



ABSTRACT

The objective of this study was to determine the effect of irradiation of pomegranate seed (PS) on chemical composition, digestibility and kinetic of gas production. Pomegranate seeds were exposed to gamma ray (GR) and electron beam (EB) at doses of 5, 10, 15 and 20 kGy. Three ruminally fistulated rams were used for obtaining ruminal fluid for *in vitro* digestibility and gas production measurements. Irradiation had no effect on chemical composition of PS. Orthogonal contrast did not show any significant effect for EB irradiation on neutral detergent fiber % (NDF%), but GR irradiation of PS at low doses (i.e.; 5 and 15 kGy) increased NDF percentage (P<0.05). Irradiation decreased condensed tannin (CT) content of PS at all doses (P<0.01). Gas production potential (*b*) and gas production potential of PS. Irradiation treatment did not affect partitioning factor. Ionizing radiation decreased PS digestibility, but EB irradiation at a dose of 20 kGy and 5, 15 and 20 kGy did not change *in vitro* dry matter digestibility and organic matter digestibility. In conclusion, the result of this study suggests that ionizing radiation processing, especially EB irradiation, can be regarded as an efficient method in decreasing CT of pomegranate seeds.

KEY WORDS condensed tannin, digestibility, irradiation, pomegranate seed.

INTRODUCTION

Pomegranate (*Punica granatum*) is an important fruit crop in tropical and subtropical regions of the world as well as in Mediterranean countries with moderate temperatures (Taher-Maddah *et al.* 2012). The fruits are globally consumed fresh, in processed forms as juice, jam and oil and in extract supplements (Prakash and Prakash, 2011). This has led to development of advanced industrial technologies which provide consumers with "ready to eat" pomegranate grains and fresh fruit juices. These products have led to production of high quantities of pomegranate byproduct biomass. Pomegranate seeds constitute about 3% of the weight of the fresh fruit. Feizi *et al.* (2005) demonstrated that pomegranate seeds can be used in animal nutrition. Although protein and oil content of pomegranate seed are considerable (Mirzaei-Aghsaghali *et al.* 2011), it has a high amount of cell wall content and is rich in antinutritional factors, particularly tannins (Parakash and Parakash, 2011). Removal of these undesirable components is essential to improve the nutritional quality of pomegranate seeds as animal feed. There is scant information on the nutritive value of pomegranate seeds for ruminants (Feizi *et al.* 2005; Shabtay *et al.* 2008; Modarresi *et al.* 2010; Mirzaei-Aghsaghali *et al.* 2011). The previous methods introduced by researchers (Chen *et al.* 1995; Duodu *et al.* 1999; Parker *et al.* 1999) to deactivate the antinutrients compounds in feeds did not necessarily completely eliminate or reduce the

amounts of them; instead in some cases the methods gave the adverse results and led to the reduced nutritive value of the feeds.

There is no information on the effect of gamma ray (GR) and electron beam (EB) irradiation on the nutritive value of pomegranate seeds. Therefore, the major aim of the present study was to evaluate the impacts of gamma and electron irradiation on the nutritional and antinutritional components, *in vitro* digestibility and rumen fermentation characteristics of pomegranate seeds.

MATERIALS AND METHODS

Samples preparation and irradiation treatments

Pomegranate seeds (PS) were obtained from the Neyriz Green Farm pomegranate factory, in Fars, Iran, during the pomegranate harvest season and were air dried before it was used in this study.

The experimental treatments were: pomegranate seeds treated by gamma ray (GR) and electron beam (EB) irradiation at doses of 5, 10, 15 and 20 kGy and the control group without irradiation. Irradiations of samples were done in Radiation Applications Research School, Nuclear Science and Technology Research Institute, Atomic Energy Organization of Iran.

Gamma-irradiation was completed by using a cobalt-60 irradiator at 20 °C. The dose rate determined by Fricke dosimetry (Holm and Berry, 1970) was 0.36 Gy/s. Threepaper packages of samples were irradiated to total doses of 15, 30 and 45 kGy in the presence of air. After irradiation and prior to sealing the plastic bags, samples were allowed to air equilibrate for 2 h. The pomegranate seed samples were irradiated under various doses of 5, 10, 15 and 20 kGy. Three poly-ethylene packages of samples were exposed to 10 MeV electron beam of a Rhodotron accelerator model TT-200 (IBA Co., Belgium) at various doses (5, 10, 15 and 20 kGy) in Radiation Applications Research School of Atomic Energy Organization of Iran. All irradiations were performed at room temperature in air, with 4 mA beam of 10 MeV electrons. Regarding the low thickness of the samples packages, single sided irradiation has been used. The required doses were delivered to the samples by adjusting the conveyer speed when each of the sample batches passed under the beam.

Chemical analysis

The dry matter (DM) of pomegranate seed was determined by drying at 60 $^{\circ}$ C for 48 h. After drying, the samples were ground through a 1 mm screen (Wiley mill, Arthur H. Thomas, Philadelphia, USA), and DM, crud protein (CP) (948.13), ether extract (EE) (954.02) and ash (924.05) were analyzed according to AOAC (1995). Neutral detergent fiber (NDF), acid detergent fiber (ADF) were analyzed according to the method of Van Soest *et al.* (1991). Condensed tannins were analyzed using the vanillin-HCl procedure according to Galyean (1997) and results are expressed as catechin equivalents (mg of catechin equivalents/g of dry sample).

In vitro study

In vitro digestibility

Two-step digestion technique (Tilly and Terry, 1963) was used to determine *in vitro* digestibility of un-irradiated and irradiated PS. Allocation of treatments into *in vitro* experimental units was done as a completely randomized design. Samples (1 g) were weighed and placed into tubes. Subsequently, 12 mL of McDougall's buffer and 8 mL of rumen fluid were added. The tube was closed with a rubber cap and incubated anaerobically for 24 h in an automatic shaker water bath, maintained at 39 °C during the process. After 24 h, the cap was opened, 0.2 mL of HgCl₂ was added, centrifuged at 10000 rpm for 10 min and the supernatant was removed. The residue was combined with 20 mL of 0.2% pepsin under acidic condition, and further incubated for 24 h.

The remaining sample after the two-stage *in vitro* incubation procedure was filtered with a Whatman paper no. 41 for determination of *in vitro* dry matter digestibility (IVDMD) and *in vitro* organic matter digestibility (IVOMD).

Values of IVDMD and IVOMD were presented as percentage (%) of digested substance from their initial amounts prior to incubation. Blanks (rumen fluid and buffer only without sample substrate) were incubated as described above and served as a correction factor to the DM and organic matter (OM) contents of residuals. The incubation was done in three replicates according to the treatments (n=3) and each replicate was represented by two tubes.

In vitro gas production

The method used for gas production measurements was as described by Theodorou *et al.* (1994). Three ruminally fistulated Sanjabi rams, as rumen fluid donors, were fed at 8:15 and 17:15 h daily with a diet of lucerne hay and whole barley (70:30, DM basis) at the maintenance requirements. Rumen fluid was collected at the morning before feeding and strained into a pre-warmed thermos flask. Pomegranate seeds were ground to pass a 0.2 mm screen. Approximately 125 mg of each substrate was weighed into 25 mL serum bottles and incubated in a water bath at 39 °C with 5 mL strained rumen fluid and 10 mL of medium (McDougall, 1948) in order to determine rate and extent of gas production by reading gas production at 2, 4, 6, 8, 10, 12, 24, 48, 72 and 96 h post-inoculation.

Each sample was incubated in three replicates. Cumulative gas production data were fitted to the model of Orskov and McDonald (1979) as follows:

 $Y = b(1 - e^{-ct})$

Where:

b: potential extent of gas production.

c: gas production rate constant.

t: incubation time.

y: gas produced at time "t".

In a separate run of gas production, the method of Blummel *et al.* (1997) was adopted to determine the partitioning factor.

Statistical analyses

Experimental data was analyzed using general linear model (GLM) and significant differences among means from a triplicate analysis at (P<0.05) were determined by LSD test using the SAS (2002) software. Orthogonal contrasts were used to detect significant differences among the treatment means.

RESULTS AND DISCUSSION

Effects of irradiation on chemical composition

Orthogonal contrast of chemical composition of PS before and after irradiation are shown in Table 1. Irradiation had no effects on DM, EE, CP, OM, ADF and nonfiber carbohydrates (NFC) percentages of pomegranate seeds (P>0.05). Orthogonal contrast indicated that the GR irradiation at low doses (i.e.; 5 and 15 kGy) significantly increased the NDF content (as %) of pomegranate seeds compared to the control group. The effect of EB irradiation on the NDF content of pomegranate seeds was no significant. Condensed tannin content of PS was significantly reduced by irradiation compared to the control. Significant differences were observed between GR and EB irradiation effects on condensed tannin reduction (P<0.05).

Chemical composition of untreated and irradiated pomegranate seed are shown in Table 2. Gamma ray and EB irradiation at a dose of 10 kGy significantly increased NDF content of PS but the effect of electron beam irradiation was not significant. Irradiations significantly decreased condensed tannin. Reduction in condensed tannin in response to irradiations was dose dependent (Table 2).

In vitro study

Gas production, kinetic analysis of gas production

Cumulative gas production of irradiation treatments and the estimated parameters of gas production are presented in

Figure 1 and in Tables 3 and 4, respectively. In total, irradiation decreased the gas production parameters.

As can be seen from Table 4, rate constant of gas production (c) was decreased but gas production potential of PS did not significantly affected by EB irradiation at doses of 5 and 20 kGy.

In vitro digestibility

In vitro digestibilities of dry matter and organic matter of untreated and irradiated PS are presented in Tables 3 and 4. *In vitro* dry matter digestibility and *in vitro* organic matter digestibility of pomegranate seeds decreased by GR and EB irradiation with the exception of 5 and 20 kGy EB irradiation.

In spite of non significant effect of irradiations on the partitioning factor, there was a difference in GV_{24} between untreated and irradiated PS (Table 6). Mean average of partitioning factor was 5.62 mg DM truly degraded/mL gas produced in 24 h. Partitioning factor ranged from 4.61 to 6.90. Orthogonal contrast showed that irradiation affected metabolizable energy, but there was no difference between gamma and electron radiation (Table 5).

The correlation coefficients of chemical composition and experimental parameters of gamma and electron irradiated PS are presented in Tables 7 and 8.

Effects on chemical composition

DM, EE, CP, OM, ADF and NFC content of irradiated pomegranate seeds was not significantly different from the control, which are in agreement with previous works (Shawrang et al. 2011; Bhat and Sridhar, 2008; Ebrahimi et al. 2009; Taghinejad et al. 2009; Farag, 1998). Although gamma irradiation was effective and led to a significant increase in NDF content of PS compared to the control (Table 2), the effect for electron beam irradiation was not significant. In common, irradiation treatment showed to be effective on the structure of cellulosic raw materials (Dela Rosa et al. 1983). However, there are some discrepancies regarding the effects of irradiation on fiber content of feeds. Electron beam irradiation in high doses (50, 100 and 150 kGy) decreased NDF and ADF content of soybean and cotton seed meal in Tahan et al. (2012) study, but Ebrahimi-Mahmoudabad and Taghinejad-Roudbaneh (2011) did not indicate any significant effect of EB irradiation (at doses of 15, 30 and 45 kGy) on NDF content of whole cottonseed, soybean and canola seeds. Ebrahimi et al. (2009) also did not find any significant effect for GR irradiation on chemical composition and fiber content of feeds. In our study EB irradiation at a dose of 10 kGy significantly increased NDF content of PS. Similarly, Khosravi et al. (2012) reported that EB irradiation significantly increased ADF content of pomegranate seeds without any significant effect on NDF.

Table 1 Orthogonal contrast of pomegranate seeds before and after irradiation (means square)

Treatments	df	OM	СР	EE	NDF	ADF	NFC	CT
Irradiation vs. control	1	0.001	0.14	0.04	101.28	35.14	148.08	10.44^{**}
GR vs. control	1	0.001	0.09	0.01	200.82^*	55.47	234.35	8.88^{**}
5 and 10 GR vs. control	1	0.04	0.01	0.003	166.20	21.38	177.84	3.66**
15 and 20 GR vs. control	1	0.01	0.18	0.01	158.99	74.29	235.96	12.44**
5 and 10 vs. 15 and 20 GR	1	0.17	0.13	0.01	0.63	26.34	15.40	3.90**
EB vs. control	1	0.01	0.15	0.08	22.43	13.39	65.99	9.93**
5 and 10 EB vs. control	1	0.01	0.09	0.05	16.37	26.93	44.39	4.07^{**}
15 and 20 EB vs. control	1	0.002	0.15	0.09	18.93	1.77	66.73	13.95**
5 and 10 vs. 15 and 20 EB	1	0.01	0.01	0.01	0.10	19.92	3.40	4.42^{**}
GR vs. EB	1	0.04	0.01	0.10	165.93	22.91	134.02	0.07^{*}
5 and 10 GR vs. 5 and 10 EB	1	0.17	0.04	0.05	87.14	1.87	60.75	0.01
15 and 20 GR vs. 15 and 20 EB	1	0.01	0.00	0.04	79.56	66.22	73.28	0.06

GR: gamma ray; EB: electron beam; df: degree of freedom; OM: organic matter; CP: crude protein; EE: ether extract; NDF: neutral detergent fiber; ADF: acid detergent fiber; NFC: non fibrous carbohydrates and CT: condense tannin (mg of CE/g of dry sample). * (P<0.05) and ** (P<0.01).

Table 2 Chemical compositions of irradiated pomegranate seeds (as g/100 g dry matter)

Treatments	OM	СР	EE	NDF	ADF	NFC	CT
Control	97.61	11.12	10.92	52.72 ^{bc}	39.18	28.29 ^{ab}	4.10^{a}
Gamma							
5 kGy	97.76	11.19	10.97	59.48 ^{ab}	45.50	17.24 ^{ab}	3.03 ^b
10 kGy	97.74	10.81	10.96	68.30 ^a	40.87	9.53 ^{ab}	1.85 ^c
15 kGy	97.42	10.66	11.05	65.95 ^{ab}	46.56	9.96 ^{ab}	1.41 ^{de}
20 kGy	97.61	10.83	11.03	63.08 ^{ab}	47.93	10.60^{ab}	$0.69^{\rm f}$
Electron							
5 kGy	97.44	11.01	11.06	42.71 [°]	40.41	34.78 ^a	3.12 ^b
10 kGy	97.58	10.67	11.16	70.30 ^a	48.33	5.48 ^b	1.59 ^d
15 kGy	97.55	10.53	11.17	57.28 ^{abc}	42.13	18.77 ^{ab}	1.21 ^e
20 kGy	97.60	10.97	11.19	56.30 ^{abc}	38.26	17.80 ^{ab}	0.52^{f}
LSD	0.79	1.02	1.13	15.36	12.67	27.32	0.26
P-value	0.9892	0.7475	0.9997	0.0438	0.3887	0.2660	0.0001
SEM	0.07	0.08	0.09	2.40	1.20	2.67	0.27

GR: gamma ray; EB: electron beam; df: degree of freedom; OM: organic matter; CP: crude protein; EE: ether extract; NDF: neutral detergent fiber; ADF: acid detergent fiber; NFC: non fibrous carbohydrates and CT: condense tannin (mg of CE/g of dry sample). The means within the same column with at least one common letter, do not have significant difference (P>0.05).

SEM: standard error of the means.

LSD: least significant difference.



Figure 1 Cumulative gas production curve of gamma and electron beam irradiated pomegranate seed

Table 3 Orthogonal contrast of in vitro gas production parameters and digestibility of GR and El	B irradiated pomegranate seeds (means square)
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Transformente	16	Param	ieters		OMD(0/)	
Treatments	di	b	с	DMD (%)	OMD (%)	
Irradiation vs. control	1	1106.90**	0.0016**	107.25**	117.58**	
GR vs. control	1	1275.03**	0.0022^{**}	170.45**	165.79**	
5 and 10 GR vs. control	1	1114.07**	0.0018^{**}	171.49**	153.36**	
15 and 20 GR vs. control	1	1012.20**	0.0020^{**}	115.36**	111.64**	
5 and 10 vs. 15 and 20 GR	1	3.66 ^{ns}	0.00^{ns}	8.31 ^{ns}	6.17 ^{ns}	
EB vs. control	1	751.75**	0.0008^{**}	43.48**	55.31*	
5 and 10 EB vs. control	1	812.17**	0.0008^{**}	114.71**	107.06**	
15 and 20 EB vs. control	1	464.82**	0.0005^{**}	1.76 ^{ns}	10.44 ^{ns}	
5 and 10 vs. 15 and 20 EB	1	72.22 ^{ns}	0.00^{*}	132.0**	75.95**	
GR vs. EB	1	171.78 ^{ns}	0.0009^{**}	104.37**	73.60**	
5 and 10 GR vs. 5 and 10 EB	1	35.70 ^{ns}	0.0003**	8.53 ^{ns}	11.0 ^{ns}	
15 and 20 GR vs. 15 and 20 EB	1	157.76 ^{ns}	0.0006^{**}	132.86**	80.42**	

GR: gamma ray; EB: electron beam; df: degree of freedom; b: gas production potential (mL/g DM); c: rate constant of gas production (mL/h); DMD: dry matter digestibility and OMD: organic matter digestibility. (P<0.05) and $^{**}(P<0.01)$. NS: non significant.

Table 4 In vitro gas production parameters and digestibility of pomegranate seeds before and after irradiation

Tractments	Param	eters		
Treatments	b	с	DMD (%)	OMD (%)
Control	185.73 ^a	0.09 ^a	47.41 ^a	47.38 ^a
Gamma				
5 kGy	162.93 ^b	0.06^{d}	39.30 ^{de}	38.54 ^{bc}
10 kGy	161.33 ^b	0.06^{d}	37.01 ^e	37.31 [°]
15 kGy	162.35 ^b	0.05^{d}	38.65 ^{de}	38.19 ^{bc}
20 kGy	164.12 ^b	0.06^{d}	40.99^{cd}	41.02 ^{bc}
Electron				
5 kGy	172.73 ^{ab}	0.08^{b}	42.50 ^{bc}	42.75 ^{ab}
10 kGy	158.43 ^b	0.06^{d}	37.18 ^e	37.38 ^c
15 kGy	165.66 ^b	0.07°	44.49^{b}	42.98 ^{ab}
20 kGy	175.31 ^{ab}	0.07^{b}	48.45^{a}	47.21 ^a
LSD	17.40	0.006	2.86	4.83
P-value	0.006	0.0001	0.0001	0.0011
SEM	1.99	0.002	0.83	0.88

b: gas production potential (mL/g DM); c: rate constant of gas production (mL/h); DMD: dry matter digestibility and OMD: organic matter digestibility.

The means within the same column with at least one common letter, do not have significant difference (P>0.05).

SEM: standard error of the means.

LSD: least significant difference.

Table 5 Orthogonal contrast of PF, GV₂₄ and ME of pomegranate seeds (means square)

Treatments	PF	GV_{24}	ME
Irradiation vs. control	0.22	2337.87**	1.16^{*}
GR vs. control	0.05	2965.95**	1.23^{*}
5 and 10 GR vs. control	0.32	2521.92**	1.48^{*}
15 and 20 GR vs. control	1.0	2421.84**	0.63^{*}
5 and 10 vs. 15 and 20 GR	3.72	1.51	0.15
EB vs. control	0.44	1389.80**	0.80^{*}
5 and 10 EB vs. control	0.002	1525.36**	0.73^{*}
15 and 20 EB vs. control	1.35	841.45**	0.50
5 and 10 vs. 15 and 20 EB	1.84	151.44^{*}	0.04
GR vs. EB	0.46	737.92**	0.05
5 and 10 GR vs. 5 and 10 EB	0.58	186.91*	0.05
15 and 20 GR vs. 15 and 20 EB	0.03	612.32**	0.009

GR: gamma ray; EB: electron beam; PF: partitioning factor (mg DM/mL gas); GV_{24} : gas volume at 24 hour (mL) and ME: metabolizable energy (MJ/kg DM). *(P<0.05) and **(P<0.01).

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Treatments	PF	GV 24	ME
Control	5.36 ^{ab}	150.38ª	7.21
Gamma			
5 kGy	5.29^{ab}	114.77 ^c	6.20
10 kGy	4.61 ^b	114.97 ^c	6.10
15 kGy	6.90 ^a	114.27 ^c	6.22
20 kGy	5.23 ^{ab}	116.90 ^c	6.70
Electron			
5 kGy	4.72 ^b	133.83 ^b	6.71
10 kGy	6.07^{ab}	111.70 ^d	5.98
15 kGy	5.82 ^{ab}	124.86 ^{bc}	6.38
20 kGy	6.54^{ab}	134.88 ^b	6.70
LSD	2.12	13.67	0.95
P-value	0.3536	0.0001	0.1458
SEM	0.24	2.59	0.11

PF: partitioning factor (mg DM/mL gas); GV₂₄: gas volume at 24 hour (mL) and ME: metabolizable energy (MJ/kg DM).

The means within the same column with at least one common letter, do not have significant difference (P>0.05).

SEM: standard error of the means.

LSD: least significant difference.

 Table 7
 Correlation coefficient (r) of the relationship between chemical composition and experimental parameters of gamma irradiated pomegranate seeds

	DMD	OMD	b	С	GP ₉₆	GP ₂₄	ME	PF
DMD	1	0.93*	0.88^*	0.87^{*}	0.38	0.88^{*}	0.93^{*}	-0.16
OMD	0.93^{*}	1	0.86^{*}	0.92^*	0.41	0.91^{*}	0.92^{*}	-0.15
CT	0.47	0.54	0.69^*	0.73^{*}	-0.01	0.73^{*}	0.25	0.07
NDF	-0.55	-0.61	-0.73*	-0.66	-0.40	-0.71*	-0.23	-0.43
ADF	-0.31	-0.29	-0.59	-0.56	-0.31	-0.59	-0.20	0.48
NFC	0.37	0.61	0.65	0.71^{*}	0.67	0.70	-0.38	0.39

DMD: dry matter digestibility; OMD: organic matter digestibility; *b*: gas production potential (mL/g DM); *c*: rate constant of gas production (mL/h); GP_{96} : gas production at 96 hour; GP_{24} : gas production at 24 hour (mL); ME: metabolizable energy (MJ/kg DM); PF: partitioning factor (mg DM/mL gas); CT: condense tannin (mg of CE/g of dry sample); NDF: neutral detergent fiber; ADF: acid detergent fiber and NFC: non fibrous carbohydrates and. *(P<0.05).

 Table 8
 Correlation coefficient (r) of the relationship between chemical composition and experimental parameters of electron irradiated pomegranate seeds

	DMD	OMD	b	с	GP ₉₆	GP ₂₄	ME	PF
DMD	1	0.88^{*}	0.68^{*}	0.75^{*}	0.37	0.77^{*}	0.93*	-0.16
OMD	0.88^{*}	1	0.77^{*}	0.71^{*}	0.46	0.82^{*}	0.92^{*}	-0.15
CT	-0.003	-0.001	0.54	0.64^{*}	0.05	0.61	0.50	-0.61
NDF	0.10	0.46	0.40	-0.49	-0.52	-0.56	-0.92	-0.81
ADF	-0.80^{*}	-0.85*	-0.71	-0.72	-0.66	-0.73	-0.67	0.48
NFC	0.48	0.24	0.69	0.85	-0.36	0.76	0.56	-0.27

DMD: dry matter digestibility; OMD: organic matter digestibility; *b*: gas production potential (mL/g DM); *c*: rate constant of gas production (mL/h); GP₉₆: gas production at 96 hour; GP₂₄: gas production at 24 hour (mL); ME: metabolizable energy (MJ/kg DM); PF: partitioning factor (mg DM/mL gas); CT: condense tannin (mg of CE/g of dry sample); NDF: neutral detergent fiber; ADF: acid detergent fiber and NFC: non fibrous carbohydrates and. * (P<0.05).

These contradictions could be due to differences in doses and types of irradiation, laboratory circumstance in cell wall analysis (Tahan *et al.* 2012), free radicals formation, depolymerization (chain-scission) or cross-linking of cellulose and glucose chain (Polvi and Nordlund, 2014; Khan *et al.* 2006; Pekel *et al.* 2004).

The efficiency of these types of reactions depends mainly on the polymer structure and radiation dose (Charlesby, 1981).

In total, our result did not indicate any beneficial effect of irradiation at low doses on chemical composition of pomegranate seeds.

Effects on tannin content

Despite some differences which exit between GR and EB irradiations for their effects on condensed tannin content of PS, these two techniques significantly (P<0.05) decreased the condensed tannin content of PS compared to control (Table 2).

The decrease in condensed tannin was positively dependent on irradiation dose. Reduction of tannin by irradiation in this study was consistent with some previous studies on gamma irradiation (El-Niely, 2007; Behgar *et al.* 2011; De Toledo *et al.* 2007) and electron irradiation (Bhat and Sridhar, 2008; Shawrang *et al.* 2011). Irradiation, generally, resulted in the degradation of tannin (Variyar *et al.* 1998) and a change in its molecular conformation (Topuz and Ozdemir, 2004). Mechanism of gamma action on tannin has been related to generation of the hydroxyl and superoxide anion radicals (Riley, 1994) which indiscriminately attacks neighbouring molecules, but mode of electron beam action on tannins has not been demonstrated.

In vitro study

Gas production profiles

Gas production parameters decreased significantly by irradiation (Table 4). Regarding to the potential gas production, there was no significant difference between gamma and electron radiation techniques.

The rate constant (fraction c) of gas production of EB irradiated PS at doses of 5 and 20 kGy decreased but the gas production potential did not affected significantly. The differences between gas parameters of lower (5 and 10 kGy) and upper (15 and 20 kGy) doses of GR and EB were not significant.

Ndlovu and Nherera (1997) reported that rate of gas production was negatively related to ADF, ADL and NDF content, a finding that agrees with our results. A negative correlation between condensed tannins and cell wall content with gas production parameters was reported by several studies (Kamalak *et al.* 2004; Ndlovu and Nherera, 1997; Nsahlai *et al.* 1994).

Focusing on the results presented in Tables 7 and 8, condensed tannins of PS showed a high positive correlation with gas parameters both in GR and EB irradiation which could be due to the tanning degradation and free phenolic compounds production that are toxic and suppressed the growth of the cellulolytic microorganism in the rumen (Chesson *et al.* 1982).

Behgar *et al.* (2011) reported that gamma radiation (at 30, 40 and 60 kGy dose) caused a decrease (P<0.05) in potential gas production of pistachio hull compared to the control.

Reductions in gas volume, fraction *b* and rate of gas production were also reported when tannic acid was added to the sunflower meal (Mohmmadabadi *et al.* 2010). In contrast, the rate of gas production in the study of Kamalak *et al.* (2004) was not related neither to chemical composition nor condensed tannins. A weak relationship between condensed tannins and gas production of tree leaves during wet and dry season in west Africa was reported by Larbi *et al.* (1998). A possible reason for these disparities could be due to differences in the nature of tannins between browse species (Jackson *et al.* 1996).

In vitro digestibility

In vitro dry matter digestibility and organic matter digestibility of PS decreased with the exception of EB at the dose of 20 kGy and 5, 15 and 20 KGy repectively. In a previous study by Ghanbari *et al.* (2012), the differences between various cottonseed meal treated by irritation (25, 50 and 75 kGy gamma and electron radiation) were not significant statistically.

Shawrang *et al.* (2011) reported that doses higher than 15 kGy of EB irradiation significantly increased dry matter digestibility of sorghum grains compared to the control. It is well documented that tannins and cell wall content of feedstuffs negatively affect their digestibility (Ndagurwa and Dube 2013; Guimaraes-Beelen *et al.* 2006; Mohmmadabadi *et al.* 2010).

These two components also influencing the growth and morphology of rumen microorganisms (O'Donovan and Brooker, 2001).

According to the results presented in Tables 7 and 8, ADF content of EB irradiated PS had significantly negative correlation with PS digestibility. There was no significant correlation between condensed tannin and digestibility of EB irradiated PS. Cell wall contents may be more important than tannins in limiting *in vitro* fermentation (Ndlovu and Nherera, 1997).

The effects of ionizing radiation on nutrients digestibility vary with irradiation dose and chemical structure and concentration of cell wall contents and tannins (Jackson *et al.* 1996).

The later may be due to differences in the nature of tannins between browse species and degradation of tannin by irradiation and adverse effect of free phenolics on rumen digestion (Chesson *et al.* 1982).

Irradiation did not change partitioning factor and gas volume at 24 h incubation (GV24) significantly decreased. Mean average of partitioning factor (PF) was 5.62 (mg DM truly degraded/mL gas produced in 24 h). Partitioning factor ranged from 4.61 to 6.90.

The theoretical range for partitioning factor of tannin-free plants suggested by Blummel *et al.* (1997) was between 2.75 and 4.41 (mg truly degraded substrate/mL gas). PF values greater than 4.41 are not theoretically possible (Makkar *et al.* 1998) and if this occurred, could simply indicate the inhibition of gas production due to the existence of tannins.

This result agreed with Makkar (2004). Irradiation changed ME of pomegranate seeds, but there is no difference among gamma and electron radiation. The mean of metabolizable energy of pomegranate seed was 6.44 MJ/kg DM, that was higher than Mirzaei-Aghsaghali *et al.* (2011)

and Taher-Maddah *et al.* (2012) who reported that estimated amounts of ME of dried pomegranate seed was 6.20 and 5.10 MJ/kg DM, respectively.

CONCLUSION

Generally irradiation at low doses could not change in chemical composition of pomegranate seed. Irradiation have the potential to reduce anti-nutritional factors. Condensed tannin content of PS was significantly decrease compared to control. There was a difference between the effects of gamma and electron radiation on reduction of condensed tannin. Electron beam radiation with a higher mean square of difference had more impact on reduction of condensed tannin as compared to gamma ray. Ionizing radiation processing can be used as an efficient method in decreasing condensed tannin content of PS, but it should be noted that potential feed value of PS could be altered by irradiation. So, further studies are needed to evaluate the definite effect of gamma and electron radiation on nutritional value and ruminal metabolism of feedstuff contained anti-nutritional factor.

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REFERENCES

- AOAC. (1995). Official Methods of Analysis. 16th Ed. Association of Official Analytical Chemists, Arlington, VA, USA.
- Behgar M., Ghasemi S., Naserian A., Borzoie A. and Fatollahi H. (2011). Gamma radiation effects on phenolics, antioxidants activity and *in vitro* digestion of pistachio (*Pistachia vera*) hull. *Radiat. Phys. Chem.* **80**, 963-967.
- Bhat R. and Sridhar K.R. (2008). Nutritional quality evaluation of electron beam- irradiated lotus (*Nelumbo nucifera*) seeds. *Food Chem.* **107**, 174-184.
- Blummel M., Steingass H. and Becker K. (1997). The relationship between *in vitro* gas production *in vitro* microbial biomass and N-15 incorporation and its implications for the prediction of voluntary feed intake of roughages. *Br. J. Nutr.* **77**, 911-921.
- Charlesby A. (1981). Crosslinking and degradation of polymers. *Radiat. Phys. Chem.* **18**, 59-66.
- Chen K.H., Huber J.T., Simas J., Theurer C.B., Yu P. and Chan S.C. (1995). Effect of enzyme treatment or steam flaking of sorghum grain on lactation and digestion in dairy cows. J. Dairy Sci. 78, 1721-1727.

- Chesson A., Stewart C.S. and Wallace R.J. (1982). Influence of plant phenolic acids on growth and cellulolytic activity of rumen bacteria. *Appl. Environ. Microbiol.* 44, 597-603.
- De Toledo T.C.F., Canniatti-Brazaca S.G., Arthur V. and Piedade S.M.S. (2007). Effects of gamma radiation on total phenolics, trypsin and tannin inhibitors in soybean grains. *Radiat. Phys. Chem.* 76, 1653-1656.
- Dela Rosa A.M., Dela Mines A.S., Banzon R.B. and Simbul Nuguid Z.F. (1983). Radiation pretreatment of cellulose for energy production. *Radiat. Phys. Chem.* 22, 861-868.
- Duodu K.G., Minnaar A. and Taylor J.R.N. (1999). Effect of cooking and irradiation on the labile vitamins and antinutrient content of a traditional African sorghum porridge and spinach relish. *Food Chem.* 66, 21-27.
- Ebrahimi S.R., Nikkhah A., Sadeghi A.A. and Raisali G. (2009). Chemical composition secondary compounds ruminal degradation and *in vitro* crude protein digestibility of gamma irradiated canola seed. *Anim. Feed Sci. Technol.* **151**, 184-193.
- Ebrahimi-Mahmoudabad S.R. and Taghinejad-Roudbaneh M. (2011). Investigation of electron beam irradiation effects on anti-nutritional factors, chemical composition and digestion kinetics of whole cottonseed, soybean and canola seeds. *Radi Phys. Chem.* **80**, 1441-1447.
- El-Niely H.F.G. (2007). Effect of radiation processing on antinutrients, *in vitro* protein digestibility and protein efficiency ratio bioassay of legume seeds. *Radiat. Phys. Chem.* **76**, 1050-1057.
- Farag M.D.E.H. (1998). The nutritive value for chicks of full-fat soybeans irradiated at up to 60 kGy. *Anim. Feed Sci. Technol.* 73, 319-328.
- Feizi R., Ghodratnama A., Zahedifar M., Danesh Mesgaran M. and Raisianzadeh M. (2005). Apparent digestibility of pomegranate seed fed to sheep. P. 222 in Proc. Br. Soc. Anim. Sci. Penicuik, England.
- Galyean M.L. (1997). Laboratory Procedure in Animal Nutrition Research. Texas A and M Research and Extension Center, Amarillo. USA.
- Ghanbari F., Ghoorchi T., Shawrang P., Mansouri H. and Torbati-Nejad N.M. (2012). Comparison of electron beam and gamma ray irradiations effects on ruminal crude protein and amino acid degradation kinetics and *in vitro* digestibility of cottonseed meal. *Radi. Phys. Chem.* 81, 672-678.
- Guimaraes-Beelen P.M., Berchielli T.T., Beelen R., Araujo Filho J. and Oliveira S.G. (2006). Characterization of condensed tannins from native legumes of the Brazilian northeastern semi-arid. *Sci. Agric.* 63, 522-528.
- Holm N.W. and Berry R.J. (1970). Manual on Radiation Dosimetry. Dekker, New York.
- Jackson F.S., Barry T.N., Lascano C. and Palmer B. (1996). The extractable and bound condensed tannin content of leaves

from tropical tree. J. Sci. Food Agric. 71, 103-110.

Kamalak A., Canbolat O., Gurbuz Y., Ozay O., Ozkan C.O. and Sakarya M. (2004). Chemical composition and *in vitro* gas production characteristics of several tannin containing tree leaves. *Livest. Res. Rural Dev.* Available at: http://www.lrrd.org.

- Khan F., Ahmad S.R. and Kronfli E. (2006). c-Radiation induced changes in the physical and chemical properties of lignocellulose. *Biomacromolecules*. 7, 2303-2309.
- Khosravi F., Fathi Nasri M.H., Modaresi S.J. and Fazaeli Rad A. (2012). Effect of electron beam irradiation on phenolic compound content and rumen degradability parameters of pomegranate seed pulp. Pp 2491-2495 in Proc. Nation. Cong. Ani. Poult. Sci. North. Country. Sari, Iran.
- Larbi A., Smith J.W., Kurdi I.O., Adekunle I.O., Raji A.M. and Lapido D.O. (1998). Chemical composition, rumen degradation and gas production characteristics of some multipurpose fodder tree shrubs during wet and dry seasons in the humid tropics. *Anim. Feed Sci. Technol.* **72**, 81-96.
- Makkar H.P.S. (2004). Recent advances in the *in vitro* gas method for evaluation of nutritional quality of feed resources. Pp. 55-88 in Assessing Quality and Safety of Animal Feeds. FAO Animal Production and Health Series. Rome.
- Makkar H.P.S., Blummel M. and Becker K. (1998). Application of an *in vitro* gas method to understand the effects of natural plant products on availability and partitioning of nutrients. Pp. 147-150 in *In vitro* Techniques for Measuring Nutrient Supply to Ruminants. E.R. Devaille, E. Owen, A.T. Adesogan, C. Rymer, J.A. Huntington and T.L.J. Lawrence, Eds. Edinburgh, Scotland.
- McDougall E.I. (1948). Studies on ruminant saliva. I. The composition and output of sheep's saliva. *Biochem. J.* **43**, 99-109.
- Mirzaei-Aghsaghali A., Maheri-Sis N., Mansouri H., Razeghi M.E., Mirza-Aghazadeh A., Cheraghi H. and Aghajanzadeh-Golshani A. (2011). Evaluating potential nutritive value of pomegranate processing by-products for ruminants using *in vitro* gas production technique. *ARPN J. Agric. Biol. Sci.* 6, 45-51.
- Modarresi S.J., Fathi Nasri M.H., Dayani O. and Rashidi L. (2010). The effect of pomegranate seed pulp feeding on DMI, performance and blood metabolites of southern Khorasan crossbred goats. *Anim. Sci. Res.* **20**, 123-132.
- Mohmmadabadi T., Chaji M. and Tabatabaei S. (2010). The effect of tannic acid on *in vitro* gas production and rumen fermentation of sunflower meal. *J. Anim. Vet. Adv.* **9**, 277-280.
- Ndagurwa H.G.T. and Dube J.S. (2013). Nutritive value and digestibility of mistletoes and woody species browsed by goats in a semi-arid savanna, southwest Zimbabwe. *Livest. Sci.* **151**, 163-170.
- Ndlovu L.R. and Nherera F.V. (1997). Chemical composition and relationship to *in vitro* gas production of Zimbabwean browsable indigenous tree species. *Anim. Feed Sci. Technol.* 69, 121-129.
- Nsahlai I.V., Siaw D.E.K.A. and Osuji P.O. (1994). The relationship between gas production and chemical composition of 23 browses of the genus Sesbania. *J. Sci. Food Agric.* **65**, 13-20.
- O'Donovan L. and Brooker J.D. (2001). Effect of hydrolysable and condensed tannins on growth morphology and metabolism of (*Streptococcus gallolyticus*), (*Streptococcus caprinus*) and (*Streptococcus bovis*). *Microbiology*. **147**, 1025-1033.
- Orskov E.R. and McDonald I. (1979). The estimation of protein

degradability in the rumen from incubation measurements weighted according to the rate of passage. J. Agric. Sci. 92, 499-503.

- Parker M.L., Grant A., Rigby N.M., Belton P.S. and Taylor J.R.N. (1999). Effect of popping on the endosperm cell walls of sorghum and maize. J. Cereal. Sci. 30, 209-216.
- Pekel N., Yoshii F., Kume T. and Guven O. (2004). Radiation crosslinking of biodegradable hydroxypropyl methylcellulose. *Carbohydr. Polym.* 55, 139-147.
- Polvi J. and Nordlund K. (2014). Low-energy irradiation effects in cellulose. J. Appl. Physiol. 115, 1-8.
- Prakash C.V.S. and Prakash I. (2011). Bioactive chemical constituents from pomegranate (*Punica granatum*) juice, seed and peel: a review. *Int. J. Res. Chem. Environ.* 1, 1-18.
- Riley R.A. (1994). Free radicals in biology: oxidative stress and the effects of ionizing radiation. *Int. J. Radi. Biol.* **65**, 27-33.
- SAS Institute. (2002). SAS[®]/STAT Software, Release 9.1. SAS Institute, Inc., Cary, NC. USA.
- Shabtay A., Eitam H., Tadmor Y., Orlov A., Meir A., Weinberg P., Weinberg Z.G., Chen Y., Brosh A., Izhaki I. and Kerem Z. (2008). Nutritive and antioxidative potential of fresh and stored pomegranate industrial byproduct as a novel beef cattle feed. J. Agric. Food Chem. 56, 10063-10070.
- Shawrang P., Sadeghi A.A., Behgar M., Zareshahi H. and Shahhoseini G. (2011). Study of chemical compositions antinutritional contents and digestibility of electron beam irradiated sorghum grains. *Food Chem.* **125**, 376-379.
- Taghinejad M., Nikkhah A., Sadeghi A.A., Raisal G. and Chamani M. (2009). Effects of gamma irradiation on chemical composition, antinutritional factors, ruminal degradation and *in vitro* protein digestibility of full-fat soybean. *Asian-Australas J. Anim. Sci.* 22, 534-541.
- Tahan G.H., Fathi Nasri M.H., Riasi A., Behgar M. and Farhang far H. (2012). Effect of electron radiation on degradation characteristics, ruminal and post ruminal DM and CP digestion of some plant protein resources. *Iranian J. Anim. Sci. Res.* 3, 422-434.
- Taher-Maddah M., Maheri-Sis N., Salamatdoustnobar S. and Ahmadzadeh A. (2012). Estimating fermentation characteristics and nutritive value of ensiled and dried pomegranate seeds for ruminants using *in vitro* gas production technique. *Open Vet. J.* 2, 40-45.
- Theodorou M.K., Williams B.A., Dhanoa M.S., McAllan A.B. and France J. (1994). A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. *Anim. Feed Sci. Technol.* 48, 185-197.
- Tilly J.M.A. and Terry R.A. (1963). A two stage technique for the *in vitro* digestion of forage crops. *J. Br. Grassl. Soc.* **18**, 104-111.
- Topuz A. and Ozdemir F. (2004). Influences of gamma irradiation and storage on the capsaicinoids of sun-dried and dehydrated paprika. *Food.* **86**, 509-515.
- Van Soest P.J., Robertson J.B. and Lewis B.A. (1991). Methods for dietary fiber, neutral detergent fiber and nonstarch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* 74, 3583-597.

Variyar P.S., Bandyopadhyay C. and Thomas P. (1998). Effect of gamma-irradiation on the phenolic acids of some Indian spices. *Int. J. Food Sci. Technol.* 33, 533-537.