

Journal of Ornamental Plants Available online on: www.jornamental.iaurasht.ac.ir ISSN (Print): 2251-6433 ISSN (Online): 2251-6441

Genetic Variability and Karyotype Analysis for 13 Accessions of *Lolium multiflorum*

Soosan Abbaszade 1*, Ali Ashraf Jafari 2, Hooshmand Safari 3 and Hooman Shirvani 4

¹ Young Researchers and Elite Club, Kermanshah Branch, Islamic Azad University, Kermanshah, Iran

² Research Institute of Forests and Rangelands, Tehran, Iran

³ Agriculture and Natural Resources Research Center, Kermanshah, Iran

⁴ Department of Agriculture, Payame Noor University, Iran

Received: 26 September 2016 Accepted: 14 October 2016 *Corresponding author's email: so.abbaszade@yahoo.com

Genetic variations were studied for 13 accessions of Lolium multiflorum using cytogenetical traits. Karyotype was prepared for 5 metaphases cells of each accession and the traits of total length (TL), long arm (LA), short arm (SA), arm ratio (AR) and centromer index (CI) were determined by micromeasure software. Six accessions were diploid and seven accessions were tetraploid, the basic chromosome number was x=7, and also three and six satellites were observed for diploid and tetraploid accessions, respectively. The accessions had an asymmetry karyotype, and variations were significant (at 1% level of probability) between accessions for karyotypic characters based on analysis of variance (ANOVA). Results of mean comparison showed that the diploid accessions had higher mean chromosome length than tetraploid accessions. The results of cluster analysis by Ward's method based on the values of intrachromosomal asymmetry index (A1) and interchromosomal asymmetry index (A2) showed that the studied accessions were fallen in three groups. The tetraploid accessions had higher intrachromosomal asymmetry than the diploid accessions. Principal components analysis based on karyotypic traits for accessions showed the first two components captured 84.37% of the total variance. Principal component analysis for grouping accessions based on Scatter plot identified four distinctive groups.

Keywords: Cytogenetic, Genetic variation, Karyotype, Lolium multiflorum.

Abstrac

INTRODUCTION

The genus Lolium L., family Poaceae (Geraminea), subfamily festucoideae, belongs to the tribe festuceae Nees (Zwierzykowski and Naganowski, 1996). This genus is one of the most important categories of temperate forage grasses that are diploid (2n = 2x = 14), although some accessions have been reported as tetraploid (2n = 4x = 28) (Mirjalili, 2014). The Lolium genus contains only eight species, including the agronomically important taxa annual or Italian ryegrass (Lolium multiflorum Lam.) and perennial ryegrass (Lolium perenne L.) (Lundqvist, 1962; Cornish et al., 1979), which are used for forage and turf purposes throughout the temperate regions of the world including North and South America, South Africa, Australia, and New Zealand (Lamp et al., 2001).

Breeding programs are dependent on genetic variation for the development of improved cultivars. Therefore, the knowledge of genetic diversity is pertinent to improving overall plant characteristics which will allow for a systematic sampling of germplasm for breeding and conservation purposes (Che and Li, 2007). A rich and diverse germplasm collection is the backbone of every successful crop improvement programs. The genetic variability is the raw material to crop breeding industry in which selection acts to evolve superior genotypes. Morphological characteristics are the strongest determinants of the agronomic value and taxonomic classification of plants. Compared with other methods morphological evaluations are direct, inexpensive, and easy. However, errors can arise; furthermore, morphological estimations are more dependent on environment (Chowdhury et al., 2002; Iruela et al., 2002; Sudupak et al., 2002).

Interpretation of processes resulting in genetic variation and evolution are made possible by chromosome studies (Hajimoniri, 1999). Gupta (1995) defined karyotype and explained that similarities and differences between the plant taxons may arise from phylogenetic relationships. Karyotypic characters (such as chromosome length, arm ratios, and secondary constrictions) can be useful for individual chromosomes identification and phylogenetic studies. Karyological studies are very important, because chromosomes include genes containing information about phenotype of the plants. Mirzaei-Nodushan and Nadarkhani (2001) studied karyotypes of nine populations of L. multiflorum and L. rigidum. They measured karyological traits such as arm length, chromosome number, and symmetry on diploid and tetraploid populations. The aim of this study was to explain the genetic diversity between accessions of Lolium multiflorum based on karyological characters.

MATERIALS AND METHODS

Plant materials

In order to evaluate the genetic variation, 13 accessions of L. multiflorum were prepared from gene bank of Research Institute of Forests and Rangelands, Tehran, Iran (Table 1).

Cytological studies

For karyotype study, freshly grown root tips were collected from the germinated seed, pretreated with α-Bromonaphtaline (4 h), and fixed in Levitsky solution (Solution A: chromium trioxide and solution B: Formaldehyde 37%) for 16 hours. Root tips were then hydrolyzed in 1%

Gene bank code	Origin	Number	Gene bank code	Origin	Number
usa	USA	G7	390	Italy	G1
374	Italy	G8	393	Italy	G2
1766	Netherlands	G9	374"	Italy	G3
1765	Netherlands	G10	1551	Russia	G4
1624	Netherlands	G11	1151	Unknown	G5
vi	Russia	G12	1268	France	G6
plc-Early	Unknown	G13			

Table 1. Gene bank code and origin of accessions of L. multiflorum.

2 Journal of Ornamental Plants, Volume 7, Number 1: 1-8, March, 2017

$$\% TF = \frac{\sum_{i=1}^{n} SA_i}{\sum_{i=1}^{n} TL_i} \times 100$$

$$DRL = \% RL_{Max} - \% RL_{Min}$$

$$VRC = \frac{\sum_{i=1}^{n} TL_i}{n}$$

$$A_1 = 1 - \frac{\sum_{i=1}^{n} \frac{S_i}{L_i}}{n}$$

$$A_2 = \frac{S}{\overline{d}}$$

NaOH at 60°C for 8 minutes; therefore, hematoxiline was used for chromosome staining (4 h). Cytotype cells were recorded on at least five well-prepared cells at metaphase stage for each population by microscope Olympus BH2 monitoring systems (At magnification of 2775). The traits of total length of chromosomes (TL), short arm length (SA), long arm length (LA), arm ratio (AR), and centromer index (CI) were measured by MicroMeasure software. Then the parameter of short arm relative length percentage (S%), long arm relative length percentage (L%), relative length percentage (RL%), total form percentage (TF%), difference of range relative length (DRL), value of relative chromatin (VRC), intrachromosomal asymmetry index (A₁), and interchromosomal asymmetry index (A₂) were calculated. The types of chromosomes were identified according to Levan *et al.* (1964), and karyotype symmetry was determined according to Stebbins (1971).

Statistical analyses

Analysis of variance was performed on the data recorded on the karyotypic traits using SPSS software package. Mean comparison were performed by Duncan's test at 5% levels of probability for means of karyotypic traits for accessions. Principal components analysis was performed on the data using SPSS software.

RESULTS

Karyotype analysis

The results showed that six accessions (374, 1766, vi, 1624, plc and 1765) were diploid, seven accessions (393, 1551, 374", 1151, 390, 1268 and usa) were tetraploid (Fig. 1), and the basic chromosome number was x = 7. Three and six satellites were observed for diploid and tetraploid accessions, respectively (Table 2).

Karyotypic formula was varied for survived accessions, so that it was 5m + 3sm + 6st for accessions 374 and 1766, and 2m + 2sm + 3st for accessions 374" and 1151. Karyotypic formula for 393, 1268 and usa was 3m + 4sm. Also, the vi had 3m + 5sm + 6st chromosomes. Accessions 1624 had 8m + 6st chromosomes, karyotypic formula was 4m+4sm+6st for plc, and finally, it was 5m + 9sm, 3m + 3sm + 1st and 3m + 1sm + 3st for 1765, 1551 and 390, respectively. In terms of the Stebbins' system, the karyotype of accessions seizes 2A classes, which are considered as median asymmetrical karyotypic in this system. Based on the intrachromosomal asymmetry index (A1) and total form percentage (TF%), accessions 374, vi, plc and 390 had the highest value for intrachromosomal asymmetry (A1) and the lowest value for TF%. Therefore, these accessions had the most karyotypic evolution. On the other hand, 1624, 1765, 393 and usa had the lowest karyotypic evolution based on these parameters. Accessions 1765 and 1268 had the lowest interchromosomal asymmetry index (A2) and the lowest karyotypic evolution based on interchromosomal asymmetry index (A2) and the most karyotypic evolution based on interchromosomal asymmetry index (A2) parameters. The diagram of the accessions distribution based on the values of intra-

Gene bank code	2n	DRL	TF%	A1	A2	SC	K.F
374 1766 vi 1624 plc 1765 393 1551 374" 1151 390	2n = 4x = 28 2n = 2x = 14 2n = 2x = 14 2n = 2x = 14 2n = 2x = 14	4.066 4.217 4.566 3.780 4.156 3.898 6.921 6.690 7.344 6.690 7.146	26.637 28.521 27.493 30.896 27.873 30.256 30.115 29.204 28.615 29.204 26.053	0.5237 0.4977 0.5117 0.4270 0.5070 0.4451 0.4453 0.4661 0.4968 0.4661 0.5294	0.1794 0.1737 0.1909 0.1715 0.1740 0.1649 0.1753 0.1745 0.1757 0.1745 0.1789	2A 2A 2A 2A 2A 2A 2A 2A 2A 2A 2A 2A 2A	5m + 3sm + 6st 5m + 3sm + 6st 3m + 5sm + 6st 8m + 6sm 4m + 4sm + 6st 5m + 9sm 3m + 4sm 3m + 3sm + 1st 2m + 2sm + 3st 2m + 2sm + 3st 3m + 1sm + 3st
1268 usa	2n = 2x = 14 2n = 2x = 14	5.699 6.693	29.220 30.032	0.4772 0.4457	0.1550 0.1897	2A 2A	3m + 4sm 3m + 4sm

Table 2. Karyotypic characters for studied accessions.



Fig. 1. Cytotypes of studied accessions.

chromosomal asymmetry index (A_1) and interchromosomal asymmetry index (A_2) and accessions grouped using cluster analysis of Ward's method based on the values of intrachromosomal asymmetry index (A_1) and interchromosomal asymmetry index (A_2) showed that the studied accessions fell into three groups (Fig. 2).

Analysis of variance and means comparison

The results of variance analysis showed a significant variation (at the 1% level of probability) between the accessions for chromosomal traits. So, the tested germplasm varied for karyotypic characteristics (Table 3). Also, the results of mean comparison by Duncan's test (Table 4) revealed that the diploid accessions 393, 1551, 390 and usa and the tetraploid accession plc had the highest length of total chromosome, and long and short arms. The tetraploid accessions 1624



Fig. 2. Scatter plot for accessions based on A1 and A2.

Table 3. Analysis of variance for karyotypic traits of the studied accessions.

S.o.V	df	TL	LA	SA	CI	AR
Accessions Error CV (%) Average	12 52	0.794** 0.154 5.95 6.58	0.183** 0.057 6.59 3.63	0.128** 0.019 7.25 1.890	0.001** 0.0002 4.5 0.30	0.299** 0.036 8.54 2.23

**: significant at the 1% level of probability.

and 1765 and diploid accessions 393, 1268 and usa had the highest centromer index and the lowest intrachromosomal asymmetry. The tetraploid accessions 374 and plc and diploid accession 390 had the highest arm ratio (AR) and intrachromosomal asymmetry index (A₁).

Principal component analysis

usa

The results of principal components analysis based on karyotypic traits for accessions showed that the first two components captured 84.37% of the total variance (Table 5). The most important traits for the first component included the length of short arm (SA), centromer

5% levels.					
Accessions	TL(µ)	LA(µ)	SA(µ)	CI	AR
374	6.31 ^{cd}	3.48 ^{cd}	1.640 ^e	0.274 ^f	2.47 ª
1766	6.03 d	3.39 d	1.720 ^{de}	0.290 cdef	2.40 ab
vi	6.55 bcd	3.68 abcd	1.802 ^{cde}	0.285 def	2.40 ab
1624	6.40 bcd	3.43 d	1.976 abc	0.318 ª	1.88 °
plc	6.84 abcd	3.85 _{ab}	1.907 bcd	0.286 def	2.51 ª
1765	6.52 bcd	3.55 bcd	1.972 abc	0.312 ab	1.97 °
393	7.14ª	3.82 abc	2.149 ª	0.309 abc	1.96 °
1551	7.11 ª	3.80 abc	2.076 ab	0.298 bcde	2.15 bc
374"	6.36 ^{cd}	3.55 bcd	1.819 ^{cde}	0.291 cdef	2.37 ab
1151	6.35 ^{cd}	3.56 bcd	1.825 ^{cde}	0.292 cdef	2.35 ab
390	6.93 abc	3.92 ª	1.805 ^{cde}	0.279 ef	2.56 ª
1268	5.98 d	3.36 d	1.748 de	0.302 abcd	2.05 °

Table 4. The means comparison by Duncan's test for studied accessions at the 5% levels.

*In each column, means with the similar letters were not significantly different at the 5% level of probability using Duncan's test.

2.126 a

3.78 abc

7.08 ab

0.308 abc

1.96 °

Traits	PRIN1	PRIN2
TL	0.495	0.858
LA	0.202	0.945
SA	0.922	0.367
CI	0.917	-0.384
AR	-0.897	0.347
VRC	0.495	0.858
DRL	0.199	0.504
TF%	0.894	-0.398
A1	-0.943	0.293
A2	-0.333	0.554
Eigen value	4.865	3.573
Eigen vector	48.647	35.726
Percentage of cumulative variance	48.647	84.373

Table 5. Eigen value, percentage of variance, cumulative variance for thefirst and second principal components.



Fig. 3. Biplot of karyotypic characters in the studied accessions based on two first components and grouping of accessions.

index (CI), and total form percentage of chromosome (TF%) with the highest positive coefficients of eigen vectors and arm ratio (AR) and intrachromosomal asymmetry index (A₁) with the highest negative coefficients of Eigen vectors. For the second component, the total length (TL) of chromosome, long arm (LA), value of relative chromatin (VRC), difference of range relative length (DRL) and interchromosomal asymmetry index (A₂) played the most important role for total variation. According to the biplot, the accessions were divided into four groups (Fig. 3).

DISCUSSION

No variation in basic chromosome number was observed in the studied accessions, but there were two groups for ploidy level. The diploid accessions' average chromosome length was higher than that of tetraploid accessions. The species *L. multiflorum* has been reported to be diploid in most research (2n = 2x = 14) (Hovin and Hill, 1966; Evans and Mecefield, 1974), but some studies have reported the tetraploid species (Wu and Chen, 1991). On the other hand, the tetraploid accessions had higher intrachromosomal asymmetry than diploid accessions. Essad (1954) based on size and symmetry of chromosomes

suggested the genus *Lolium* to be divided into three classes: 1- *L. temulentum* – *L. remotum*, 2- *L. perenne* – *L. multiflorum*, and 3- *L. rigidum*; the latest resembles the second group more than the first.

The satellites were observed for all accessions, and variations were observed in the number, length and location of satellites on chromosomes. Mirzaei-Nodushan and Nadarkhani (2001) reported the number of one or two satellites for two species of *Lolium*. The accessions varied in karyotypic formula and all accessions were composed of metacentric and subtelocentric chromosomes. Essad (1954) studied karyotypes of five species of *Lolium* and suggested three classes for the genus. A detailed investigation of karyotype was conducted by Malik and Thomas (1966).

In terms of the Stebbins' system, the karyotype of accessions are considered as median asymmetrical karyotypic in this system and this index cannot show the diversity between accessions in this species. The accessions had an asymmetry karyotype and variation was significant between accessions in karyotypic characters. Karyotypic evolution for L. multiflorum was more than intrachromosomal asymmetry, given the fact that the studied accessions and intrachromosomal asymmetry were increased by the development of ploidy level from diploidy to tetraploidy. Terrell (1968) had a comprehensive review on the genus Lolium. He recognized eight species in the genus and divided it into two sections based on breeding system. The accessions 374, 390 and vi displayed the highest karyotypic asymmetry based on intra and interchromosomal asymmetry and had the highest karyotypic evolution. The accessions 1765, 1624, 393 and usa had the moderate of karyotipic evolution and the accessions 1268, 1551, 374", 1151, 1766 and plc had the lowest karyotypic evolution based on intrachromosomal asymmetry index (A1) and interchromosomal asymmetry index (A2). Essad (1954) studied karyotypes of five species of Lolium and suggested three classes for the genus. In addition, the accessions 390, 374, vi and plc had the highest karyotypic evolution and the accessions 1624 and 1765 had the lowest one.

The results of variance analysis showed a significant variation (1% level) between the accessions in chromosomal traits. Cytogenetic divergence and extensive variations in karyotypic traits in Lolium germplasem were reported by other researchers (Mirzaei-Nodushan and Nadarkhani, 2001; Mirjalili, 2014). The results of principal components analysis based on karyotypic traits for accessions showed that the first two components accounted for 84.37% of the total variance. Multivariate methods of analysis, such as the PCA and cluster analyses used in the present study, revealed the germplasm groupings in many genetic resources such as Lolium (Mirjalili, 2014). According to the biplot, accessions 390, 374, vi and plc (Group 1) had the highest value based on the second component and the lowest values for the first component. So, these accessions had the highest chromosome length, intrachromosomal asymmetry index (A₁), interchromosomal asymmetry Index (A₂), and evolutionary accession based on karyotype characteristics. The accessions 1551, 393 and usa (Group 2) based on the first and second components showed the highest values implying that they had the highest chromosome length and asymmetric karyotype based on interchromosomal asymmetry and the lowest asymmetric karyotype based on intrachromosomal asymmetry. In addition, the accessions of Group 2 had the moderate of karyotipic evolution. The accessions 1151, 1268, 1766 and 374" (Group 3) had the lowest values for first and second components; therefore, they had the shortest chromosomes and the lowest symmetrical karyotypes based on interchromosomal asymmetry. The accessions of group 3 had the lowest karyotypic evolution based on intrachromosomal asymmetry. The accessions 1765 and 1624 (Group 4) had the lowest values for second component and the highest values for first component, so they had the shortest chromosomes and the lowest symmetrical karyotype. According to ploidy levels for accessions, we observed that the tetraploid accessions had the highest evolutionary of karyotypes. Mirjalili (2014) classified genotypes using karyotypic traits.

Literature Cited

- Che, Y.H. and Li, L.H. 2007. Genetic diversity of prolamines in *Agropyron mongolicum* Keng indigenous to northern China. Genetic Resources and Crop Evolution, 54: 1145-1151.
- Chowdhury, M.A., Vandenberg, V. and Warkentin, T. 2002. Cultivar identification and genetic relationship among selected breeding lines and cultivars in chickpea (*Cicer arietinum* L.). Euphytica, 127: 317–325.
- Cornish, M.A., Hayward, M.D. and Lawrence, M.J. 1979. Self-incompatibility in ryegrass. I genetic control in diploid *Lolium perenne*. Heredity, 43: 95-106.
- Essad, S. 1954. Contribution a la systematique du genere *Lolium*. Annales Institution Nationale Research Agronomie. Paris, Series B, Orsay 8 (Thesis).
- Evans, M. and Mecefield, B. 1974. Effects of B chromosomes homoeologus pairing in *Lolium* multiflorum × Lolium perenne species hybrids. Chromosoma, 45: 369-378.
- Gupta. P.K. 1995. Cytogenetic. Rastoginad Company, Marual, India. 3-4 p.
- Hajimoniri, M. 1999. Cytogenetic and morphometric studies on canola (*Brassica napus*). MSc. Thesis on plant science. North of Tehran Islamic Azad University.
- Hovin, A.W. and Hill, D. 1966. B chromosomes, tgeir origim and relation to meiosis in inter species *Lolium* hybrids. American Journal of Botany, 53: 702-708.
- Iruela, M., Rubio, J., Cubero, J.I., Gil, J. and Millan, T. 2002. Phylogenetic analysis in the genus *Cicer* and cultivated chickpea using RAPD and ISSR markers. Theoretical and Applied Genetics, 104: 643–651. Doi: 10.1007/s001220100751.
- Lamp, C.A., Forbes, S.J. and Cade, J.W. 2001. Grasses of temperate Australia- A fild guide. Inkata Press (1 st Edition) and CH Jerram & Associates Science Publishers (Revised Edition).
- Levan, A., Fredga, K. and Sandberg, A. 1964. Nomenclature for centrometric position on chromosomes. Hereditas, 52: 201-220.
- Lundqvist, A. 1962. The nature of the two loci incompatibility system in grasses. I. the hypothesis of a duplicative origin. Hereditas, 48: 153-168.
- Malik, C.P. and Thomas, P.T. 1966. Karyotypic studies in some *Lolium* and *Festuca* species. Caryologia, 19: 167-196.
- Mirjalili, S.A. 2014. Karyological studies in the genus *Lolium* in Iran. International Journal of Biosciences, 4 (4): 176 -183.
- Mirzaei-Nodushan, H. and Nadarkhani, H. 2001. Karytypic investigation of tetraploid populations of *Lolium* sp. Iranian Journal of Rangelands and Forests Plant Breeding and Genetic Research, 4: 87-116.
- Stebbins, G.L. 1971. Chromosomal evolution in higher plants. Edward Arnold, London.
- Sudupak, A., Akkaya, S. and. Kence, A. 2002. Analysis of genetic relationships among perennial and annual Cicer species growing in Turkey using RAPD markers. Theoretical and Applied Genetics, 105: 1220–1228. Doi: 10.1007/s00122-002-1060-8.
- Terrell, E.E. 1968. A taxonomic revision of the genus *Lolium*. Agricultural Research Service, US. Department of Agriculture. Technical Bulletin 1392.
- Wu, D. and Chen, L. 1991. The Karyotype analysis of oryzon ryegrass and three lines. Journal of Shanghai Agricultural College, 9: 253-259.
- Zwierzykowski, Z. and Naganowski, B. 1996. Taxonomy, cytogenetics and phylogenetic relationship in the *Lolium- Festuca* complex (*Poaceae*): I. *Lolium* - a review; Fragmenta Floristica ET Geobotanica, 41: 521-536.

How to cite this article:

Abbaszade, S., Ashraf Jafari, A., Safari, H., and Shirvani, H. 2017. Genetic variability and karyotype analysis for 13 accessions of *Lolium multiflorum*. *Journal of Ornamental Plants*, *7(1)*, *1-8*. URL: http://jornamental.iaurasht.ac.ir/article_528900_0134625dd7bf12d91ab55f67aa7585b0.pdf

