

Anatomo-Pathological Consequences of Mycotoxins Contamination in Rabbits Feed

Review Article

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Received on: 17 May 2019

Revised on: 28 May 2019

Accepted on: 15 Jun 2019

Online Published on: Sep 2019

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Online version is available on: www.ijas.ir

ABSTRACT

Mycotoxins are secondary metabolites produced by certain filamentous microscopic fungi, which occur naturally in the environment and may persist in animal feed. A mycotoxin contaminated diet may lead to feed refusal, poor feed conversion, diminished body weight gain and causes pathological effects associated with gross and histological changes, which are responsible for great economical losses. The present review summarizes the pathological lesions caused by the most widespread mycotoxins, aflatoxins, ochratoxins and their interaction, potentially hazardous ingredients of rabbit feed.

KEY WORDS feed, mycotoxins, pathological lesions, rabbits.

INTRODUCTION

Mycotoxins are secondary metabolites produced by certain filamentous microscopic fungi, which occur naturally in the environment and can commonly grow on a variety of crops, including wheat, maize and soybean (Goswami and Kistler, 2004; Murphy *et al.* 2006; Marin *et al.* 2013). They comprise a group of several hundreds of chemically different toxic compounds (Sweeney and Dobson, 1998) and their occurrence in cereal grains and animal feed have been reported worldwide (Placinta *et al.* 1999). Food and feed-stuffs prepared using mycotoxin-containing crops deteriorate nutritional content and represent a potential risk for animal and human health (Hussein and Brasel, 2001; Bennett and Klick, 2003).

The mycotoxins are cytotoxic, disrupting various cellular structures such as membranes, and interfering with vital cellular processes such as protein, RNA and DNA synthesis (Guerra *et al.* 2000). They destroy the tissues by oxidizing proteins, have immunosuppressive effects and increase dis-

eases incidence (Kumar *et al.* 2008). Mycotoxins may cause fever (Cannon *et al.* 1982), gastrointestinal problems, internal bleeding, haemorrhages or bruising, stomach ulcers (Aziz *et al.* 1995), mouth sores, kidney or liver damage (Szilágyi *et al.* 1994), central nervous system problems (Gabal *et al.* 1986), immune-suppression (Richard *et al.* 1991; Kumar *et al.* 2008), tumour-genesis, eye and lung problems, hypertrophy of the adrenal cortex, reproductive organ problems (Szilágyi *et al.* 1994), damage to the heart muscle, tachycardia, skin problems (Fairhurst *et al.* 1987), bone marrow and spleen problems (Niyo *et al.* 1988), blood abnormalities (Mizutani *et al.* 1997), rectal prolapses and increased vascular fragility.

In addition to the toxic effects, a mycotoxin contaminated diet may lead to other consequences like feed refusal, poor feed conversion and diminished body weight gain (Council on Agricultural Science and Technology, 1989; Council on Agricultural Science and Technology, 2003; Kolpin *et al.* 2014) which are responsible for great economical losses. These effects, in both humans and animals, depend on a

number of factors including intake levels, duration of exposure, toxin species, mechanisms of action, metabolism, and defense mechanisms (Galvano *et al.* 2001; Hussein and Brasel, 2001).

The most frequent sources of mycotoxins are the fungal genera *Aspergillus*, *Fusarium*, and *Penicillium* (Bennett and Klich, 2003). While *Fusarium* species are destructive plant pathogens producing mycotoxins before or immediately post harvesting, *Penicillium* and *Aspergillus* species are more commonly found as contaminants of commodities and foods during drying and subsequent storage (Sweeney and Dobson, 1998). Weather conditions, including moisture level and temperature have greatest influence on mold growth and mycotoxin production: while *Aspergillus* prefers warmer tropical areas, *Fusarium* and *Penicillium* also grow in European temperate areas (Bryden, 2012).

Rabbit feed ingredients that constitute complete feed products are derived from different raw materials and the contamination of feed materials would represent an important potential hazard. Contamination of animal feed with mycotoxins still occurs very often, despite great efforts in preventing it. Multitoxin studies on animal feed reported that 75-100% of samples analysed contained more than one mycotoxin (Streit *et al.* 2012; Schatzmayr and Streit, 2013).

Two environmentally important mycotoxins, which have gained an immense importance due to their biological effects and widespread toxicity, are ochratoxin A (OTA) and aflatoxin B₁ (AFB₁). WHO-IARC, (1993) designated AFB₁ as group-1 and OTA as group-2B carcinogen (Hussein and Brasel, 2001) and are most frequently detected in the agricultural commodities (Bennett and Klich, 2003; Gabarty and Abou, 2016; Jan *et al.* 2017).

Aflatoxins are a family of extremely toxic, mutagenic and carcinogenic compounds produced by fungi belonging to genus *Aspergillus flavus* and *Aspergillus parasiticus* (Jacobsen *et al.* 2007; Cortes *et al.* 2010); ochratoxins, potentially as important as the aflatoxins (Bennett and Klich, 2003), are commonly produced by two species of fungi, *Penicillium verrucosum* Dierckx and *Aspergillus ochraceus* Wilhelm (Frisvad and Samson, 1991). The alarming feature of mycotoxins is their occurrence in combination and in low dose levels frequently in cereals / feedstuffs which may exert a greater degree of damage (additive or synergistic) in comparison to the individual effects (Roberts *et al.* 1981; Raina and Singh, 1991; Adebajo, 1993; Sundaram *et al.* 1999; Prabu *et al.* 2013).

The toxicity and clinical signs observed in animals when more than one mycotoxin is present in feed are complex and diverse (Prabu *et al.* 2013). The effects observed during multiple mycotoxin exposure can differ greatly from the effects observed in animals exposed to a single mycotoxin (Huff *et al.* 1988).

Animals are usually exposed to mycotoxins through their diet and depending on different factors such as age, sex, route of administration: this can result in acute, sub-acute or chronic mycotoxicosis. Rabbits are highly susceptible to mycotoxins and they can have very pervasive yet sub-clinical effects on their health which very often remain unnoticed: when the clinical symptoms of mycotoxin poisoning can be observed, significant damage has already occurred.

The aim of this review is to summarize the toxicopathological effects caused by the most widespread mycotoxins (aflatoxins and ochratoxin and their interactions), potentially hazardous ingredients of rabbit feed.

Aflatoxins

Aflatoxins are a group of naturally occurring carcinogens that are known to contaminate different food stuffs causing serious consequences in human and animal health. Aflatoxins have been reported to affect the various body organs like liver, kidneys, lungs, brain, testes and many endocrine and exocrine organs, the heart, skeletal muscles and the different body systems (Bbosa *et al.* 2013).

They are produced by fungi belonging to genus *Aspergillus flavus* and *Aspergillus parasiticus* (Jacobsen *et al.* 2007; Cortes *et al.* 2010). The four major known aflatoxins include AFB₁, AFB₂, AFG₁, and AFG₂ where the "B" and the "G" refer to the blue and green fluorescent colors produced under UV light on thin layer chromatography plates, while the subscript numbers 1 and 2 indicate major or minor compounds (Thrasher, 2012; Bbosa *et al.* 2013). The aflatoxins display potency of toxicity, carcinogenicity, mutagenicity in the order of AFB₁ > AFG₁ > AFB₂ > AFG₂ (Cortes *et al.* 2010). AFG₂ occurs in high quantities though less toxic while AFB₁ is the most toxic of all the aflatoxin. The World Health Organization (WHO) classifies AFB₁ as a class 1 carcinogen (WHO-IARC, 2000).

Aflatoxicosis is a condition caused by aflatoxins which can result in acute, sub-acute or chronic mycotoxicosis. The acute primary aflatoxicosis is produced when moderate to high levels of aflatoxins are consumed; the chronic primary aflatoxicosis results from ingestion of low to moderate levels of aflatoxins (USAID, 2012)

Animals get exposed to aflatoxins by two major routes: (a) by direct ingestion of aflatoxin-contaminated foods (Agag, 2004) and (b) by inhalation of dust particles of aflatoxins, especially AFB₁ in contaminated foods in industries and factories (Coulombe, 1994).

The lethal toxicity of AFB₁ varies in different animals. Rabbits are highly susceptible to aflatoxins having the least median lethal dose (LD₅₀) of any animal species studied (Prabu *et al.* 2013; Cardona *et al.* 1991; Lakkawar *et al.* 2004).

AFB₁ as low as 15 mg/kg feed causes high levels of morbidity and mortality (Makkar and Singh, 2009; Mezes and Balogh, 2009).

Experimental AFB₁ toxicosis is known to cause alterations in enzyme levels along with patho-anatomical changes in vital organs (Clark *et al.* 1980; Abdelhamid *et al.* 1990; Sahoo *et al.* 1993). Short exposure to large doses of aflatoxins produces acute toxicity including fever oedema, vomiting, abdominal pain, inappetence, lethargy ataxia, rough hair coat and pale potentially fatal liver failure (Lakkawar *et al.* 2004).

The prolonged rabbit feeding of AFB₁ in a diet, beyond 40 days, results in cumulative toxicosis which is manifested by altered clinical signs. Symptoms are characterized by anorexia, diarrhoea, decrease in body weight gain, lethargy, fur chewing and dehydration (Clark *et al.* 1980; Abdelhamid *et al.* 1990; Prabu *et al.* 2013; Lakkawar *et al.* 2004). It has been reported that a decrease in body weight is one of the earliest indicators of clinical aflatoxicosis in animals (Clark *et al.* 1980).

The gastrointestinal tract (GIT) is the main route of entry of aflatoxins as a result of consumption of aflatoxin-contaminated foods especially AFB₁. Orally-ingested AFB₁ is most efficiently absorbed from the small intestine at the duodenum (Kumagai, 1989) and the effects of AFB₁ on the GIT are indirect following alterations in the liver's detoxification mechanism and a possible reduction in nutrient uptake (Stetinova *et al.* 1998).

The liver and kidneys are the most affected organs followed by stomach, intestine, lungs, heart, spleen, gonads, thyroid and brain (Lakkawar *et al.* 2004; Prabu *et al.* 2013). AFB₁ is a potent hepatotoxic and hepato-carcinogenic mycotoxin which requires bio-activation to an active AFB₁-8,9-epoxide (Essigmann *et al.* 1982) which binds to DNA resulting in abnormal cellular proliferation, leading to mutagenesis and carcinogenesis (Guengerich, 2001). AFB₁ is metabolized in the liver by the cellular cytochrome-P450 enzyme system to form the reactive intermediate, AFB₁-8,9-epoxide, which in turn reacts with macromolecules such as lipids and DNA, leading to lipid peroxidation and cellular injury (Stresser *et al.* 1994). The mitochondrial damage following aflatoxicosis can result in a decrease in the oxidation of fats by these organelles, with a concomitant accumulation of lipids, necrotic changes, fibroblastic proliferation, mononuclear cellular infiltration, bile duct hyperplasia (Sahoo *et al.* 1993; Mclean and Dutton, 1995; Lakkawar *et al.* 2004). Icterus observed in terminal stages may be due to an increase in the levels of bilirubin as a sequela to hepatic necrosis and cholestasis resulting from decreased cytochromed P450, increased heme oxygenase and biliverdin reductase activities along with an increase in heme catabolism in the liver (Guerre *et al.* 1997). AFB₁ has been re-

ported to cause pallor discoloration of liver and hepatomegaly, congestion of liver parenchyma, cytoplasmic vacuolation or fatty change of hepatocytes, necrosis of hepatocytes and newly formed bile ducts (Krishna *et al.* 1991; Sahoo *et al.* 1993; Churamani *et al.* 1995; Singh *et al.* 1999; Vinita *et al.* 2003; Lakkawar *et al.* 2004).

Haemolytic anaemia and strong cytotoxic effects have been also observed (Verma and Metha, 1998).

The renal lesions are secondary to those observed in the liver. The target site of action of AFB₁ is the glomerular region causing reduction in the glomerular filtration rate and glucose reabsorption (Glahn *et al.* 1991). Kidneys appear congested, slightly enlarged and show a moderate degree of nephrosis (Ahmed *et al.* 2012). The urinary bladder appears distended with tick yellowish turbid urine (Lakkawar *et al.* 2004).

Aflatoxin have reported to have serious acute effects on the respiratory systems. Besides the liver, the lung and trachea are also capable of activating AFB₁ and rabbit lung and tracheal microsomes show high activity for this reaction (Daniels *et al.* 1990). Bio-activation related toxicity of AFB₁ has also been observed in tracheal mucosa following intra-tracheal instillation of AFB₁ in rabbits (Mezes, 2008; Coulombe *et al.* 1986). The pulmonary inflammation and oedematous changes observed might also be due to the production of eicosanoids stimulated by the AFB₁. This might have also contributed to the manifestation of dyspnoea in the early stages of toxicosis (Massey *et al.* 1995; Lakkawar *et al.* 2004).

Furthermore, aflatoxins have been reported to disrupt the reproductive system in both male and female animals after ingestion of aflatoxin-contaminated foods. Short exposure to large doses of aflatoxins produces acute toxicity; sub-symptomatic exposure to aflatoxins is known to produce male reproductive toxic effects with several manifestation.

The principal target organ in causing male reproductive toxicity is the testis and various aspects of spermatogenesis and androgen biosynthesis are affected (Baker and Greene, 1987; Sahoo *et al.* 1993). However, the epididymis and vas deferens also are a target for the action of such reproductive toxicants (Akbarsha *et al.* 2000). The severity of pathological changes in epididymis are aflatoxins (AF) dose dependent and summarized as interstitial oedema and atrophy of epididymal tubules, associated with congestion of the blood vessels and capillaries.

The seminiferous tubules show degeneration of the epithelium and a reduction in the number of mature spermatids in aflatoxin treated rabbits (Lakkawar *et al.* 2004). Salem *et al.* (2001) reported a relative decrease in testes weight and an increase in the number of abnormal/dead sperms following a 9-week administration of sublethal doses of AFB₁ to mature male rabbits.

Aflatoxins have reported to have serious acute effects on cardiovascular systems including vascular fragility, endothelial injury and haemorrhaging in tissues (Harriet, 2003).

Heart can show epicardial congestion (Lakkawar *et al.* 2004).

The coagulation defect and bleeding associated with aflatoxicosis has previously been attributed to either vitamin K antagonism, decreased hepatic protein and coagulation factor synthesis or disseminated intravascular coagulation secondary to hepatocellular degeneration and necrosis (Baker and Greene, 1987).

In the brain or central nervous system, the neurons have a high metabolic rate but little capacity for anaerobic metabolism and subsequently, inadequate oxygen flows to the brain kills the neuronal brain cells within minutes. Aflatoxins and its metabolites and other products such as the reactive oxygen species (ROS) like the AFB-8,9-epoxides may interfere with the normal functioning of the nerve cells by forming DNA adducts, protein adducts, oxidative stress factors, mitochondrial directed apoptosis of the nerve cells as well as inhibiting their synthesis of protein, RNA and DNA (Bbosa *et al.* 2013). AFB₁ is known to alter the distribution of acetylcholine esterase (AChE) in the brain affecting cholinergic transmissions at the nerve endings and thus can result in manifestations of nervousness and behavioural deficiency (Egbunike and Ikegwuonu, 1984; Lakkawar *et al.* 2004). Sahoo *et al.* (1993) reported vascular congestion and focal mononuclear infiltration in the meninges along with perivascular cuffing, mild neuronal degeneration and gliosis following the oral administration of AFB₁ to New Zealand White Rabbits at the rate of 0.0625 mg/day/animal for a period of 30 days.

AFB₁ is recorded to be teratogen in rabbits (El-Nahla *et al.* 2013). The dose of 0.1 mg/kg AFB₁ body weight AFB₁ is the minimum teratogenic dose that interfere with intrauterine development during 6th-18th days of gestation (Wangikar *et al.* 2005).

Ochratoxins

Ochratoxin, commonly produced by two species of fungi, *Penicillium verrucosum* and *Aspergillus ochraceus* (Frisvad and Samson, 1991; Ostry *et al.* 2013; Saleemi *et al.* 2015; Valtchev *et al.* 2015), is potentially as important as the aflatoxins (Bennett and Klich, 2003). The most abundant and most toxic mycotoxin within the ochratoxins is ochratoxin-A (OT-A) (Marquardt and Frohlich, 1992), which occurs in maize, cereal grains such as wheat and barley and oil seeds such as soybean and peanuts (Manning *et al.* 2003). The OT-A is toxic to numerous animal species, the kidney being the main target organ in birds and mammals, but not adult ruminants (Pfohl-Leszkowicz and Manderville, 2007). The most consistent effect of ochratoxicosis in different species

including rodents, guinea pigs, swine and poultry is growth depression (Thacker and Carlton, 1977; Dwivedi and Burns, 1985; Stein *et al.* 1985; Dwivedi and Burns, 1986; Rati *et al.* 1991; Harvey *et al.* 1994).

OT-A is well known nephrotoxic, hepatotoxic, immunosuppressive, mutagenic, cardio-vascular toxic, teratogenic and possible human carcinogen (Ahmed *et al.* 2012; Hussain *et al.* 2016). In addition to these effects, OT-A is also considered as potent myelotoxic agent (Moura *et al.* 2004).

OT-A is also extremely cytotoxic and may cause red blood cell haemolysis in rabbits (Zofair *et al.* 1996). OT-A at low doses influences energy metabolism such as carbohydrate, amino acid, cofactors and vitamins. However, in the high doses pathways influenced by OT-A are associated with the different body systems including circulatory, digestive, endocrine, excretory system (Jan *et al.* 2017).

Rabbits are comparatively more susceptible to OT-A than mice, rats and guinea pigs (Ponnuchamy, 2000). OT-A has been observed to be acutely toxic to young rabbits with LD50 of 10 mg OT-A/kg body weight (Mir *et al.* 1999).

Long term exposures have been observed to favor accumulation in the tissues and cause severe impairment of health, wide spread pathoanatomical alterations, and even death (Mir and Dwivedi, 2000; Mir and Dwivedi, 2010). OT-A has been known to be actively reabsorbed through the proximal convoluted tubules (Stein *et al.* 1985). OT-A produces significant nephrotoxicity with the pale, soft and enlarged kidneys showing discoloured foci over the surface. The proximal tubule of the kidney is the primary site targeted in OT-A induced nephrotoxicity (Suzuki *et al.* 1975). The reason behind the high sensitivity of kidneys to the toxin might be correlated to the fact that the kidneys constitute the primary excretory pathway for OA and its heavy blood circulation (25% of cardiac output) (Marquardt and Frohlich, 1992).

Testis, being an organ with rapid meiotic and mitotic cell divisions, could be a possible target of OT-A as it inhibits DNA, RNA and protein synthesis (Marquardt and Frohlich, 1992). OT-A (~99%) is bound to plasma proteins, mainly albumin, which can't be excreted by glomerular filtrate. However, unbound portion (~1%) can be found in the urinary filtrate. The remaining OT-A is only excreted via organic anion transporter (OAT) route, which prone the proximal tubular epithelial cells to damage, by virtue of depletion of indigenous dicarboxylic acid (glutarate, ketoglutarate) on expense of OT-A internalization (Sekine *et al.* 2006; Khatoon *et al.* 2016). Several natural and experimental OT-A exposure studies have recorded similar changes in the kidney function (Khan *et al.* 2014). The increase in the serum biomarkers of kidneys damage, and the mechanistic nephrotoxicity associated with the OT-A, has

also been augmented by histological alteration in the tubular cells of nephrons (Jan *et al.* 2017). Microscopically, kidneys reveal degeneration of the proximal convoluted tubules and the testes atrophic (Prabu *et al.* 2013).

OT-A has been reported to cause direct effects on lymphocytes and plasma cells in primary and secondary lymphoid organs (Dwivedi and Burns, 1985).

OT-A has also been reported to be teratogenic in rabbits (Wangikar *et al.* 2005), rats (Still *et al.* 1971; Brown *et al.* 1976; Mayura *et al.* 1982; Abdel-Wahhab *et al.* 1999), mice (Hayes *et al.* 1974; Arora and Fr^ool, 1981), hamsters (Hood *et al.* 1976). Teratogenic effects were found among the 0.1 mg/kg dose group in the form of a significant increase in the incidence of gross anomalies (wrist drop, rudimentary tail, knuckling of fetlock and agenesis of tail), skeletal anomalies (agenesis of caudal vertebrae, incomplete ossification of skull bones and wavy ribs) and soft tissue anomalies (internal hydrocephalus, microphthalmia and kidney agenesis).

Ochratoxicosis induces also biochemical alterations in the rabbits (Mir and Dwivedi, 2010).

Aflatoxins and ochratoxins interaction

The alarming feature of mycotoxins is their occurrence in combination and in low dose levels frequently in cereals/feedstuffs which may exert a greater degree of damage to animal health (additive or synergistic) in comparison to the individual effects (Roberts *et al.* 1981; Raina and Singh, 1991; Adebajo, 1993; Sundaram *et al.* 1999; Prabu *et al.* 2013). The effects observed during multiple mycotoxin exposure can differ greatly from the effects observed in animals exposed to a single mycotoxin (Huff *et al.* 1988).

Simultaneous occurrence of these two mycotoxins in feedstuffs under natural conditions was observed during screening of different food and feed stuffs (Mir, 1998).

Although the scientific literature offers a broad variety of information on the effects of individual mycotoxins in various animal species, concurrent exposure to multiple mycotoxins is more likely in the livestock industry (Zaki *et al.* 2012). Occurrence of spontaneous aflatoxicosis and ochratoxicosis, individually and in interaction, the combined presence of low dose levels of AFB₁ ranging from 0.03 to 2.06 ppm and OT-A ranging from 0.01 to 1.23 ppm in cereals/feedstuffs have been frequently reported in different species of animals (Raina and Singh, 1991; Dwivedi *et al.* 2004).

There exists an interaction in terms of synergism in the toxicological effects when both the chemicals are fed together. Heavy mortalities of up to 75% in German Angora rabbits in Kangra district of Himachal Pradesh (India) were reported due to simultaneous spontaneous aflatoxicosis and ochratoxicosis with high levels of AFB₁ and OT-A in the

feed (Sharma, 1998). In experimental simultaneous exposure to OT-A e AFB₁ in rabbits, during long term feeding of AFB₁ and OT-A in combination, the intensity of clinical symptoms was comparatively more severe as than when either of these mycotoxins was given alone (Doerr and Huff, 1981; Wangikar *et al.* 2005; Prabu *et al.* 2013). The changes were more severe as evidenced by gross, histological, ultra structural, immunological and enzyme antioxidant changes observed both in liver and kidneys, suggesting an additive interaction of AFB₁ and OT-A in rabbits (Prabu *et al.* 2013). Similar presence of changes both in the liver and kidney has been reported by several workers in guinea pigs (Richard *et al.* 1975). OT-A and AFB₁, when administered in combination in pregnant rabbits, resulted in significant increase in the incidence of various fetal anomalies and post implantation losses and decreased fetal weights (Wangikar *et al.* 2005). Testis revealed severe disruption of the normal spermatogonial cell pattern and marked atrophic changes (Prabu *et al.* 2013). Similar results were observed in previous investigation with different combinations of OT-A and AFB₁ in rats (Wangikar *et al.* 2004b; Wangikar *et al.* 2004c), suggesting that in combination, these toxins might have possibly antagonistic interaction.

Mycotoxins comprise a group of several hundreds of chemically different toxic compounds (Moss, 1996; Rotter *et al.* 1996; Sweeney and Dobson, 1998) and their occurrence in cereal grains and animal feed have been reported worldwide (Placinta *et al.* 1999). Food and feedstuffs prepared using mycotoxin-containing crops deteriorate nutritional content and represent a potential risk for animal and human health (Hussein and Brasel, 2001; Bennett and Klick, 2003).

Consumption of a mycotoxin contaminated diet may induce acute and long-term chronic effects resulting in teratogenic, carcinogenic, and oestrogenic or immunosuppressive effects. Direct consequences of consumption of mycotoxin contaminated animal feed include: reduced feed intake, feed refusal, poor feed conversion, diminished body weight gain, increased disease incidence (due to immunosuppression), and reduced reproductive capacities (Fink-Gremmels and Malekinejad, 2007; Morgavi and Riley, 2007; Pestka, 2007; Voss *et al.* 2007) which leads to economic losses (Huwig *et al.* 2001; Wu, 2004; Wu, 2006).

It was widely reported that aflatoxins affect the various body organs like liver, kidneys, lungs, brain, testes and many endocrine and exocrine organs, the heart, skeletal muscles and the different body systems (Bbosa *et al.* 2013).

In particular, rabbits are highly susceptible to aflatoxins having the least median lethal dose (LD₅₀) of any animal species studied (Prabu *et al.* 2013; Cardona *et al.* 1991; Lakkawar *et al.* 2004). AFB₁ as low as 15 mg/kg feed causes high levels of morbidity and mortality (Makkar and

Singh, 2009; Mezes and Balogh, 2009). OT-A has been observed to be acutely toxic to young rabbits with LD50 of 10 mg OT-A/kg body weight (Mir *et al.* 1999). Rabbits are comparatively more susceptible to OT-A than mice, rats and guinea pigs (Ponnuchamy, 2000). Long term exposures have been observed to favor accumulation in the tissues and cause severe impairment of health, wide spread pathological anatomical alterations, and even death (Mir and Dwivedi, 2000; Mir and Dwivedi, 2010). OT-A has been known to be actively reabsorbed through the proximal convoluted tubules (Stein *et al.* 1985).

Multi-toxin occurrence may be one important explanation for divergences in effect levels described in the scientific literature, where defined, mostly purified mycotoxins are used in most studies. In field outbreaks, naturally contaminated feeds may contain multiple mycotoxins and thus apparently lower contamination levels of a single specific mycotoxin can be associated with more severe effects (Zaki *et al.* 2012). The presence of AFB₁ and OT-A, both alone and in interaction at very low dose levels of 0.5-1 ppm in animal feeds and cereals can cause significant toxicity in rabbits with gross and histological changes and might be a potential threat to animal health (Prabu *et al.* 2013).

CONCLUSION

Based on literature review and data available, it has to be pointed out that far more work has to be done on this particular research field, especially in case of mycotoxins subacute contamination range as well as with combinations of more than two toxins.

REFERENCES

- Abdelhamid A.M., El-Shawaf I., El-Ayoty S.A., Ali M.M. and Gamil T. (1990). Effect of low level of dietary aflatoxins on Baladi rabbits. *Arch. Tierenahr.* **45**, 517-537.
- Abdel-Wahhab M.A., Nada S.A. and Arbid M.S. (1999). Ochratoxicosis: Prevention of developmental toxicity by l-methionine in rats. *J. Appl. Toxicol.* **19**, 7-12.
- Adebajo L.O. (1993). Survey of aflatoxins and ochratoxin A in store tubers of *Cyperus esculantus*. *Mycopathologia.* **124**, 41-46.
- Agag B.I. (2004). Mycotoxins in foods and feeds: Aflatoxins. *Ass. Univ. Bull. Environ. Res.* **7**, 173-191.
- Ahmed K., El Mahady M.M., Badawy S.A., Ahmed Y.F. and Aly M.A. (2012). Pathological consequences of aflatoxins in male rabbit: cellular, genetic and oxidative damage. *Glob. Vet.* **8**, 721-731.
- Akbarsha M.A., Averal H.I., Girija R., Anandhi S. and Faridha B.A. (2000). Male reproductive toxicity of vincristine: Ultrastructural changes in the epididymal epithelial apical cell. *Cytobios.* **102**, 85-93.
- Arora R.G. and Fr'ol'en H. (1981). Interference of mycotoxins with prenatal development of the mouse. II. Ochratoxin A induced teratogenic effects in relation to the dose and stage of gestation. *Acta Vet. Scandinavica.* **22**, 534-552.
- Aziz N.H., El-Aziz A.M.A. and Omran R.M.A. (1995). Effects of T-2 mycotoxin on histopathological changes in rabbits. *Bio-med. Lett.* **51**, 271-281.
- Baker D.C. and Greene R.A. (1987). Coagulation defects of aflatoxin induced rabbits. *Vet. Pathol.* **24**, 62-70.
- Bbosa G.S., Kytia D., Lubega A., Ogwal-Okeng J., Anokbongbo W.W. and Kyegombe D.B. (2013). Review of the biological and health effects of aflatoxins on body organs and body systems Pp. in Aflatoxins - Recent Advances and Future Prospects, M. Razzaghi-Abyaneh, Ed. Intech Construction Inc., Philadelphia, USA.
- Bennett J.W. and Klich M. (2003). Mycotoxins. *Clin. Microbiol. Rev.* **16**, 497-516.
- Bryden W.L. (2012). Mycotoxin contamination of the feed supply chain: Implications for animal productivity and feed security. *Anim. Feed Sci. Technol.* **173**, 134-150.
- Brown M.H., Szczech G.M. and Purmalis B.P. (1976). Teratogenic and toxic effects of ochratoxin A in rats. *Toxicol. Appl. Pharmacol.* **37**, 331-337.
- Cannon M., Cranston W.I., Hellon R.F. and Townsend Y. (1982). Inhibition, by trichothecene antibiotics, of brain protein synthesis and fever in rabbits. *J. Physiol.* **322**, 447-455.
- Cardona T.D., LLangantileke S.G. and Noomhorm A. (1991). Aflatoxin research on grain in Asia: its problems and possible solutions. Pp. 309-322 in Mycotoxin Prevention and Control in Foodgrains. R.L. Semple, A.S. Frio, P.A. Hicks and J.V. Lozare, Eds. FAO, Bangkok, Thailand.
- Churamani C.P., Chattopadhyay S.K., Pawaiya R.S. and Johri T.S. (1995). Patho-anatomical studies on cumulative aflatoxicosis induced with low dose in rabbits. *Indian J. Vet. Pathol.* **19**, 119-122.
- Clark J.D., Jain A.V. and Hatch R.C. (1980). Experimentally induced chronic aflatoxicosis in rabbits. *Am. J. Vet. Res.* **41**, 1841-1845.
- Cortés G., Carvajal M., Méndez-Ramírez I., Ávila-González E., Chilpa-Galván N., Castillo-Urueta P. and Flores C.M. (2010). Identification and quantification of aflatoxins and aflatoxicol from poultry feed and their recovery poultry litter. *Poult. Sci.* **89**, 993-1001.
- Coulombe R.A.Jr., Wilson D.W., Hsieh D.P.H., Plopper C.G. and Serabjit-Singh C.J. (1986). Metabolism of aflatoxin B₁ in the upper airways of the rabbit: Role of the nonciliated tracheal epithelial cell. *Cancer Res.* **46**, 4091-4096.
- Coulombe R.A.Jr. (1994). Nonhepatic disposition and effects of aflatoxin B₁. Pp. 89-101 in The Toxicology of Aflatoxins: Human Health, Veterinary and Agricultural significance. D. Eaton and J. Groopman, Eds. Academic Press, USA.
- Council on Agricultural Science and Technology (CAST) Report. (1989). Mycotoxins: Economics and Health Risks. Task Force Report No. 116, Ames, Iowa, USA.
- Council on Agricultural Science and Technology (CAST) Report. (2003). Mycotoxins: Risks in plant, animal, and human systems. Task Force Report No. 139, Ames, Iowa, USA.
- Daniels J.M., Liu L., Stewart R.K. and Massey T.E. (1990). Bio-transformation of aflatoxin B₁ in rabbit lung and liver microsomes. *Carcinogenesis.* **11**, 823-827.

- Doerr J.A. and Huff W.E. (1981). Effect of young broiler chickens of combining low levels of dietary aflatoxin and ochratoxin A. *Poult. Sci.* **60**, 1648-1660.
- Dwivedi P. and Burns R.B. (1985). Immunosuppressive effects of ochratoxins A in young turkeys. *Avian Pathol.* **14**, 213-221.
- Dwivedi P. and Burns R.B. (1986). The natural occurrence of ochratoxin A and its effects in poultry. A review. Part I: epidemiology and toxicology. *World's Poult. Sci. J.* **42**, 32-42.
- Dwivedi P., Sharma A.K. and Telang A.G. (2004). NATP-CGP Project on Ochratoxicosis in Animals with Special Reference to Teratogenicity, Diagnosis and Ameliorative Measures. Annual Report Division of Pathology, IVRI, Izatnagar, New Delhi, India.
- Egbunike G.N. and Ikegwonu F.L. (1984). Effect of aflatoxicosis on acetyl- cholinesterase activity in the brain and adenylophysis of the male rat. *Neurosci. Lett.* **52**, 171-174.
- El-Nahla S., Imam H., Moussa E., Ibrahim A. and Ghanam A. (2013). Teratogenic effects of aflatoxin in rabbits (*Oryctolagus cuniculus*). *J. Vet. Anat.* **6**, 67-85.
- Essigmann J.M., Croy R.G., Bennett R.A. and Wogan G.N. (1982). Metabolic activation of aflatoxin B₁: Patterns of DNA adduct formation, removal and excretion in relation to carcinogenesis. *Drug Metab. Rev.* **13**, 581-602.
- Fairhurst S., Maxwell S.A., Seawin J.W. and Swanston D.W. (1987). Skin effects of trichothecenes and their amelioration by decontamination. *Toxicology.* **46**, 307-319.
- Fink-Gremmels J. and Malekinejad H. (2007). Clinical effects and biochemical mechanisms associated with exposure to the mycoestrogen zearalenone. *Anim. Feed Sci. Technol.* **137(3)**, 326-341.
- Frisvad J.C. and Samson R.A. (1991). Filamentous fungi in foods and feeds: ecology, spoilage and mycotoxin production. Pp. 31-68 in Handbook of applied mycology: Food and Feeds. D.K. Arora, K.G. Mukerji and E.H. Marth, Eds. Marcel Dekker, New York.
- Gabal M.A., Awad Y.L., Morcos M.B., Barakat A.M. and Malik G. (1986). Fusario toxicoses of farm animals and mycotoxic leucoencephalomalacia of the equine associated with the finding of trichothecenes in feedstuffs. *Vet. Hum. Toxicol.* **28**, 207-212.
- Gabarty A. and Abou El Nour S. (2016). Impact of wheat flour infestation by some insects on its quantity and quality loss, fungal contamination and mycotoxins. *Int. J. Agric. Biol.* **18**, 1122-1130.
- Galvano F., Piva A., Ritieni A. and Galvano G. (2001). Dietary strategies to counteract the effects of mycotoxins: A review. *J. Food Prot.* **64**, 120-131.
- Glahn R.P., Beers K.W., Bottje W.G., Wideman Jr R.F. and Huff W.E. (1991). Aflatoxicosis alters avian renal function, calcium and vitamin D metabolism. *J. Toxicol. Environ. Health.* **34**, 309-321.
- Goswami R. and Kistler H.C. (2004). Heading for disaster: *Fusarium graminearum* on cereal crops. *Mol. Plant Pathol.* **5**, 515-525.
- Guengerich F.P. (2001). Forging the links between metabolism and carcinogenesis. *Mutat. Res.* **488**, 195-209.
- Guerre P., Burgat V. and Galtier P. (1997). Dose-related increase in liver heme catabolism during rabbit aflatoxicosis. *Toxicol. Lett.* **92**, 101-108.
- Guerre P., Eeckhoutte C., Burgat V. and Galtier P. (2000). The effects of T-2 toxin exposure on liver drug metabolizing enzymes in rabbit. *Food Addit. Contam.* **17**, 1019-1026.
- Harriet A.M. (2003). Is indoor mold contamination a threat to health? *J. Environ. Health.* **62**, 0022-0892.
- Harvey R.B., Kubena L.F., Elissalde M.H., Rottinghaus G.E. and Corrier D.E. (1994). Administration of ochratoxin A and T-2 toxin to growing swine. *Am. J. Vet. Res.* **55**, 1757-1761.
- Hayes W.A., Hood R.D. and Lee H.L. (1974). Teratogenic effects of ochratoxin A in mice. *Teratology.* **9**, 93-97.
- Hood R.D., Naughton M.J. and Hayes W.A. (1976). Prenatal effects of ochratoxin A in hamsters. *Teratology.* **13**, 11-14.
- Huff W.E., Kubena L.F., Harvey R.B. and Doerr J. (1988). Mycotoxin interactions in poultry and swine. *J. Anim. Sci.* **66**, 2351-2355.
- Hussain Z., Khan M.Z., Saleemi M.K., Khan A. and Rafique S. (2016). Clinicopathological effects of prolonged intoxication of aflatoxin B₁ in broiler chicken. *Pakistan Vet. J.* **36**, 477-481.
- Hussein H.S. and Brasel J.M. (2001). Toxicity, metabolism and impact of mycotoxins in humans and animals. *Toxicology.* **167**, 101-134.
- Huwig A., Freimund S., Kappeli O. and Dutler H. (2001). Mycotoxin detoxification of animals feed by different adsorbents. *Toxicol. Lett.* **122**, 179-188.
- Jacobsen B.J., Coppock R.W. and Mostrom M.S. (2007). Mycotoxins and mycotoxicoses. Extension Bulletin, Montana State University, Bozeman, Montana.
- Jan R., Sadique U., Hassan Z.U., Farid K., Ahmad S., Khan S. and Khan H. (2017). Toxicopathological and reproductive effects of concurrent oral administration of ochratoxin A and endosulfan in pregnant rabbits (*Oryctolagus cuniculus*). *Pakistan Vet. J.* **37**, 19-24.
- Khan W.A., Khan M.Z., Khan A., Ul Hassan Z. and Saleemi M.K. (2014). Potential for amelioration of aflatoxin B₁-induced immunotoxic effects in progeny of White Leghorn breeder hens co-exposed to vitamin E. *J. Immunotoxicol.* **11**, 116-125.
- Khatoun A., Khan M.Z., Khan A. and Javed I. (2016). Toxicopathological and serum biochemical alterations induced by ochratoxin a in broiler chicks and their amelioration by locally available bentonite clay. *Pakistan J. Agric. Sci.* **53**, 977-984.
- Kolpin D.W., Schenzel J., Meyer M.T., Phillips P.J., Hubbard L.E., Scott T.M. and Bucheli T.D. (2014). Mycotoxins: Diffuse and point source contributions of natural contaminants of emerging concern to streams. *Sci. Total Environ.* **470**, 669-676.
- Krishna L., Dawra R.K., Vaid J. and Gupta V.K. (1991). An outbreak of aflatoxicosis in Angora rabbits. *Vet. Hum. Toxicol.* **32**, 159-161.
- Kumagai S. (1989). Intestinal absorption and excretion of aflatoxins in rats. *Toxicol. Appl. Pharmacol.* **97**, 88-97.
- Kumar M., Dwivedi P., Sharma A.K., Telang A.G., Patil R.D. and Singh N.D. (2008). Immunotoxicity of ochratoxin a and citri-

- inin in New Zealand White Rabbits. *World Rabbit Sci.* **16**, 7-12.
- Lakkawar A.W., Chattopadhyay S. and Tripurari S.J. (2004). Experimental aflatoxin B₁ toxicosis in young rabbits—a clinical and pathoanatomical study. *Slovenia Vet. Res.* **41**, 73-81.
- Makkar H.P.S. and Singh B. (2009). Aflatoxicosis in rabbits. *J. Appl. Rabbit Res.* **14**, 218-221.
- Manning B.B., Ulloa R.M., Li M.H., Robinson E.H. and Rottinghaus G.E. (2003). Ochratoxin A fed to channel catfish (*Ictalurus punctatus*) causes reduced growth and lesions of hepatopancreatic tissue. *Aquacult.* **219**, 739-750.
- Marin S., Ramos A.J., Cano-Sancho G. and Sanchis V. (2013). Mycotoxins: Occurrence, toxicology, and exposure assessment. *Food Chem. Toxicol.* **60**, 218-237.
- Marquardt R.R. and Frohlich A.A. (1992). A review of recent advances in understanding ochratoxicosis. *J. Anim. Sci.* **70**, 3968-3988.
- Massey T.E., Stewart R.K., Danniels J.M. and Liu L. (1995). Biochemical and molecular aspects of mammalian susceptibility to aflatoxin B₁ carcinogenicity. *Proc. Soc. Exp. Biol. Med.* **208**, 213-227.
- Mayura K., Reddy R.V., Hayes A.W. and Berndt W.O. (1982). Embryocidal, fetotoxic and teratogenic effects of ochratoxin A in rats. *Toxicology.* **25**, 175-185.
- McLean M. and Dutton M.F. (1995). Cellular interactions and metabolism of aflatoxin: an update. *Pharmacol. Ther.* **65**, 163-192.
- Mezes M. (2008). Mycotoxins and other contaminants in rabbit feed. Pp. 491-506 in Proc. 9th World Rabbit Congr., Verona, Italy.
- Mezes M. and Balogh K. (2009). Mycotoxins in Rabbit feed: A review. *World Rabbit Sci.* **17**, 53-62.
- Mir M.S. (1998). Studies on pathology and immunology of ochratoxin A induced toxicosis and its interaction with *Pasteurella multocida* infection in rats. MS Thesis. Veterinary Research Institute, Izatnagar, India.
- Mir M.S., Dwivedi P. and Charan K. (1999). Ochratoxin A induced acute toxicity in rabbits. *Indian J. Vet. Pathol.* **23**, 8-13.
- Mir M.S. and Dwivedi P. (2000). Clinical studies on experimental subacute ochratoxicosis in rabbits. *Indian J. Vet. Pathol.* **24**, 99-101.
- Mir M.S. and Dwivedi P. (2010). Immunopathology of ochratoxicosis-A in New Zealand White Rabbits (*Oryctolagus cuniculus*). *Vet. Scandinavica.* **5**, 1-10.
- Mizutani Y., Ito Y. and Ohtsubo K. (1997). Inhibition of platelet aggregation *in vitro* by trichothecene mycotoxins. *Mycotoxins.* **44**, 41-44.
- Morgavi D. and Riley R.T. (2007). An historical overview of field disease outbreaks known or suspected to be caused by consumption of feeds contaminated with *Fusarium* toxins. *Anim. Feed Sci. Technol.* **137**(3), 201-212.
- Moss M.O. (1996). Mycotoxins: Centenary review. *Mycol. Res.* **100**, 513-523.
- Moura M.A., Machado C.H., Porfírio L.C. and Freire R.B. (2004). Effects of ochratoxin A on broiler leukocytes. *Br. J. Poultry Sci.* **6**, 187-190.
- Murphy P.A., Hendrich S., Landgren C. and Bryant C.M. (2006). Food mycotoxins: An update. *J. Food Sci.* **71**, 51-65.
- Niyo K.A., Richard J.L., Niyo Y. and Tiffany L. (1988). Pathologic, hematologic, and serologic changes in rabbits given T-2 mycotoxin orally and exposed to aerosols of *Aspergillus fumigatus* conidia. *Am. J. Vet. Res.* **49**, 2151-2160.
- Ostry V., Mair F. and Ruprich J. (2013). Producers and important dietary sources of ochratoxin A and citrinin. *Toxins.* **5**, 1574-1586.
- Pestka J.J. (2007). Deoxynivalenol: Toxicity, mechanisms and health risks. *Anim. Feed Sci. Technol.* **137**(3), 283-298.
- Pfohl-Leskowicz A. and Manderville R.A. (2007). Ochratoxin A: an overview on toxicity and carcinogenicity in animals and humans. *Mol. Nutr. Food Res.* **51**, 1189-1192.
- Placinta C.M., D'Mello J.P.F. and Macdonald A.M.C. (1999). A review of worldwide contamination of cereal grains and animal feed with *Fusarium* mycotoxins. *Anim. Feed Sci. Technol.* **78**, 21-37.
- Ponnuchamy V. (2000). Studies on pathology and immunology of ochratoxin-A induced toxicosis and its interaction with *Salmonella typhimurium* infection in guinea pigs. MS Thesis. Veterinary Research Institute, Izatnagar, India.
- Prabu P.C., Dwivedi P. and Sharma A.K. (2013). Toxicopathological studies on the effects of aflatoxin B₁, ochratoxin A and their interaction in New Zealand White Rabbits. *Exp. Toxicol. Pathol.* **65**, 277-286.
- Raina J.S. and Singh B. (1991). Prevalence and pathology of mycotoxicosis in poultry in Punjab. *Indian J. Animal Sci.* **61**, 671-676.
- Rati E.R., Shantha T. and Ramesh H.P. (1991). Effect of long term feeding and withdrawal of aflatoxin B₁ and ochratoxin A on kidney cell transformation in albino rats. *Indian J. Exp. Biol.* **29**, 813-817.
- Richard J.L., Thurston J.R., Deyoe B.L. and Booth G.D. (1975). Effect of ochratoxin and aflatoxin on serum proteins, complement activity and antibody production to *Brucella abortus* in guinea pigs. *Appl. Microbiol.* **29**, 27-29.
- Richard L.L., Bray G.A. and Ryan D.H. (1991). Mycotoxins as immunomodulatory in animal systems. Pp. 196-220 in Mycotoxins. Cancer and Health. G.A. Bray and D.H. Ryan, Eds. Louisiana State University Press, USA.
- Roberts B.A., Glancy E.M. and Patterson D.S.P. (1981). Rapid economical method for determination of aflatoxin and ochratoxin in animal feed stuffs. *J. Assoc. Off. Anal. Chem.* **64**(4), 961-963.
- Rotter B.A., Prelusky D.B. and Pestka J.J. (1996). Toxicology of deoxinivalenol (vomitoxin). *J. Toxicol. Environ. Health.* **48**, 1-34.
- Sahoo P.K., Chattopadhyay S.K., Johri T.S., Charan K. and Sikdar A. (1993). Pathology of experimental aflatoxicosis in rabbits. *Indian J. Anim. Sci.* **63**, 268-273.
- Saleemi M.K., Khan M.Z., Khan A., Hassan Z.U., Khan W.A., Rafique S., Fatima Z. and Sultan A. (2015). Embryotoxic and histopathological investigation of *in ovo* inoculation of aflatoxicogenic fungal extracts in chicken embryos. *Pakistan Vet. J.* **35**, 403-408.
- Salem M.H., Kamel K.I., Yousef M.I., Hassan G.A. and El-Nouty F.D. (2001). Protective role of ascorbic acid to enhance semen

- quality in rabbits treated with sublethal doses of aflatoxin B₁. *Toxicology*. **162**, 209-218.
- Schatzmayr G. and Streit E. (2013). Global occurrence of mycotoxins in the food and feed chain: facts and figures. *World Mycotoxin J.* **6**, 213-222.
- Sekine T., Miyazaki H. and Endou H. (2006). Molecular physiology of renal organic anion transporters. *Am. J. Physiol.* **290**, 251-261.
- Sharma P. (1998). Himachal Pradesh rabbit farmers fear toxin in feed: mortality, infertility cases on rise. *The Hindustan Times*. **210**, 1-3.
- Singh K.P., Singh R., Singh Y.P., Telang A.G. and Mehrotra M.L. (1999). Investigation of mortality due to aflatoxicosis in New Zealand White Rabbits. *Indian J. Vet. Pathol.* **23**, 83-84.
- Stein A.F., Phillips T.D., Kubena L.F. and Harvey R.B. (1985). Renal tubular secretion and reabsorption as factors in ochratoxicosis: effects of probenecid on nephrotoxicity. *J. Toxicol. Environ. Health.* **16**, 593-604.
- Stetinova V., Grossmann V. and Kvetina J. (1998). Changes in the gastrointestinal tract, cardiovascular function and some drug metabolizing processes in rats and guinea pigs intoxicated with aflatoxin B₁. *Polish J. Pharmacol.* **50**, 135-141.
- Still P.E., Macklin A.W., Ribelin W.E. and Smalley E.B. (1971). Relationship of ochratoxin A to fetal death in laboratory and domestic animals. *Nature*. **31**, 563-564.
- Streit E., Schatzmayr G., Tassis P., Tzika E., Marin D., Taranu I., Tabuc C., Nicolau A., Aprodu I., Puel O. and Oswald I.P. (2012). Current situation of mycotoxin contamination and co-occurrence in animal feed-focus on Europe. *Toxins*. **4**, 788-809.
- Stresser D.M., Bailey G.S. and Williams D.E. (1994). Indole-3-carbinol and ̂-naphthoflavone induction of aflatoxin B₁ metabolism and cytochrome P450 associated with bioactivation and detoxication of aflatoxin B₁ in the rats. *Drug Metab. Dispos.* **22**, 383-391.
- Sundaram T.K., Chandrasekaran D., Natrajan A. and Mohan B. (1999). Multimycotoxins: An overview of their occurrence in feed ingredients and feeds in Namakkal. Pp. 23-37 in 18th Ann. Conf. Soc. Toxicol., Lucknow, India.
- Suzuki S., Kozuka Y., Satoh T. and Yamazaki M. (1975). Studies on the nephrotoxicity of ochratoxin A in rats. *Toxicol. Appl. Pharmacol.* **34**, 479-490.
- Sweeney M.J. and Dobson A.D.W. (1998). Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species. *Int. J. Food Microbiol.* **43**, 141-158.
- Szilágyi M., Fekete S., Huszenicza Gy. and Albert M. (1994). Biochemical and physiological effects of long-term sublethal T-2 toxin feeding in rabbits. *Acta Biol. Hung.* **45**, 69-76.
- Thacker H.L. and Carlton W.W. (1977). Ochratoxin A mycotoxicosis in the guinea pig. *Food Cosmet. Toxicol.* **15**, 563-574.
- Thrasher J.D. (2012). Aflatoxicosis in animals. Aflatoxins and Health. Available at: www.alphaboostjuice.com/aflatoxicosis_in_animals.pdf.
- USAID (2012). Aflatoxin: A Synthesis of the Research in Health, Agriculture and Trade. Feed the Future: The Office of Regional Economic Integration USAID East Africa Regional Mission Nairobi, Kenya. Available at: www.eastafrica.usaid.gov/research_in_Health_Agriculture_and_Trade/pdf_10-15.
- Valtchev I., Koynarski T., Sotirov L., Nikolov Y. and Petkov P. (2015). Effect of aflatoxin B1 on moult duck's natural immunity. *Pakistan Vet. J.* **35**, 67-70.
- Verma R.J. and Mehta D.N. (1998). Occurrence of hemolytic anemia during aflatoxicosis. *Indian J. Environ. Toxicol.* **8**, 5-7.
- Vinita R., Prasad L.N. and Sinha B.K. (2003). Pathological and histochemical changes in liver and kidneys in experimental aflatoxicosis in rabbits. *Indian J. Vet. Pathol.* **27**, 57-65.
- Voss K.A., Smith G.W. and Haschek W.M. (2007). Fumonisin: Toxicokinetics, mechanism of action and toxicity. *Anim. Feed Sci. Technol.* **137**, 299-325.
- Wangikar P.B., Dwivedi P. and Sinha N. (2004b). Effect in rat of simultaneous prenatal exposure to ochratoxin A and aflatoxin B₁ I Maternal toxicity and fetal malformations. *Birth Defects Res. Part B: Dev. Reprod. Toxicol.* **71**, 343-351.
- Wangikar P.B., Dwivedi P., Sharma A.K. and Sinha N. (2004c). Effect in rat of simultaneous prenatal exposure to ochratoxin A and aflatoxin B1. II. Histopathological features of teratological anomalies induced in fetuses. *Birth Defects Res. Part B: Dev. Reprod. Toxicol.* **71**, 352-358.
- Wangikar P.B., Dwivedi P., Sinhab N., Sharma A.K. and Telanga A.G. (2005). Teratogenic effects in rabbits of simultaneous exposure to ochratoxin A and aflatoxin B₁ with special reference to microscopic effects. *Toxicology*. **215**, 37-47.
- WHO-World Health Organization International Agency for Research on Cancer. (1993). Toxins derived from *Fusarium moniliforme*: Fumonisin B₁ and B₂ and fusarin C. World Health Organisation. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK513589>.
- WHO-World Health Organization International Agency for Research on Cancer. (2000). Hazardous chemicals in humans and environmental health: International programme on chemical safety, Geneva, Switzerland. World Health Organisation. Available at: <https://apps.who.int/iris/handle/10665/66161>.
- Wu F. (2004). Mycotoxins risk assessment for the purpose of setting international regulatory standards. *Environ. Sci. Technol.* **38**, 4049-4055.
- Wu F. (2006). Economic impact of fumonisin and aflatoxin regulations on global corn and peanut markets. Pp. 277-289 in The Mycotoxin Factbook. Food and Feed Topics. D. Barug, D. Bhatnager, H.P. van Egmond, J.W. van der Kamp, W.A. van Osenbruggen and A. Visconti, Eds. Wageningen Academic Publishers, Wageningen, The Netherlands.
- Zaki M.M., El-Midany S.A., Shaheen H. M and Rizzo L. (2012). Mycotoxins in animals: Occurrence, effects, prevention and management. *J. Toxicol. Environ. Health Sci.* **4**, 13-28.
- Zofair S.M., Mathew S. and Verma R.J. (1996). Ochratoxin induced hemolysis in rabbits. *Indian. J. Exp. Biol.* **34**, 592-593.