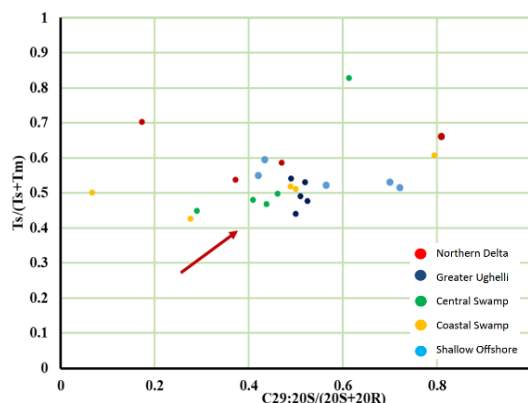


Fig 16. $C_{29} 20S/(20S + 20R)$ regular sterane ratio versus $Ts/(Ts + Tm)$ ratio for the analyzed crude oil samples from the five depobelts of the Niger Delta: The direction of increasing thermal maturity is shown with the red arrow (Hunt 1996).

This means that after the peak point of the oil-generative window, the $C_{29}: 20S/(20S+20R)$ ratio will remain constant, maintaining the equilibrium value, only $Ts/(Ts+Tm)$ ratio will be increasing with the increase in the thermal maturity. The $Ts/(Ts+Tm)$ ranged from 0.54 to 0.70 for the Northern depobelt oils, 0.44 to 0.54 for greater Ughelli oils, 0.45 to 0.83 for Central Swamp oils, 0.43 to 0.61 for Coastal Swamp oils and 0.51 to 0.60 for Shallow Offshore oils (Table 2). The $Ts/(Ts + Tm)$ ratios and $C_{29}:20S/20S + 20R$ ratio values do not show much significant differences across the five depobelts. This might be related to similarities in source organic matters and/or conditions of deposition of these oils (Dahi et al. 1999). The $C_{29} 20S/(20S + 20R)$ sterane ratio for the Northern depobelt oils ranged from 0.17 to 0.81, 0.49-0.53, for the Greater Ughelli oils, 0.29-0.61 for the Central Swamp oils, 0.07 to 0.79 for the Coastal Swamp oils and 0.42 to 0.72 for the Shallow offshore oils (Table

2). Fig 16 shows $Ts/(Ts + Tm)$ ratio versus $C_{29} 20S/(20S + 20R)$ regular sterane ratio of the crude oils. The $Ts/(Ts + Tm)$ values show that some of the oils had already attained thermal equilibrium while some are just close to thermal equilibrium. The $C_{32}H: 22S/(22S+22R)$ hopane ratio also indicated thermal maturity for the crude oils. $C_{32}H: 22S/(22S+22R)$ ratio increases from 0 - 0.6 to reach equilibrium during thermal maturity, and values of 0.50 to 0.54 are indicative of oil generative window (El-Sabagh et al. 2017). Crude oil samples from the Northern depobelt have $C_{32}H: 22S/(22S+22R)$ values that ranged from 0.57 to 0.60, 0.52 to 0.59 for the Greater Ughelli oils, 0.55 to 0.64 for Central Swamp oils, 0.55 to 0.61 for the Coastal Swamp oils and 0.56 to 0.60 for Shallow Offshore oils. These values suggested that most of these crude oils have reached thermal equilibrium with a high thermal maturity level (Fig 17; El-Sabagh et al. 2017).

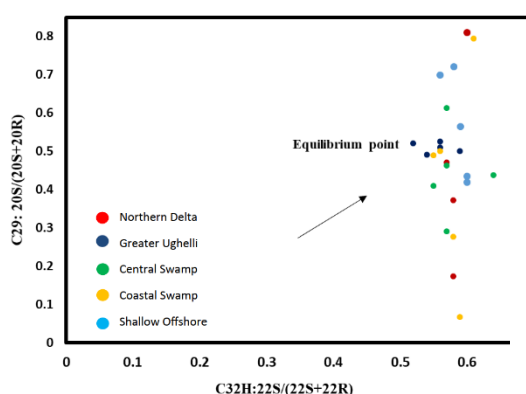


Fig 17. Cross plot of $C_{32}H: 22S/(22S+22R)$ versus $C_{29}: 20S/(20S+20R)$; the arrow show the direction of increasing thermal maturity (El-Sabagh et al. 2017).

5. Conclusions

The results of biomarker characterization by Gas chromatography–Mass spectrometry (GC-MS) indicated that the analyzed crude oils belong to the same family (a single petroleum system) with source organic matter originating from land plants (terrigenous environment) and minor organic matter input from planktonic diatoms/bacterial. Palaeoenvironmental analysis base on (Pr/Ph), Pr/n-C₁₇, Ph/n-C₁₈ ratio, indicated sub-oxic to oxic paleoenvironmental conditions of deposition. The ratios of n-alkanes and the acyclic isoprenoids indicated mainly marine algal type II and a mixed type II/III kerogen for the studied crude oils. Most of the crude oil samples have C₃₂H: 22S/(22S+22R) values that suggested they have reached thermal equilibrium with a high thermal maturity level. This is supported by the Ts/(Ts + Tm), C₂₉:20S/20S + 20R, oleanane index and CPI values. The biomarker characteristics and distribution pattern in the various depobelts can be used as a fingerprints for crude oil and source rocks correlation across the Niger Delta.

Acknowledgements

We sincerely express our gratitude to the African Centre of Excellence, Centre for Oilfield Chemicals Research, University of Port Harcourt Nigeria for the baseline support provided for this research. We equally acknowledge the anonymous reviewers for their constructive criticism and thoughtful comments that were beneficial to the improvement of the revised manuscript. Acknowledgments are due to Matosaab integrated services limited (Environmental & Laboratory Services) Port Harcourt Nigeria for their valuable assistance in carrying out the GC-MS analyses and Oil Company XY for providing the crude oil samples used for this study.

References

- Adegoke OS, Oyebamiji AS, Edet JJ, Osterloff PL, Ulu OK (2017) Cenozoic Foraminifera and Calcareous Nannofossil Biostratigraphy of the Niger Delta. Elsevier Radarweg 29, PO Box 211, 1000 AE Amsterdam, Netherlands.
- Akinlua A, Ajayi TR (2009) Geochemical Characterization of Central Niger Delta Oils, *Journal of Petroleum Geology* 32(4):373-382.
- Anyanwu TC, Ekpo BO, Oriji BA (2021) Geochemical Characterization of the Coastal and Offshore Niger Delta Crude Oils, Nigeria, *International Journal of Advanced Academic Research* 7 (12):73-87.
- Avbovbo AA, (1978) Tertiary lithostratigraphy of Niger Delta, *American Association of Petroleum Geologists Bulletin* 62:295-300.
- Bilotti FD, Shaw JH, Cupich RM, Lakings RM (2005) Detachment fold, Niger Delta, in Shaw JH, Connors C, Suppe J, eds., Seismic interpretation of contractional fault related folds, *AAPG Studies in Geology* 53:103–104.
- Brownfield ME, Charpentier RR (2006) Geology and total petroleum systems of the Gulf of Guinea Province of west Africa, *U.S Geological Survey Bulletin* 2207-C: 32.
- Burke K (1972) Longshore drift, submarine canyons, and submarine fans in development of Niger Delta, *American Association of Petroleum Geologists* 56:1975-1983.
- Bustin RM, (1988) Sedimentology and characteristics of dispersed organic matter in Tertiary Niger Delta: origin of source rocks in a deltaic environment, *American Association of Petroleum Geologists Bulletin* 72:277-298
- Corredor F, Shaw JH, Bilotti F (2005) Structural styles in the deep-water fold and thrust belts of the Niger Delta, *American Association of Petroleum Geologists Bulletin* 89(6): 753–780.
- Dahi J, Moldowan M, Sundaraman P (1999) Relationship of biomarker distribution to depositional environment: Phosphoria Formation, Montana, U.S.A, *Organic Geochemistry* 20(7):1001–1017.
- Didyk BM, Simoneit BRT, Brassell SC, Eglinton G (1978) Organic geochemical indicators of palaeoenvironmental conditions of sedimentation, *Nature* 272:216–22.
- Doust H, Omatsola E (1990) Niger Delta. In: Edwards JD, Santogrossi PA (Eds.), Divergent/passive Margin Basin, *American Association of Petroleum Geologists* 48:239–248.
- Duan Y, Ma L (2001) Lipid geochemistry in a sediment core from Ruoergai Marsh deposit (Eastern Qinghai-Tibet Plateau, China), *Organic Geochemistry* 32:1429-1442.
- Ekpo BO, Essien N, Neji PA, Etsenake RO (2018) Geochemical fingerprinting of western offshore Niger Delta oils, *Journal of Petroleum Science and Engineering* 160:452–464.
- Ekweozor CM, Okogun JI, Ekong DEU, Maxwell JM (1979) Preliminary organic geochemical studies of samples from the Niger Delta (Nigeria): Analyses of crude oils for triterpanes, *Chemical Geology* 27: 11–28.
- El Diasty WS, Moldowan JM (2012) Application of biological markers in the recognition of the geochemical characteristics of some crude oils from Abu Gharadig Basin, north Western Desert Egypt, *Marine and Petroleum Geology* 35:28-40.
- El-Sabagh SM, Ebiad MA, Rashad AM, El-Naggar AY, Badr IHA, El Nady MM, Abdullah ES (2017) Characterization Based on Biomarkers Distribution of Some Crude Oils in Gulf of Suez Area – Egypt, *Journal of materials and Environmental Sciences* 8:1-15.
- El-Sheshtawy HS, Khalil N, Farouk S (2020) Biodegradation Effect on the Campanian Oil Shale of Egypt, *Geomicrobiology Journal* 37(8): 746-752 .
- Eneogwe C, Ekundayo O, Patterson B (2002) Source-derived oleanenes identified in Niger Delta oils, *Journal of Petroleum Geology* 25:83–94.

- Farouk S, Ahmad F, Baioumy H, Lehmann B, Mohammed IQ, Al-Kahtany K (2020). Geochemical characteristics of carbonaceous chalk near the Cretaceous/Paleogene transition, central Jordan: Strong metal enrichment of redox-sensitive and biophile elements from remineralized calcitic plankton, *Marine and Petroleum Geology* 120:104535.
- Genik GJ, (1993) Petroleum geology of Cretaceous-Tertiary rift basins in Niger, Chad and Central African Republic, *American Association of Petroleum Geologists Bulletin* 77:1405–1434.
- GhasemShirazi B, Bakhshandeh L, Yazdi A (2014) Biozonation and Paleobathymetry on Foraminifera Upper Cretaceous Deposites of Central Iran Basins (Isfahan, Baharestan Section), *Open Journal of Geology*, 4 (8): :343-353.
- Grantham PJ, Posthuma J, Baak A (1983) Triterpanes in a number of Far-Eastern crude oils. In: *Advances in Organic Geochemistry 1981* (Bjoroy M, Albrecht C, Cornford C, et al. eds.), John Wiley & Sons, New York 675–83.
- Hedberg HD (1968) Significance of high-wax oils with respect to genesis of petroleum, *American Association of Petroleum Geologists Bulletin* 52:736-750.
- Huang WY, Meinshein WG (1979) Sterols as ecological indicators, *Geochimica et Cosmochimica Acta* 43:739–45.
- Hunt J (1996) *Petroleum geochemistry and geology* 2nd ed., Freeman and Company, New York 743.
- Jehangir Khan M, Ghazi S, Mehmood M, Yazdi A, Naseem AA, Serwar U, Zaheer A, Ullah H (2021) Sedimentological and provenance analysis of the Cretaceous Moro formation Rakhi Gorge, Eastern Sulaiman Range, Pakistan, *Iranian Journal of Earth Sciences* 13 (4), 252-266.
- Knox G.J, Omatsol EM, (1989) Development of the Cenozoic Niger Delta in terms of the “Escalator Regression” model and impact on hydrocarbon distribution, *Proceedings KNGMG Symposium 'Coastal Lowlands, Geology and Geotechnology* 181-202.
- Kulke H (1995) Nigeria. In: Kulke H (Ed.), *Regional Petroleum Geology of the World. Part II: Africa, America, Australia and Antarctica*. Gebrüder Borntraeger, Berlin.
- Moldowan JM, Dahl J, Huizinga BJ (1994) The molecular fossil record of oleanane and its relation to angiosperms, *Science* 265:768–71.
- Moldowan JM, Sundararaman P, Schoell M (1986) Sensitivity of biomarker properties to depositional environment and/or source input in the Lower Toarcian of South West Germany, *Organic Geochemistry* 10:915–26.
- Okosun EA, Osterloff P (2013) Ostracod, Diatom and Radiolarian Biostratigraphy of the Niger Delta, Nigeria, *Earth Science Research* 3(1): 72-93.
- Olade MA, (1975) Evolution of the Nigerian Benue Trough (Aulocogen): A Tectonic Model, *Geologic Magazine* 12:575-583.
- Onojake MC, Abrakasa S (2021) The Occurrence and Distribution of Oleanane Biomarkers in crude oils as an Index, *Journal of Petroleum Science and Technology* 11(1): 43-50.
- Onojake MC, Osuji LC, Oforka NC (2013) Preliminary hydrocarbon analysis of crude oils from Umutu/Bomu fields, south west Niger Delta Nigeria, *Egyptian Journal of Petroleum* 22:217–224.
- Onuoha KM, (1981) Sediment loading and subsidence in the Niger Delta sedimentary basin, *Journal of Mining Geology* 18 (1):138–140.
- Onyia V, Adejobi A, Ibie E, Nkeme U, Haack R (2002) Regional chronostratigraphic interpretation in northwestern Niger Delta, *NAPE Bulletin* 16:81–92.
- Ouirsson G, Albrecht P, Rohmer M (1984) The microbial origin of fossil fuels. *Scientific American* 251:44–51.
- Peters KE, Fraser TH, Amris W, Rustanto B, Hermanto E (1999) Geochemistry of crude oils from eastern Indonesia. *American Association of Petroleum Geologists Bulletin* 83:927-1942.
- Peters KE, Moldowan JM (1993) *The Biomarker Guide: Interpreting Molecular Fossils in Petroleum and Ancient Sediments*. Prentice Hall, Englewood Cliffs, New Jersey.
- Peters KE, Walters CC, Moldowan JM (2005) *The Biomarker Guide: Biomarkers and isotopes in petroleum systems and Earth history* (Vol. 2), 2nd edition, Cambridge university press, New York.
- Powell TG, McKirdy DM (1973) Relationship between ratio of pristane to phytane, crude oil composition and geological environment in Australia, *Nature* 243:37–9
- Reijers TJA (2011) Stratigraphy and sedimentology of the Niger Delta, *Geologos* 17(3): 133–162 .
- Reijers TJA, Petters SW, Nwajide CS (1997) The Niger Delta basin, in Selley RC ed., *African basins: Amsterdam, Elsevier Science, Sedimentary Basins of the World* 151– 172.
- Salari, M, Yazdi A (2017) Engineering Classification of Genou Limestone Mass, *The Electronic Journal of Geotechnical Engineering*. Oklahoma State University. 22(14): 5553-5560.
- Sharifi Teshnizi E, Yazdi A, Rahnamarad J (2021) Geotechnical Characteristics of Liquefaction in Shahid-Rajaei Port Site (Bandar Abbas, Hormozgan Province) by Using GIS, *Geotechnical Geology* 17 (2), 613-626.
- Short KC, Stauble AJ (1965) Outline of Geology of Niger Delta, *American Association of Petroleum Geologists Bulletin* 51(5):761-779.
- Sonibare O, Alimi H, Jarvie D, Ehinola OA (2008) Origin and occurrence of crude oil in the Niger delta, Nigeria, *Journal of Petroleum Science and Engineering* 61:99–107.
- Ten Haven HL, de Leeuw JW, Rullkotter J, Sinninghe DJS (1987) Restricted utility of the pristane/phytane

- ratio as a palaeoenvironmental indicator, *Nature* 330:641–3.
- Ten Haven HL, Rullkotter J (1988) The diagenetic fate of taraxer-14-ene and oleanene isomers, *Geochimica et Cosmochimica Acta* 52:2543–8.
- Tissot P, Welte D (1984) Petroleum formation and occurrence, 2nd ed., Springer Verlag, Berlin, 699.
- Turcotte DL (1977) On the thermal and subsidence history of sedimentary basins, *Journal Geophysics Research* 82:3762–3766.
- Udo OT, Ekweozor CM (1990) Significance of oleanane occurrence in shales of the Opuama Channel Complex, Niger Delta, *Energy & Fuels* 4:248–54.
- Ukpong AJ, Anyanwu TC (2018a) Late Eocene—Early Oligocene Foraminiferal Biostratigraphy and Palaeoenvironment of Sediments from “Beta- 24 Well” Niger Delta Basin, South Eastern Nigeria, *European Academic Research* 6(2):871-891.
- Ukpong AJ, Anyanwu TC (2018b) Foraminiferal Distribution, Stratigraphy & Palaeoenvironment of the U-12 Well, Niger Delta Basin, Nigeria, *International Journal for Scientific Research and Development* 6(5):1043- 1050.
- Ukpong AJ, Ikediasor KC, Anyanwu TC, Osung EW, Ekhaliu OM (2017) Foraminiferal Biozonation of —Well K-271, Greater Ughelli Depobelt, Niger Delta Basin, South Eastern Nigeria, *EPRA International Journal of Multidisciplinary Research* 3(10):23-32.
- Van Graas G (1990) Biomarker maturity parameters for high maturities: calibration of the working range up to the oil/condensate threshold, *Organic Geochemistry* 16: 1025-1032.
- Wang Z, Stout S, Fingas M (2006) Forensic Fingerprinting of Biomarkers for Oil Spill Characterization and Source Identification, *Environmental Forensics* 7:105–146.
- Waples DW (1983) A reappraisal of anoxia and organic richness, with emphasis on Cretaceous of North Atlantic, *American Association of Petroleum Geologists Bulletin* 67:963–78.
- Whitehead EV (1974) The structure of petroleum pentacyclanes. In: *Advances in Organic Geochemistry 1973* (Tissot B, Bienner F eds.), Editions Technip, Paris, 225–43.