Studies on Physicochemical and Structural Properties of Marshmallow (*Althaea officinalis*) Seed Mucilage

S. Moazzezi^a, A. H. Elhamirad^{b*}, L. Nateghi^c, M. H. Haddad Khodaparast^d, F. Zarei^e

^{*a*} MSc of the Department of Food Science and Technology, Sabzevar Branch, Islamic Azad University, Sabzevar, Iran.

^b Associate Professor of the Department of Food Science and Technology, Sabzevar Branch, Islamic Azad University, Sabzevar, Iran.

^c Associate Professor of the Department of Food Science and Technology, Varamin Branch, Islamic Azad University, Varamin, Iran.

^d Professor of the Department of Food Science and Technology, Ferdowsi University of Mashhad, Mashhad,

Iran.

^e PhD in Food Science, Food and Drug Administration, Tehran, Iran.

Received: 12 May 2021

Accepted: 21 June 2021

ABSTRACT: Marshmallow (*Althaea officinalis*) belonging to *Malvaceae* family possesses mucilage containing cells in stem, petiole, petals and seeds showing antimicrobial activity, anti-inflammatory, immunomodulatory effects among others. In this study, in order to determine the characteristics of marshmallow seed mucilage, as a potential new source of hydrocolloid, some instrument methods (Scanning Electron Microscopy, Fourier transform infrared spectroscopy, nuclear magnetic resonance spectroscopy, etc.) were used and viscosity of mucilage was determined. SEM analysis showed that the mucilage had an amorphous structure and disordered particle size. Mucilage solution at pH of 7 had negative charge; zeta potential of -22.4 mV, electrical conductivity of -1.753 mS /cm and particle size being 255.1 d.nm. Its glass transition temperature (Tg) was 37.9 °C and the melting process started at 34.3 °C to 182.2 °C. An endothermal peak was observed at 92.6°C. Heat of infusion was 199.19 J/g. The most important functional groups identified by FTIR were an asymmetric stretching of double-bond (C=0) in the deprotonated carboxylated groups at 14.3 and 1627 cm⁻¹ vibratory stretching ring of pyranose at 1280 cm⁻¹ and 1115 cm⁻¹ as well as glycoside bonds at 617 and 780 cm⁻¹. Marshmallow seed mucilage solution showed a shear-thinning (pseudoplastic) behavior and the most predominant elements found in the mucilage were carbon (26.59%), potassium (26.39%).

Keywords: DSC, Marshmallow, NMR, SEM, Seed Mucilage, Viscosity.

Introduction

In recent years' interest in use of natural polymers as gelling, thickening, stabilizing and emulsifier agents has increased (Zatz *et al.*, 1989). In addition, hydrocolloids have many secondary functional properties, including body forming, film forming,

purifying, clouding and foam stabilizing agents, crystallization inhibition, etc. (Glicksman, 1982). One of active compounds in plants is mucilage which is a high molecular weight biopolymer produced naturally during normal plant growth. The most common source of mucilage is seeds. However, it is also found in skin, leaves, flower, root, bulb, tubercles and fruits

^{*}Corresponding Author: ah_elhami@iaus.ac.ir

(Niknam, 1999). Hydrolysis of mucilage apart from uronic acid reveals pentose and hexose with the most common ones being arabinose, xylose, rhamnose, mannose, galactose and glucose (Moafeghy, 1992). Marshmallow (Althaea officinalis) belonging to Malvaceae family possesses mucilage containing cells in stem, petiole, petals and seeds showing antimicrobial activity, anti-inflammatory, immunomodulatory effects among others (Hashemluyan et al., 2010; Al-Snafi, 2013). Marshmallow extract has intense antioxidant activity as all studies demonstrated its antioxidant properties (Elmastas et al., 2004). Polysaccharides found in this plant possesses antioxidant property whith 69% antioxidant activity being resulted from its α-Tocopheral (Kardosava and Machova, 2006). As natural gums and mucilage these polymers are inexpensive available and bio compatible and they are being much attention as compared to synthetic and semisynthetic compounds because they have lower toxicity more reasonable cost and a non-allergenic nature (Kulkarni et al., 2005). Hashemluyan carried out a study on the extraction and determination of terpene, phenols, and mucilage content in stem, leaves, plants and seeds of marshmallow (Althaea officinalis) and compared their mucilage percentage. The result showed that these organs contain high mucilage content; however, its seeds contain the highest mucilage percentage (Hashemluyan, 2010). In 2010, Rani and co-workers investigated the phytoconstituents of marshmallow seeds (Althea officinalis) and introduced three phytoconstituents, n-hexacos-2-envl-1, 5olide (altheahexacosanyl lactone), 2 β hydroxycalamene (altheacalamene) and 5,6dihydroxycoumarin-5-dodecanoate-6 β-Dglucopyranoside (altheacoumarin glucoside) with along known phytoconstituents such as lauric acid, βwhich sitosterol and lanosterol their structures were revealed by optical analysis

and chemical reactions (Rani et al., 2010). In 1983, Capek et al. extracted polysaccharides from marshmallow (Althea officinalis) root and examined their arabinans structure. According to their results, the water-soluble arabinans had a branched structure with some α - L- arabinofaranosyl branches that are linked by different bonds $(1 \rightarrow 5)$, $(1 \rightarrow 2)$ and $(1 \rightarrow 3)$ and some of the L-arabinosyl groups are involved in branches through O-2, O-3, and O-5. There was a good agreement between the results obtained by chemical methods and 13Cn-m-r. The derived polymer had structural properties similar to those of L-arabinan isolated from other plant sources (Capek et al., 1983). The aim of this study was to investigate the physicochemical and structural characteristics marshmallow of seed mucilage.

Materials and Methods

Marshmallow seeds were purchased from (Pakan-bazr Co, Isfahan, Iran). In order to extract Marshmallow seed mucilage a slightly modified method of Farahnaky et al., (2013) was used. Marshmallow seed were ground and hydrated (seed powder: water, 1:50 w/v at room temperature for all night. The mixture was centrifuged with (Rotofix 32 A, Germany) at 4000 rpm for 20 min at 25 °C to separate the mucilage. The gum solution, was then separated, dried at 50 °C, ground and then packed in hermetic sealed plastic bags and kept in cool and dry place for later experiments. Determination of zeta potential, conductivity and particle size by using Malvern Zetasizer Nano series. Nano _ Zs (Malvern Instrument Worcestershire, UK), zeta potential of 100 (g/L) solutions was measured through the disposable folded capillary cell (DTS 1060) at 25 °C. The pH value of solutions ranged from 6.29-6.69. The conductivity and particle size of samples were also measured. A differential scanning calorimeter (INNUO DSC-500B, China) was used for DSC

analysis of mallow seed mucilage. 5 mg sample was placed into a platinum cup and sealed. The temperature ranged from 20-300 °C under nitrogen atmosphere with a heating rate being 10°C /min (Stopic et al., 1997). FTIR spectra observed for the mucilage were recorded on a FTIR spectrometer (perkin elmer spectrum 400, USA). The dry powder was mixed with KBr and pressed to form pellets under mechanical pressure. The FT-IR spectra were obtained at 4000 and 400/cm (Sahoo et al., 2010). Scanning Electron Microscope (SEM) (TESCAN Vega-3 SBU model, Czech Republic) with acceleration voltage of 5.0 kV was used. The samples were mounted on an aluminum stub with double-sided adhesive tape. The tape was attached to the stub and the sample powder was scattered carefully over its surface. The stub with the sample was then coated with a thin layer of gold to make the sample conductive. The specimen was subjected to SEM analysis (Nep and Conway, 2010). Quantitative elemental analysis and distribution of the elements (by X-ray mapping) in the Si/C composite was examined using the energy dispersive X-ray Spectroscopy (EDS) analyzer attached with NMR spectra of 1H and 13C of the SEM. mucilage were recorded in an NMR (250MHz) spectrometer (Bruker Avance model, Germany). The test mucoadhesive agent (100mg) was dissolved in D2O and chemical variations were reported in ppm against an internal standard TSP (3trimethylsilylpropionic-2, 2, 3, 3, -d4 acid, sodium salt, 98% D) for 1H NMR and 1,4dioxane (d 66.67 ppm) for 13C spectra. Proton NMR spectra were obtained at a base frequency of 250MHz, with 16 transitions and delay time 1.5 s and for 13C, the base frequency was 100MHz, with 3000 scans and delay time 2 s (Hamcerencu et al., 2008). The viscosity of 40 (g/L) solutions was measured by use of a rheometer (Anton paar, MCR300, CC27, Austria). The effect of shear rate on the apparent viscosity of hydrocolloid solution at 1-1000 S-1 was investigated. The data obtained from the measurements were subjected to univariate variance analysis one-way analysis (ANOVA) to determine significant differences among the samples and values were compared by use of Tukey's test defined at $P \le 0.01$. All measurements were carried out in triplicate order and reported as mean \pm SD from independent trials. Data analysis was performed by SPSS software (version 16; IBM Corporation, Armonk, NY, USA).

Results and Discussion

Zeta potential indicates the surface and sub - surface charge of suspended particles. It is an indicator of colloid solutions stability. If the particles contained in suspensions have very high or very low zeta potential. They will show the tendency towards repulsion and aggregation was reduced (Acedo-Carrillo et al., 2006). As a general rule, the stability or un stability of a suspension may be determined by zeta potential. Particles are stable when their zeta potential is > 30 mV and / or < -30 mV. Although some exceptions have been reported (Wang et al., 2010). Zeta potential is an indicator for measurement of pure of emulsion particles often charge representing the charged part of the biopolymer (Jindal et al., 2013). Various factors including pH, ion strength, type and concentration of protein and polysaccharide macromolecules affect surface charge, electrophoretic movement and complex surface charge, electrophoresis movement and complex zeta potential (Khoshmanzar et shown in Figure al.. 2013). As 1 marshmallow mucilage solution (1mg/ml) showed negative charge at pH 7 and a zeta potential of -22.4 mV showing low stability due to its low zeta potential.

Particle size and particle size distribution of hydrocolloids are important parameters for the solubility and strength of an emulsion (Mathur, 2012). Particle size (as average diameter, Z- average) in 1 mg/ml marshmallow seed mucilage solution passed 0.45 micron filter is shown in Figure 2 Particle Zaverage of marshmallow mucilage aqueous dispersion was 255.1 d.nm, while its PDI (polydispersity index) was 0.66. The results reveal micro particles polydispersity. with high Researchers reported that particle size in gums affects their swelling in water due to the surface changes exposed to water, in turn, influence, the inherent viscosity as well as molecular mass of gums. Also particle size also affects the swelling rate and molecular weight of guar gum (Wang et al., 2003).

Electrokinetic measurement is one way to study complex surface chemical exchangeprocesses at the mineral surface/liquid interface. All electro kinetic phenomena are related to the development of electrical double-layer particles/electrolyte at interface. Additionally, electrophoretic movement of suspended particles depends on zeta potential. It has been concluded that conductivity in any solution is a function of the concentrations of electrolytes, ions as well as electrical charges leading to an increase in electrical conductivity. Electrical conductivity and dielectric constant increase with heat and electrical stress with these changes being due to the decomposition of compounds and subsequent formation of compounds having smaller molecules. Given the previous studies it could certainly said that ZP has a great effect on mucilage properties (Singh and Bothara, 2014). marshmallow seed mucilage solution (1



Fig. 1. Zeta potential data of marshmallow seed mucilage.



Fig. 2. Particle size data of marshmallow seed mucilage.

mg/ml) has an electrical conductivity value of -1.753 mS/cm.

DSC is a useful method for investigation of physical and chemical changes occurring during heating process in the polymers. This technique is based on the thermal changes. When a sample is subjected to thermal changes (received or lost thermal energy), it experiences a physical change such as phase transition. The amount of heat entered (or exit) the sample is different from the heat entered (or exit) a standard (Skoog et al., Changes including 2011). melting, crystallization, freezing and glass transition all being the indicators of compounds identification may be evaluated by this method (Haines et al., 1998). Glass transition temperature (Tg) the is temperature at which density, hardness and stiffness of polymer increase and its elongation percentage decreases drastically, i.e. it is the intermediate temperature between melted and hard states of matters. Above the Tg, secondary bonds between molecules are considerably weaker than thermal movement, since the polymer becomes rubbery and acquires a certain elasticity and plastic deformation without fracture occurs (Mathot et al., 1994; Maglic et al., 1984; Höne et al., 1996). Thermogram of marshmallow seed mucilage is given in Figure. 3. Suggesting that the melting process started at 34.3 °C to 182.2 °C. An

endothermal peak indicating the melting process was observed at 92.6 °C suggesting its amorphous structure with the heat of fusion was found to be 199.19 J/g.

FTIR is widely used for describing the molecular polymer and investigating the type and number of functional groups in the structure. FTIR was used for understanding functional groups present in the the marshmallow seed mucilage (Baxter et al., 1992). Characterization using FTIR is often due to the identification of functional group and how they are connected to the structure of polymer. As shown in Figure. 4, The peak observed at 3430 cm-1 may be attributed to the symmetrical vibrations of carbon hydrogen bond (-C-H) in the aliphatic chain of methylene group (-CH2) as well as acid carboxylic functional group (-COOH). The spectra observed at 1627d cm-1 are related to symmetrical vibration of carboxyl doublebond (C=O) in deprotonated carboxylate groups. The mucilage spectrum reveals that the bands found at 1000-1300 cm-1 (1115 and 1280 cm-1) are related to the vibrations of oxygen – carbon (-C-O), while the band observed at 617 and 780 cm-1 demonstrates the simple bonds of carbon – carbon (C-C) and out - plane vibrations of hydrogen carbon (=C-H). FTIR revealed the presence of carboxyl groups likely used as the position where bivalent cautions are connected (Bramhachari et al., 2007)



Fig. 3. Differential Scanning Calorimetry (DSC) Characterization of marshmallow seed mucilage Using DSC analyzer.

S. Moazzezi et al.



Fig. 4. FTIR spectral characterization of marshmallow seed mucilage using thermo scientific FTIR spectrophotometer.

SEM is an appropriate method for physical observations and morphological evaluations (Qi et al., 2005). In the present study, the micrograph of different levels of marshmallow seed mucilage suggests its surface amorphous nature with its characteristics may be attributed to the hydration capacity of mucilage. SEM analysis also indicates that the mucilage had different particle size and disordered morphology suggesting its fiber nature (Figure 5a and b), although the surface structure and/or topography of mucilage might be affected by the method of extraction or purification (Qian et al., 2009). It has been stated that particle size and specific surface area of gum and mucilage affect their hydration behavior in turn inherent influence the viscosity and molecular mass. Wang empirically reported that particle size influenced the rate of water absorption as well as molecular mass of guar some gum rich in galactomannan (Wang et al., 2002). A very rough surface with large wrinkle was observed likely caused by partly collapsed polymer gel network when drying (Figure 5c), In some part a continuous system but with a disordered and uneven structure having less roughness, rounded edges and different particle size was observed (Figure 5d), The analysis of elements may determine some chemical elements as minerals contained in the sample such as carbon, hydrogen, nitrogen and sulphur quantitatively and qualitatively. The results obtained by EDS showed the contents of elements as follows: C, N, O, Mg, S, Cl, K, and Ca at 26.59%, 5.34%, 18.32%, 2.04%, 4.22%, 9.40%, 26.39% and 7.69% respectively.

Apparent viscosity of 40 (g/L) marshmallow seed mucilage solution is given in figure. 6 apparent viscosity decreased from 0.296 to 0.0147 Pa as hear rate increased from 0.1 to 1000 s-1 showing dependency on the shear rate suggesting that the mucilage is pseudoplastic.

¹H and ¹³C NMR spectral characterization of marshmallow seed mucilage are shown in Figure 7 and Figure 8 respectively. According to Table 1, all compounds in certain amount are present in the mucilage extracted from marshmallow seed mucilage, which is a non–starch polysaccharide. The higher integral value or the greater peak area the higher the amount of that type of proton in the compound. Among the suggested compounds, rhamnose was more prominent. The peak area for protons bound to second carbon type, (-C-H) in aliphatic chain is greater because it has a greater integral area. On the other hand, first carbon type, methyl

J. FBT, IAU, Vol. 12, No. 1, 29-38, 2022

carbon (-CH3) is present only in the structure of rhamnose being visible in that spectrum. In addition to (-C-H) peaks in H-NMR spectra, the peaks observed for the proton of hydroxyl group (-O-H) and carboxyl (-COOH), having a great area, suggest the presence of all above compounds. Given the presented structures, the most predominant proton groups were methyl first carbon type – bound proton, second carbon type – bound proton, and oxygen – bound proton known as hydroxylic proton (-O-H) which is an alcoholic, hydroxylic or carboxylic type.



Fig. 5. Scanning electron microscopy of marshmallow seed mucilage at different magnification using (TESCAN Vega-3 SBU model, Czech Republic).



Shear rate (1/s)

Fig. 6. The apparent viscosity of 40 g/L marshmallow seed mucilage solution at shear rate of 0.1-1000 1/s.

Table 1. Position, Area and kind of peaks in 1H-NMR spectra

Peak height (Integral)	2.019	0.936		1.000	2.815		1.452	1.438		
Chemical shift (ppm)	1.034^{a}	1.062^{b}	1.636 ^c	2.134 ^d	2.918 ^e	2.983^{f}	3.626 ^g	3.827 ^h	3.855 ⁱ	4.604 ^j
Glucuronic acid (Gu)	-	-	-	-	+	+	+	+	+	+
Galactronic acid (Ga)	-	-	-	-	+	+	+	+	+	+
Galactose (G)	-	-	-	-	+	+	+	+	+	+
Rhamnose (Ra)	+	+	+	-	+	+	+	-	+	+
Arabinogalactan (Ar)	-	-	-	-	+	+	+	+	+	+

^{a,b,c}: Hydrogen of methyl (-CH₃) group in structures

e.f.g.h: Hydrogen bonded carbon ((II) type) (-C-H-) in structures

^{i,j}: Hydrogen bonded oxygen in hydroxyl group (-O-H-) in structures

According to Table 1, 13C-NMR peaks observed for Suggested compounds were revealed and C-13 indicates the all compounds given the observed peaks. The peaks appeared at 26-31 ppm indicate first carbon type, methyl carbon (-CH3), only being found in rhamnose structure. The peaks observed at 63 - 100 ppm related to CII where hydrodxy groups are connected to it at different positions being referred to as alcoholic carbon (-C-OH) or acid carbon (-COOH). This type of carbon is found in all compounds are shown in Table 2, appeared in the respective spectrum. Only the peak for aldehyde carbon (-C=O), commonly found at 200 ppm, did not appeared likely due to peaks overlapping and/ or the impurities in the matter.

Conclusion

The results indicated that the mucilage

has an amorphous structure with different particle size showing a non-Newtonian shear-thinning (pseudoplastic) behavior. Marshmallow seed mucilage has microprticles (255.1 d.nm) with high polydispersity (PDI =0.66). It has low stability because of having low zeta potential (-22.4mV). However, it may be used for stabilization of negatively charged colloid systems or sedimentation of positively charged system due to possessing of negative charges. Its Tg was 37.9 °C being attributed to the amorphous structure of the mucilage and the heat of infusion was 199.19 J/g. The most important functional groups identified by FTIR were an asymmetric stretching of double-bond (C=0) in the deprotonated carboxylated groups at 14.3 and 1627 cm⁻¹ vibratory stretching ring of pyranose at 1280 cm⁻¹ and 1115 cm⁻¹ as well as glycoside bonds at 617 and 780 cm^{-1} .



Fig. 7. 1D ¹H NMR Spectral characterization of marshmallow seed mucilage using bruker advance II 250 NMR spectrophotometer.

Chemical shift (ppm)										
Glucuronic acid (Gu)	-	-	-	-	-	+	+	+		
Galactronic acid (Ga)	-	-	-	-	+	+	-	+		
Galactose (G)	-	-	-	-	+	+	+	+		
Rhamnose (Ra)	+	+	-	-	+	+	+	+		
Arabinogalactan (Ar)	-	-	-	-	+	+	-	-		

Table 2. Position and Area of peaks in C-NMR spectra

^{a,b,c}: Carbon of methyl (-CH₃) and reveal (-C) of methyl in structure

e,f,g,h: Carbon ((II) type) bonded hydroxyl group and/or carboxylic acid (-COOH) in aliphatic chain in structure



Fig. 8. 1D ¹³C NMR Spectral characterization of marshmallow seed mucilage using bruker advance II 250 NMR spectrophotometer.

Acknowledgements

This paper was supported by the Department of Food Science and Technology of Islamic Azad University, Sabzevar, Iran.

References

Acedo-Carrilloa, J. I., Rosas-Durazoa, A., Herrera-Urbinac, R., Rinaudod, M., Goycooleab, F. M. & Valdeza, M. A. (2006). Zeta potential and drop growth of oil in water emulsions stabilized with mesquite gum. Carbohydrate Polymers, 65, 327–336.

Baxter, A., Dillon, M., Taylor, K. D. A. & Roberts, G.A.F. (1992). Improved method for it determination of the degree of N-acetylation of chitosan. International Journal of Biological Macromolecules. 14,166–169.

Bramhachari, P. V., Kishor, P. B. K., Devi, R. R., Kumar, R., Rao, B. R. & Dubey, S.K. (2007). Isolation and characterization of mucous exopolysaccharide (EPS) produced by Vibrio furnissii strain VB0S3. *Journal of Microbiology and Biotechnology*. 17, 44–51.

Capek, P., Oman, R., Kardosov, A. & Rosik, J. (1983). Polysaccharides from the roots of the marshmallow (*Althea offcinalis* L.): structure of an Arabian. Carbohydrate Research. 117, 133-140.

Chen, H.H., Xu, S.Y. & Wang, Z. (2006). Gelation properties of flaxseed gum. Journal of Food Engineering. 77, 295–303.

Elmastas, M., Ozturk, L., Gokce, I., Erenler, R. & Aboul- Enein, H.Y. (2004). Detremination of antioxidant activity of Marshmallow flower (*Althaea officinalis*). *Analaticals Letters*, 37, 1859-1869.

Farahnaky, A., Bakhshizadeh-Shirazi, S. H., Mesbahi, G. H., Majzoobi, M., Rezvani, E. & Schleining, G. (2013). Ultrasound-assisted isolation of mucilaginous hydrocolloids from Salvia macrosiphon seeds and studying their functional properties. Innovative Food Science & Emerging Technologies. 20, 182-190.

Glicksman, M. (1982). Food Hydrocolloids, Vol 1, 2 and 3, FL: CRC Press Inc.

Höne, G. W. H., Hemminger, W. & Flammersheim, H. F. (1996). Differential scanning calorimetry: An introduction for practitioners. (2nd. Ed.). New York.

Hamcerencu, M., Desbrieres, J., Khoukh, A., Popa, M. & Riess, G. (2008). Synthesis and characterization of new unsaturated esters of Gellan Gum. *Carbohydrate Polymers*. 71, 92-100.

Haines, P. J., Reading, M. & Wilburn, F. W. (1998). Differential thermal analysis and differential scanning calorimetry. In Brown ME (ed): Handbook of Thermal Analysis and Calorimetry, vol 1. The Netherlands: Elsevier Science BV. 279–361

Iwe, M. O., Obaje, P. O. & Akpapunam, M. A. (2004). Physicochemical properties of Cissus gum powder extracted with the aid of

edible starches. *Plant* Foods for Human Nutrition. 59, 161–168.

Jindal, M., Kumar, V., Rana, V. & Tiwary, A.K. (2013). Physico–chemical, mechanical and electrical performance of bael fruit gumechitosan IPN films. *Food Hydrocolloids*. 30,192-199.

Kulkarni, G. T., Gowthamarajan, K., Dhobe, R. R., Yohanan, F. & Suresh, B. (2005). Development of controlled release spheriods using natural polysaccharide as release modifier. *Drug Delivery*. 12, 201-206

Kardosava, A. & Machova, E. (2006). Antioxidant activity of medicinal plant polysaccharides. *Fitoterapia*. 77367-73.

Mathot V. B. F. (1994). Calorimetry and thermal analysis of polymers. *Journal of Thermal Analysis*. 45, 577–578.

Maglic, K. D., Cezairliyau, V. E. & Peletsky, P. (1984). Compendium of thermophysical property measurment methods. (Vol. 1). New York: Plumen Press.

Mathur, P., Saroha, K., Syan, N., Verma, S., Nanda, S. & Valecha, V. (2011). An overview on recent advancements and developments in gastroretentive buoyant drug delivery system. Pelagia *Research Library*. 2: 161-169.

Moafeghy, A. (1992). Isolation and determination of mucilage polysaccharides from plantagoes with tissue and farming cultivated. MSc. Thesis, School of science, Tehran University.

Mhaskar, K. S., Blatter, E. & Caius, J. F. (2000). Kirtikar and Basu's illustrated Indian medicinal plants. 2,405–407. Delhi: Sri Satguru Publication.

Niknam, V. (1999). Identification of secondary metabolits (N- aliphatic composition, mucilage polyssacharides, saponines, sterols, phenolic compositions) PH.D. Thesis, School of science, Tehran University.

Nep, E. I. & Conway B. R. (2010). Characterization of Grewia Gum, a potential pharmaceutical excipient. *Journal of Excipients and Food Chemicals.* 1, 30–40.

Qi, W. & Cui, S. W. (2005). Understanding the physical properties of food polysaccharides. In: Cui SW, editor. Food Carbohydrates: Chemistry, Physical Properties, and Applications. 3 rd ed. Boca Raton, Florida: Taylor and Francis; 161-262.

Qian, J., Chen, W., Zhang, W. & Zhang, H. (2009). Adulteration identification of some fungal polysaccharides with SEM, XRD, IR and optical rotation: a primary approach. *Carbohydrate Polymers*. 78, 620-625.

Singh, S. & Bothara, B. S. (2014). Physicochemical and structural characterization of mucilage isolated from seeds of Diospyros melonoxylon Roxb. *Brazilian Journal of Pharmaceutical Sciences*. 50, 713-720.

Stopic, S. R., Ilic, I. E. & Uskokovic, O. P. (1997). Effect of Pd, Cu, and Ni additions on the kinetics of NiCl2 reduction by hydrogen. *Metallurgical and Materials Transactions*. 28, 1241–1248.

Sahoo, N., Manchikanti, P. & Dey, S. (2010). Herbal drugs: standards and regulation. *Fitoterapia*. 81, 462–471.

Skoog, D. A., Holler, F. J., Crouch, S. R. (2011). Instrumental Analysis. India edition: Cengage Learning. 982-984

Wang, Q. I., Ellis, P. R. & Ross-Murphy, S. B. (2002). Dissolution kinetics of guar gum powders - 1. Methods for commercial polydisperse samples. *Carbohydrate Polymers*. 49,131-137.

Wang, J., Feng, S., Wang, S. & Chen, Z. (2010). Evaluation of cationic nanoparticles of biodegradable copolymers as siRNA delivery system for hepatitis B treatment. *International* Journal of Pharmaceutics. 400, 194–200.

Wang, Q., Ellis, P. R. & Ross-Murphy, S.B. (2003). Dissolution kinetics of guar gum powders—II. Effects of concentration and molecular weight. *Carbohydrate Polymers*. 53, 75–83.

Wang, Q. I., Ellis, P. R. & Ross-Murphy, S. B. (2006). Dissolution kinetics of guar gum powders - 3. Effect of particle size. *Carbohydrate Polymers*. 64, 239-246,

Zatz, J. L. & Kushla, G. P. (1989). In: pharmaceutical dosage Forms-Disperse systems, M. M. Reiger and G.S. Banker, Ed; Marcel Dekker Inc., New York, 2, 508.